

E-ISSN: 2278-4136 P-ISSN: 2349-8234 www.phytojournal.com JPP 2020; 9(6): 276-279 Received: 15-09-2020 Accepted: 19-10-2020

Himanshu Rai

Department of Seed Science and Technology, CCS Haryana Agriculture University, Hisar, Haryana, India

Ramesh Nath Gupta

Department of Plant Pathology, Bihar Agricultural University, Sabour, Bihar, India

Shyam Babu Sah Department of Entomology, Bihar Agricultural University, Sabour, Bihar, India

OS Dahiya

Department of Seed Science and Technology, CCS Haryana Agriculture University, Hisar, Haryana, India

Corresponding Author: Ramesh Nath Gupta Department of Plant Pathology, Bihar Agricultural University, Sabour, Bihar, India

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



Journal

Seed characters based identification key for important varieties of Indian mustard (*Brassica juncea* L. Czern and Coss)

Himanshu Rai, Ramesh Nath Gupta, Shyam Babu Sah and OS Dahiya

Abstract

Indian mustard (*Brassica juncea* L. Czern & Coss) is an important economical oilseed crop worldwide. An experiment was carried out at the Department of Seed Science & Technology, CCS Haryana Agriculture University, Hisar, Haryana to prepare an identification key for mustard varieties. It was based on seed characters of twenty Indian mustard varieties release for general cultivation from the various organizations. The characterization of varieties was based on the morphological characters as well as response of chemical seed tests. Mustard varieties were distinguished from each other on the basis of seed colour, test weight, oil content, phenol test, modified phenol test, potassium hydroxide test, peroxidase test and 2, 4-D auxin test. These seed characters based identification key can be an effective tool for maintaining the genetic purity of the varieties during seed certification process.

Keywords: Indian mustard, seed, variety, identification, characterization

Introduction

Indian mustard (Brassica juncea L. Czern & Coss) belonging to family Brassicaceae is an important economical oilseed crop, grown across the world for its rich source of oil and potential meal. In India, it is mainly cultivated in the states of Assam, Bihar, Gujarat, Haryana, Himachal Pradesh, Jammu and Kashmir, Madhya Pradesh, Odisha, Punjab, Rajasthan, Uttar Pradesh and West Bengal during Rabi season. At present large number of Indian mustard varieties are available for commercial cultivation in our country. The present trend of continuous release of new Indian mustard varieties from central and state varietal release committees has warranted to developed suitable techniques for varietal identification at the Seed Testing Laboratory level particularly when the seeds are submitted for the purity testing. Maintenance of genetic purity of varieties is of primary importance for preventing varietal deterioration during successive regeneration cycles and for ensuring varietal performance at the expected level. The purity of seed material can be maintained by combining the morphological and chemical test based characters of the seeds. The seed characteristics and chemical tests reveal differences among the varieties as reported by various workers ^[1-4]. The results of these tests are usually distinct and can be easily interpreted for identification of the varieties. Therefore, an investigation was carried out to know the various seed characters and chemical test based characters of Indian mustard varieties for effective utilization in varietal characterization during purity testing.

Materials and Methods

The experiment was conducted in the Seed Testing Laboratory of the Department of Seed Science & Technology, CCS Haryana Agriculture University, Hisar, (Haryana) during 2014 to 2016. The investigation materials comprised of twenty Indian mustard varieties *viz.*, RH30, RH8812, RH8113, RH0749, RB50, RH0406, RB24, RH0119, RH9304, RH9801, RH819, RH781, Varuna, NRCDR02, NRCDR601, NRCHB101, DRMRIJ31, NPJ112, RGN73 and Kranti. These varieties were released from State and Central Variety Release Committees for commercial cultivation of the Indian mustard. The characters observed during the investigation were seed colour, test weight, oil content and chemical tests *viz.*, phenol test, modified phenol test, potassium hydroxide test, Peroxidase activity test and 2, 4-D auxin test. The procedures for recording observations were considered as under:

Seed colour

Visual group observations were recorded on the seed lots and grouped into Dark Brown, Brown and Reddish Brown.

Seed size (Test weight)

Based on thousand seed weight (in gm), groups were made as Small, Medium and Bold.

Oil content

The oil content of each seed lot was estimated by using Nuclear Magnetic Resonance (NMR) MK111 Newport Analyser equipped with 2.0 ml sample viol assembly. The varieties were grouped into Low and Medium oil content groups.

Phenol test

Soaking of the seeds in water for 16 hours under ambient conditions followed by transferring 50 seeds in 15 cm petridishes between two layers of filter paper soaked in 1% phenol solution. Care was taken to keep the hilum region on the lower side. The final observation on the colour was marked as Dark Brown, Dark Red and Dark Grey ^[5].

Modified phenol test

Seeds were soaked for 4 hr in 0.40% solution of CuSO4 for adding Cu++ ions and another set in 0.60% Na2CO3 for adding Na+ ions. Then the seeds were overnight placed in 2.0% phenol solution for colour development. Based on the colour they were marked as Dark Brown, Brown, Reddish Brown and Strong Brown^[6].

Potassium hydroxide test

Seeds were soaked in 5.0% KOH solutions for 2 hr. at room temperature and marked for colour development into three groups viz. Dark Brown, Brown and Light Brown^[7].

Peroxidase activity test

Seeds were soaked in water for 24 hours followed by incubating them in 0.05% of Guaiacol solution for 20 minutes and then in 0.10% H2O2. The Reddish Brown coloured was quantified by D-64 spectrophotometer at 480 nm ^[8].

2, 4-D auxin test

Seeds were grown by placing them on two layers of filter paper moistened in 5.0 ppm solution of 2,4-D auxin in the petridishes at 25 0 C. Seedlings were evaluated after 7 days and 10 seedlings were selected randomly for measuring length (in cm).

Results and Discussions

Varietal identification at the time of seed testing stage requires precise tests conducted in a series to obtain conclusive results. The results obtained in the series of tests conducted on the seeds of different varieties can be used to make 'identification key' for the concerned varieties, which is quick, easy, and reproducible ^[7, 9]. Distinctness means the variety should be clearly distinct from existing varieties. On the basis of seed size, all the genotypes were categorized into three groups, i.e. small, intermediate and bold type ^[11]. Uniformity indicates the similarity of characteristics features among the plants of selected variety. Stability means that the variety should produce stable results year after year. Seventy eight genotypes of mustard were grouped by using DUS test traits ^[10]. The seeds of the twenty Indian mustard varieties were subjected the series of tests under laboratory conditions to obtain the results as presented in table 1.

| Table 1: Characterization of India | an mustard varieties based on seed characters | 5 |
|------------------------------------|---|---|
|------------------------------------|---|---|

| Variety | Seed colour | Seed size (Test weight in gm) | Oil content (%) | Phenol test | Modified phenol test | KOH test | Peroxidase activity test | 2,4-D auxin test |
|----------|---------------|----------------------------------|-----------------|-------------|-------------------------|-------------|-----------------------------|------------------|
| RH30 | Dark Brown | Medium (5.95) | 39.37 Medium | Dark brown | Brown | Light brown | Low | High susceptible |
| RH8812 | Dark Brown | Medium (5.12) | 38.88 Medium | Dark brown | Dark brown | Light brown | High | Susceptible |
| RH8113 | Dark Brown | Medium (5.29) | 38.81 Medium | Dark brown | Dark brown | Dark brown | Low | Susceptible |
| RH0749 | Dark Brown | Medium (5.41) | 39.08 Medium | Dark brown | Brown | Brown | Medium | Tolerant |
| RB50 | Dark Brown | Medium (5.30) | 38.26 Medium | Dark grey | Brown | Light brown | High | High susceptible |
| RH0406 | Brown | Bold (6.01) | 38.77 Medium | Dark grey | Dark brown | Dark brown | Low | Tolerant |
| RB24 | Dark Brown | Medium (5.74) | 38.22 Medium | Dark red | Brown | Brown | Medium | High susceptible |
| RH0119 | Dark Brown | Medium (5.87) | 38.10 Medium | Dark brown | Reddish brown | Light brown | High | Susceptible |
| RH9304 | Reddish Brown | Medium (5.89) | 38.20 Medium | Dark grey | Brown | Brown | Medium | Susceptible |
| RH9801 | Dark Brown | Medium (5.95) | 38.40 Medium | Dark brown | Reddish brown | Light brown | Medium | High susceptible |
| RH819 | Brown | Medium (5.82) | 38.34 Medium | Dark brown | Strong brown | Dark brown | High | Tolerant |
| RH781 | Dark Brown | Medium (5.73) | 37.69 Low | Dark red | Reddish brown | Light brown | High | Tolerant |
| Varuna | Brown | Medium (5.58) | 38.22 Medium | Dark grey | Brown | Brown | High | High susceptible |
| NRCDR02 | Dark Brown | Medium (5.67) | 38.20 Medium | Dark grey | Dark brown | Dark brown | High | Susceptible |
| NRCDR601 | Dark Brown | Medium (5.84) | 37.92 Low | Dark brown | Strong brown | Dark brown | Medium | High susceptible |
| NRCHB101 | Brown | Small (4.42) | 37.75 Low | Dark grey | Dark brown | Brown | High | High susceptible |
| DRMRIJ31 | Brown | Medium (5.98) | 38.30 Medium | Dark brown | Brown | Light brown | High | Susceptible |
| NPJ112 | Dark Brown | Small (4.84) | 38.02 Medium | Dark brown | Dark brown | Dark brown | Medium | Susceptible |
| RGN73 | Dark Brown | Medium (5.72) | 38.28 Medium | Dark brown | Reddish brown | Light brown | Medium | Susceptible |
| Kranti | Brown | Small (4.77) | 38.28 Medium | Dark brown | Strong brown | Light brown | Medium | High susceptible |

Based on the variation in the seed characters by morphological and chemical treatments based response, an effort was made to categorize the varieties to distinguish them from each other in a systematic way, which ultimately resulted in the Key for Identification. The key is based on the above morphological and laboratory test's based characters in the following sequence:

Seed Colour \rightarrow Seed Size (Test Weight) \rightarrow Oil Content \rightarrow Phenol Test \rightarrow Modified Phenol Test \rightarrow Potassium hydroxide Test \rightarrow Peroxidase Test \rightarrow 2, 4-D auxin Test.

Seed colour Dark brown: RH30, RH8812, RH8113, RH0749, RB50, RB24, RH0119, RH9801, RH781, NRCDR02, NRCDR601, NPJ112, RGN73 Û Seed Size (Test Weight) Small: NPJ112 Medium: RH30, RH8812, RH8113, RH0749, RB50, RB24, RH0119, RH9801, RH781, NRCDR02, NRCDR601, RGN73 Л **Oil Content** Low: RH781, NRCDR601 Phenol Test Û Dark brown: NRCDR601 Dark red: RH781 Ţ **Oil Content** Medium: RH30, RH8812, RH8113, RH0749, RB50, RB24, RH0119, RH9801, NRCDR02, RGN73 Л **Phenol Test** Dark brown: RH30, RH8812, RH8113, RH0749, RH0119, RH9801, RGN73 Ţ Modified Phenol Test Dark brown: RH8812, RH8113 Û **KOH Test** Dark brown: RH8113 Light brown: RH8812 Л **Modified Phenol Test** Brown: RH30, RH0749 T KOH Test Brown: RH0749 Light brown: RH30 Û **Modified Phenol Test** Reddish brown: RH0119, RH9801, RGN73 П **KOH Test** Light brown: RH0119, RH9801, RGN73 T **Peroxidase Test** High: RH Medium: RH9801, RGN73 Û 2, 4-D auxin Test Highly susceptible: RH9801 Susceptible: RGN73 Л **Modified Phenol Test** Û Strong brown: Nil Ţ **Phenol Test** Dark red: RB24 Dark grey: RB50, NRCDR02 Modified Phenol Test Dark brown: NRCDR02 Brown: RB50 Π Seed Size (Test Weight) Bold: Nil T Seed Colour Brown: RH0406, RH819, Varuna, NRCHB101, DRMRIJ31, Kranti Π Seed Size (Test Weight) Small: NRCHB101, Kranti П

```
~ 278 ~
```

| Oil Content | | | | |
|---------------------------------|--|--|--|--|
| Low: NRCHB101 | | | | |
| Medium: Kranti | | | | |
| 1, | | | | |
| Seed Size (Test Weight) | | | | |
| Medium: RH819, Varuna, DRMRIJ31 | | | | |
| \downarrow | | | | |
| Oil content | | | | |
| Low: Nil | | | | |
| Medium: RH819, Varuna, DRMRIJ31 | | | | |
| Π | | | | |
| Phenol Test | | | | |
| Dark brown: RH819, DRMRIJ31 | | | | |
| Ţ | | | | |
| Modified Phenol Test | | | | |
| Dark brown: Nil | | | | |
| Brown: DRMRIJ31 | | | | |
| Reddish brown: Nil | | | | |
| Strong brown: RH819 | | | | |
| \mathbb{T} | | | | |
| Phenol Test | | | | |
| Dark red: Nil | | | | |
| Dark grey: Varuna | | | | |
| \downarrow | | | | |
| Seed Size (Test Weight) | | | | |
| Bold: RH0406 | | | | |
| ↓ | | | | |
| Seed Colour | | | | |
| Reddish brown: RH9304 | | | | |

This Identification Key based on differential response of the seeds of varieties towards different laboratory based tests and observations clearly indicate the possibility of varietal identification of Indian mustard varieties in Laboratory also. The protocol can include many more varieties, as several slots are still vacant in the Key. Further, this Key can provide a potent way to save one full crop season which otherwise will be required in field identification of varieties.

Conclusion

Laboratory based observations based on the morphological and chemical tests on the seeds of the mustard can be used to structure an Identification key for easily characterization the varieties. The present study based on eight characters of the seeds was successfully analysed for identifying twenty varieties of Indian mustard in laboratory.

References

- 1. Agrawal RL, Pawar A. Identification of soybean varieties based on seed and seedling characteristics. Seed Research 1990;18:77-81.
- Chemelar F, Hemelar F, Mostovoj K. On the application of some old and on the introduction of new methods for testing genuineness of variety in the laboratory. Procedurings International Seed Testing Association 1938;10:68-74.
- 3. Joshi MG, Banerjee SK. Genetics of phenol colour reaction in emmer wheat. Proceedings International Seed Association 1970;35:207-211.
- 4. Gupta N, Monika AJ, Navraj KS, Sharma RC. Utility of phenol test in varietal characterization. Crop Improvement 2007;34(1):77-81.
- 5. Jaiswal JP, Agrawal RL. Varietal purity determination in rice modification of the phenol test. Seed Science Technology 1995;23:33-42.

- 6. Banerjee SK, Chandra SS. Modified phenol test for the varietal identification of wheat seed. Seed Science &Technology 1977;5:53-60.
- Agrawal PK. Cultivar purity tests. Techniques in Seed Science and Technology. South Asian Publishers, New Delhi 1987; P160.
- McKee. Chemicals and biochemical techniques for varietal identification. Seed Science Technology 1973;1:181-199.
- 9. Kumar A, Chaudhary RK, Kapoor RL, Dahiya OS. Identification of pearl millet hybrids and their parental lines using seeds and seedling characters, chemical tests and gel electrophoresis. Seed Science Technology 1995;23:21-32.
- 10. Yadav A, Singh D, Arya RK. Morphological characterization of Indian mustard (*B. juncea*) genotypes and their application for DUS testing. Indian Journal of Agricultural Sciences 2013;83:1305-1316.
- 11. Katiyar PK, Dixit GP, Singh BB. Morphological characterization of green gram (*Vigna radiata*) varieties and their application for distinctness, uniformity and stability testing. Indian Journal of Agricultural Sciences 2008;78(5):439-444.