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Effect of paclobutrazol and daminozide on vegetative attributes of chrysanthemum (*Dendranthema grandiflora* Tzvelve)

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Abstract

The present investigation was conducted to evaluate the vegetative parameters of chrysanthemum cv. Dolly White in response to different concentrations of paclobutrazol and daminozide at Floriculture and Landscaping Block, College of Horticulture, VCSG Uttarakhand University of Horticulture and Forestry, Bharsar, Pauri Garwal from June to December, 2019. The experiment was laid out on Randomized Complete Block Design with nine treatments. The treatments used were control, paclobutrazol (30, 60, 90 and 120 ppm) and daminozide (250, 500, 750 and 500 ppm). Each treatment was replicated thrice. The cuttings were planted in pots of size 15 cm × 15 cm (length and diameter). Paclobutrazol and daminozide were applied at 30 and 60 days after planting of cuttings.

The results of investigation revealed that shortest plant height (18.22 ± 0.35 cm) was recorded from the plant drenched with paclobutrazol @ 120 ppm and found statistically at par with paclobutrazol @ 60 and 90 ppm (19.82 ± 0.64 cm and 18.69 ± 0.57 cm, respectively). The diameter of main stem and plant spread (5.02 ± 0.05 mm and 21.33 ± 0.24 cm, respectively) were recorded maximum in plants sprayed with daminozide @ 1000 ppm and found statistically at par with paclobutrazol @ 90 ppm (4.93 ± 0.01 mm and 20.32 ± 0.38 cm, respectively).

Thus, it can be concluded that drenching of paclobutrazol @ 60 ppm was found effective in getting dwarf plant. For improving maximum plant spread paclobutrazol @ 90 ppm was found best.

Keywords: paclobutrazol, daminozide, dolly white

Introduction

Chrysanthemum is one of the most demanded flower crop in domestic and international markets. The wide range of flower size, colour, shape and form of the flower grab attention of consumers. The flowers are used as both cut as well as loose flower. Beside this the dwarf and compact growth plants are suitable for pot and bedding purpose. Nowadays due to continuous increase in population and urbanization there is shortage of land for cultivation of plants. Therefore, peoples living in urban areas shifting their interest from growing tall and bushy plants to compact and dwarf type one. The small sized plants help in greater space utilization efficiency. Ahmad *et al.* (2015) [1] reported that production of small and compact plants offer an economic benefit to producer. The use of plant growth retardants is an effective technique to restrict stem elongation to achieve aesthetically and commercially pleasing plants (Rezazadeh *et al.*, 2016) [20]. The growth retardants are very useful for manipulating the shape, size and form of plants.

It is herbaceous, perennial plant. The leaves are spirally arranged on erect, branched stems and the flowers are borne in capitula. The distinguishing character of this family is that a large number of tiny flowers are closely mounted on a flattened stem end which given the whole compositae inflorescence a false appearance of a single bloom. The flowering time of the most of cultivars is controlled by modifying the natural day length (Bhattacharjee, 2003) [3]. The national chrysanthemum societies in each country classify the cultivars grown on the basis of bloom characteristics *viz.*, the relative number of two kinds of florets, the physical shape of florets and the direction of growth and arrangement of florets. The chrysanthemum bloom contains two types of florets *i.e.* disc floret and ray floret. Disc floret is situated in centre, tubular in shape with five united petals whereas ray floret surrounds the disc florets and comparatively longer in shape, colour and direction of growth (Kher, 1988) [13].

Materials and Methods

A field experiment was conducted at College of Horticulture, VCSG, UHF, Bharsar, Pauri Garhwal, Uttarakhand, India during June- December, 2019. The experiment consist of ten treatments *viz.*, T₁ Control (Water spray); T₂ Paclobutrazol @ 30 ppm (Soil drenching); T₃

Paclobutrazol @ 60 ppm (Soil drenching); T₄ Paclobutrazol @ 90 ppm (Soil drenching); T₅ Paclobutrazol @ 120 ppm (Soil drenching); T₆ Daminozide @ 250 ppm (Foliar spray); T₇ Daminozide @ 500 ppm (Foliar spray); T₈ Daminozide @ 750 ppm (Foliar spray); T₉ Daminozide @ 1000 ppm (Foliar spray); The treatments were replicated thrice in Randomized Complete Block Design. The farmyard manure and vermicompost were applied as per treatment allocation to the plots uniformly. Inorganic fertilizers were applied in the form of calcium ammonium nitrate, single super phosphate and muriate of potash. Half dose of nitrogen and full dose of phosphorous and potash were applied as a basal dose and remaining half dose of nitrogen was applied 30 days after transplanting. The terminal cuttings of chrysanthemum for experimental purpose were taken from a healthy stock plants maintained at Floriculture and Landscaping Block, Bharsar, Uttarakhand in the month of June. The length of cutting was about 5-7 cm. The cuttings were prepared by removing basal leaves and reducing the leaf area of remaining leaves to half. The basal portion of cuttings was dipped in IBA solution @ 1000 ppm and planted in a raised nursery bed in shady conditions and then transplanted in plastic pots (length 15 cm and diameter 15 cm) under shade condition. The pots were filled with growing media containing sand + soil + FYM + Vermicompost (1:1:1:1). In each pot a single cutting was planted at the centre. Paclobutrazol and Daminozide were applied 30 and 60 days after transplanting of cuttings. They were applied in early morning hours.

Results and Discussion

The shortest plant height (18.21 ± 0.35 cm) was recorded from the plants drenched with T₅ (paclobutrazol @ 120 ppm) and found statistically at par with T₄ (18.69 ± 0.57 cm) and T₃ (19.82 ± 0.64 cm). Similarly, minimum internodal length (2.92 ± 0.22 cm) was observed in T₄ (paclobutrazol @ 90 ppm) and found statistically at par with T₅ (2.96 ± 0.09 cm) and T₃ (3.22 ± 0.08 cm). The results also revealed that all the applied treatments significantly reduced plant height and internodal length as compared to control. The reduction in plant height and internodal length by the application of different concentrations of paclobutrazol might be due to their inhibitory role on cell division and cell elongation of apical meristematic cells. Similar findings have been reported by Karlovic *et al.* (2004)^[10] and Dorajeero *et al.* (2012)^[5] in chrysanthemum. Rademacher (2000)^[19] reported that paclobutrazol has an inhibitory effect on gibberellin biosynthesis, which block cytochrome P-450 dependent monooxygenases there by leads to inhibition oxidation of ent-kaurene in to ent-kaurenoic acid. Thus, blocks the biosynthesis of the active gibberellin which results to reduction in internodal length and plant height (Mahgoub *et al.*, 2006)^[15]. Similarly, suppression of plant height and internodal length under different concentrations of daminozide might be due to it reduce the conversion of GA to active forms of GA and reducing translocation of GA to actively growing tissues. The above result is corroborated with the finding of Singh (2016)^[23] in chrysanthemum. The suppression in plant height under paclobutrazol and daminozide could be attributed due to less internodal elongation registered under these treatments. The results also revealed that increase in concentration of these chemicals there was significant reduction in plant height when compared to control. The results are in close conformity with the

findings of Pinto *et al.* (2005)^[18] in zinnia and Dorajeero *et al.* (2012)^[5] in chrysanthemum. Both the growth retardants reduce the level of endogenous GA₃, that ultimately leads to the shortening of internodes. Similar findings of shortening of internodal length were also reported by Moond and Rakesh (2006)^[16] in chrysanthemum crop, Zalewaska (1989)^[25] in chrysanthemum cultivars and Gad *et al.* (1997)^[7] in *Fuchsia magellanica*.

The maximum diameter of main stem (5.02 ± 0.05 mm) was noticed in treatment T₉ (Daminozide @ 1000 ppm) and found statistically at par with T₄ (4.93 ± 0.01 mm). In case of plant spread, maximum spread (21.33 ± 0.24 cm) was observed in T₉ (daminozide @ 1000 ppm) and found statistically at par T₄ (20.32 ± 0.38 cm). Application of plant growth retardant play a vital role in anti-auxin activity, disturb carbohydrate metabolism, inhibition of cell division and elongation of apical meristem (Jagdale *et al.*, 2017)^[9]. This might be due to the action of growth retardants suppress the formation of apical buds that causes reduction in terminal growth. Therefore, more auxin would have been available for the lateral buds of the plants which ultimately results in improving stem diameter and increasing plant spread. Similar findings were reported by Kim *et al.* (2010)^[12] in chrysanthemum, El-Sheibany *et al.* (2007)^[6] in local cultivar of chrysanthemum, Singh (2016)^[23] in chrysanthemum and Kumar *et al.* (2007)^[14] in marigold, Sen and Maharana (1971)^[22] in chrysanthemum and Gowda and Jayanhi (1991)^[8] in marigold.

The minimum leaf area (11.74 ± 0.45 cm²) was recorded in T₅ and found statistically at par with T₂ (12.74 ± 0.42 cm²), T₃ (12.56 ± 0.59 cm²), and T₄ (12.00 ± 0.32 cm²). It might be due to because it blocks the GA₃ biosynthesis which causes reduction in speed of cell division and extension so that leaf area was retard. These findings are in conformity with Dani *et al.*, (2010)^[4] in marigold and Saiyad *et al.* (2010)^[21] in gaillardia and Nasrullah *et al.* (2012)^[17] in bougainvillea. Tanimoto (1987)^[24] reported that paclobutrazol has the greatest effects on expanding or elongating cells, where inhibition of gibberellin synthesis rapidly causes reductions in stem elongation and leaf expansion so the leaf area was found minimum in paclobutrazol. The maximum leaf area (15.15 ± 0.39 cm²) was recorded in T₉ (daminozide @ 1000 ppm) and found statistically at par with T₁ (14.05 ± 0.32 cm²), T₆ (14.12 ± 0.75 cm²), T₇ (14.48 ± 0.19 cm²) and T₈ (14.73 ± 0.15 cm²). The non significant effect of daminozide application on leaf area have been reported by Pinto *et al.* (2005)^[18] in zinnia and Bhat *et al.* (2011)^[2] in *Erysimum marshallii*.

Conclusion

The results indicated that all the applied concentration of paclobutrazol and daminozide was found significantly effective in reducing plant height as compared to control. The shortest plant height was recorded in plants drenched with paclobutrazol @ 120 ppm and found statistically at par with paclobutrazol @ 90 ppm and @ 60 ppm. The plants spread was found maximum in plants sprayed with daminozide @ 1000 ppm and found statistically at par with daminozide @ 750 ppm and paclobutrazol @ 90 ppm.

Therefore, from the above findings it can be concluded that drenching of paclobutrazol @ 60 ppm was found effective in getting dwarf plant of chrysanthemum cv. Dolly White. To get maximum plant spread drenching of paclobutrazol @ 90 ppm can be recommended.

Table 1: Effect of different concentrations of paclobutrazol and daminozide on vegetative attributes of chrysanthemum cv. Dolly White

Treatments	Plant height (cm) ± S.E(m)	Internodal length (cm) ± S.E(m)	Diameter of main stem (mm) ± S.E(m)	Plant spread (cm) ± S.E(m)	Leaf area (cm ²) ± S.E(m)
T ₁ (Control)	25.77 ± 0.59	3.87 ± 0.02	4.50 ± 0.07	15.40 ± 0.31	14.05 ± 0.32
T ₂ (Paclobutrazol @ 30 ppm)	20.63* ± 0.52	3.32* ± 0.08	4.67* ± 0.05	16.66* ± 0.48	12.74* ± 0.42
T ₃ (Paclobutrazol @ 60 ppm)	19.82* ± 0.64	3.22* ± 0.08	4.84* ± 0.02	18.93* ± 0.29	12.56* ± 0.59
T ₄ (Paclobutrazol @ 90 ppm)	18.69* ± 0.57	2.92* ± 0.22	4.93* ± 0.01	20.32* ± 0.38	12.00* ± 0.32
T ₅ (Paclobutrazol @ 120 ppm)	18.21* ± 0.35	2.96* ± 0.09	4.74* ± 0.04	16.76* ± 0.45	11.74* ± 0.45
T ₆ (Daminozide @ 250 ppm)	22.85* ± 0.46	3.53* ± 0.05	4.63* ± 0.02	17.04* ± 0.46	14.12 ± 0.75
T ₇ (Daminozide @ 500 ppm)	22.65* ± 0.56	3.47* ± 0.13	4.79* ± 0.03	18.23* ± 0.40	14.48 ± 0.19
T ₈ (Daminozide @ 750 ppm)	21.33* ± 0.52	3.42* ± 0.07	4.87* ± 0.02	20.20* ± 0.33	14.73 ± 0.15
T ₉ (Daminozide @ 1000 ppm)	20.66* ± 0.52	3.35* ± 0.12	5.02* ± 0.05	21.33* ± 0.24	15.15 ± 0.39
S.E.(d)	0.77	0.14	0.05	0.52	0.54
C.D. (0.05)	1.66	0.31	0.10	1.12	1.16

*significant at 0.05 level of significance

References

- Ahmad I, Whipker BE, Dole JM. Paclobutrazol or ancymidol effects on postharvest performance of potted ornamental plants and plugs. Hort Science 2015;50(9):1370-1374.
- Bhat MA, Tahir I, Islam WSST. Effect of cycocel and B-Nine (growth retardants) on growth and flowering of *Erysimum marshallii* (Henfr.). Bois. Journal of Plant Sciences 2011;6(2):95-101.
- Bhattacharjee SK. Advances in ornamental horticulture. Pointer Publishers, Jaipur, India 2003, 35.
- Dani KN, Patil SJ, Patel RJ, Patel NA. Effect of growth retardants on flowering and yield of African marigold (*Tagetes erecta* L.) cv. Double Orange under South Gujarat conditions. The Asian Journal of Horticulture 2010;5:287-290.
- Dorajeero AVD, Mokashi AN, Patil VS, Venugopal CK, Lingaraji S, Koti RV. Effect of foliar application of growth regulators on growth, yield and economics in garland chrysanthemum (*Chrysanthemum coronarium* L.). Karnataka Journal of Agricultural Science 2012;25(3):409-413.
- El-Sheibany OM, Malki NA, Barras-Ali A. Effect of application of growth retardant ALAR on some foliage characters of local cultivar of chrysanthemum. Journal of Science and Its Applications 2007;1(2):15-20.
- Gad M, Schmidt G, Gerzson L. Comparison of application methods of growth retardants on the growth and flowering of *Fuchsia magellanica* Lam. Horticultural Science 1997;21(2):263-64.
- Gowda NJV, Jayanthi R. Effect of cycocel and maleic hydrazide on growth and flowering of African marigold (*Tagetes erecta* L.). Progressive Horticulture 1991;23:114-118.
- Jagdale AR, Khobragade YR, Panchbhai DM, Ghormade AC, Bhaskarwar AC. Growth and flowering of annual chrysanthemum influenced by cycocel An paclobutrazol. Journal of Soil and Crops 2017;27(1):143-146.
- Karlovic K, Vrsek I, Sindrak Z, Zidovec V. Influence of growth regulators on the height and number of inflorescence shoots in the chrysanthemum cultivar Revert. Agriculturae Conspectus Scientificu 2004;69(2-3):63-66.
- Kamounsis AP, Sereli C. Paclobutrazol affects growth and flower bud production in gardenia under different light regimes. Horticultural Science 1999;34:674-675.
- Kim YH, Khan AL, Hamayun M, Kim JT, Lee JH, Hwang IC *et al.* Effects of prohexadione calcium on growth and gibberellins contents of *Chrysanthemum morifolium* R. cv. Monalisa White. Horticultural Science 2010;23:423-27.
- Kher MA. Training of chrysanthemum, In: India ed, M.A. Kher, Associated publishing company. New Delhi 1988, 26.
- Kumar TS, Balakrishna V, Jawaharlal M, Ganga M. Response of micro-nutrients on flowering, yield and Xanthophyll content in African Marigold (*Tagetes erecta* Linn.) Journal of Ornamental Horticulture 2007;10(3):153-156.
- Mahgoub MH, Aziz NJGE, Youssef AA. Influence of foliar spray with paclobutrazol or glutathione on growth, flowering and chemical composition of *Calendula officinalis* L. Plant. Journal of Applied Sciences Research 2006;2(11):879-883.
- Moond SK, Rakesh. Effect of GA₃, CCC and MH on vegetative growth and yield of chrysanthemum. Horticultural Science 2006;35:258-59.
- Nasrullah N, Wati YM, Utami DW. Induction of Bougainvillea flower (*Bougainvillea spectabilis* Wild) with retardant and medium composition at polluted environment. Jurnal Lanskap Indonesia 2012;4:59-65.
- Pinto ACR, Rodrigues TDJD, Leite IC, Barbosa JC. Growth retardants on development and ornamental quality of potted Lilliput *Zinnia elegans* Jacq. Science Agriculture 2005, 337-345.
- Rademacher W. Growth retardants: Effects on gibberellin biosynthesis and other metabolic pathways. Annual Review of Plant Biology 2000;51:501-531.
- Rezazadah A, Harkess RL, Bi G. Effect of plant growth regulators on growth and flowering of potted red firespike. Horticulture Technology 2016;26(1):6-11.
- Saiyad MY, Jadav RG, Parmar AB, Chauhan KM. Effect of plant growth retardants and pinching on growth, flowering and yield of gaillardia (*Gaillardia pulchella* Foug.) cv. Lorenziana. The Asian Journal of Horticulture 2010;5(1):121-122.
- Sen SK, Maharana T. Growth and flowering response of chrysanthemum to growth regulator treatments. The Punjab Horticultural Journal 1971;11(3/4):274-277.
- Singh N. Effect of growth retardants on growth and flowering of chrysanthemum (*Chrysanthemum morifolium* Ramat.). M.Sc. thesis submitted to PAU, Ludhiana 2016.
- Tanimoto E. Gibberellin-dependent root elongation in *Lactuca sativa*: Recovery from growth retardant-suppressed elongation with thickening by low concentration of GA₃. Plant and Cell Physiology 1987;28:963-973.
- Zalewska M. Growth regulators in pot culture of chrysanthemum cultivars Palom and Promyk. Acta Horticulturae 1989;251:335-40.