

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234

www.phytojournal.com JPP 2020; 9(5): 1641-1643 Received: 19-07-2020 Accepted: 21-08-2020

Kishan Kumar Sharma

Department of Plant Pathology, College of Agriculture, Raipur, Chhattisgarh, India

AS Kotasthane

Department of Plant Pathology, College of Agriculture, Raipur, Chhattisgarh, India

Rathod Parshuram

Department of Plant Pathology, College of Agriculture, Raipur, Chhattisgarh, India

Corresponding Author: Kishan Kumar Sharma Department of Plant Pathology, College of Agriculture, Raipur, Chhattisgarh, India

$$PGI = \frac{DC-DT}{DC} \times 100$$

~ 1641 ~

In vitro evaluation of potential Trichoderma mutants against collar rot pathogen (Sclerotium rolfsii Sacc.) of chickpea

Kishan Kumar Sharma, AS Kotasthane and Rathod Parshuram

Abstract

Collar rot of chickpea is an important disease which causes severe loss in chickpea production. The present study was carried out to evaluate four potential *Trichoderma* mutants against *Sclerotium rolfsii*. Among the four potential *Trichoderma* mutants tested the maximum inhibition was observed in BARC mutant (81.50%) over control which was followed by mutants M-136 (81%), M-23 (80.5%) and M-18 (79%) respectively.

Keywords: Trichoderma, collar rot pathogen, Sclerotium rolfsii Sacc., chickpea

Introduction

Chickpea (*Cicer arietinum* L.) is an important legume crop and good source of vegetable protein. Chickpea crop can be attacked by several pests and diseases. (Singh *et al.*, 2015) ^[12]. Collar rot of chickpea is one of the economic important and most destructive disease of chickpea caused by Sclerotium rolfsii Sacc. It is prevalent in those areas which have high soil moisture and warm temperature with mortality rate from 10-100 per cent (Kumari and Ghatak, 2018) ^[6]. Seedling mortality due to attack of this pathogen is reported from 54.7 to 95 per cent and yield reduction in field condition is reported from 22 to 50 per cent (Ahsan et al., 2018)^[1]. Due to wide host range and sclerotial formation as a resting structure by the pathogen it survives in the soil for longer period of time which makes management process quite difficult (Wavare *et al.*, 2017). The antagonistic fungus *Trichoderma* spp. have capability to break the outer sclerotial shell that leads to its destruction along with several histological changes such as deformation, decay of cytoplasmic content and cell wall lysis (Rawat and Tewari, 2011)^[11]. Since application of toxic chemicals for management of this disease are hazardous and have residual effect in soil. Use of biological control agents are the best alternative to these toxic chemicals. Among the bio-control agents, Trichoderma viride and Gliocladium virens were found to be most effective against S. rolfsii. Trichoderma spp. has been found as an effective BCA against many seed and soil borne pathogens (Eziashi et al., 2006)^[4]. The mycelial growth inhibition of different pathogens by Trichoderma isolates is due to the production of diffusible compounds, lytic enzymes and water soluble metabolites. (Harmanet al.2004; Yedidia et al.2003)^[16]. Hence, present investigation was carried out to screen out the most compatible combinations of floral extracts, bio-control agents and fungicides to find out efficient management practices against collar rot of chickpea caused by S. rolfsii.

Material and methods

Potential *Trichoderma* mutants and the pathogen were grown on PDA (Potato Dextrose Agar) medium each separately and by using 7 days old cultures, 5 mm diameter disc of the potential mutants and pathogen were taken into consideration. The Petri plates (90 mm) were inoculated aseptically with *S. rolfsii* and potential mutants, by placing 5 mm diameter culture blocks at 70 mm apart from each other. Three repetitions of each treatment were kept and the petri plates with only pathogen served as control. Afterward, the plates were incubated at temperature $(28\pm2^{\circ}C)$ and the radial growth of the test organism and pathogen was measured after 7 days of incubation. The per cent growth inhibition (PGI) was worked out by using the formula given by Vincent (1947)^[13].

Where,

PGI= Per cent growth inhibition

DC = Average diameter of mycelial colony of control set DT = Average diameter of mycelia colony of treated set

Results and discussion

In vitro antagonistic potential of different potential *Trichoderma* mutants against fungal plant pathogen *Sclerotium rolfsii* studied by following dual culture method, and growth assessed 5 days after inoculation. All the potential *Trichoderma* mutants showed varied range of antagonism against *Sclerotium rolfsii* ranging from 79 per cent to 81.50 per cent. Among potential *Trichoderma* mutants, BARC mutant showed maximum inhibiting effect on the growth of *Sclerotium rolfsii* (81.50%) over control. While remaining mutants also showed good inhibitory effect on the growth of *Sclerotium rolfsii* in order M-136 (81%), M-23 (80.5%) and M-18 (79%) respectively.

The antagonistic fungus *Trichoderma* spp. have capability to break the outer sclerotial shell that leads to its destruction along with several histological changes such as deformation, decay of cytoplasmic content and cell wall lysis (Rawat and Tewari, 2011)^[11]. The present study revealed that different *Trichoderma* mutants have capacities as biological weapons in inhibiting the pathogens. This might be due to the production of secondary metabolites and antibiotics production, which diffused into the PDA and air filed spores which showed detrimental effect towards growth of *S. rolfsii* as well as due to higher competitive ability of potential *Trichoderma* mutants. Overall BARC mutant was found to be more efficient against *S. rolfsii* which indicates that it can be exploited as potential candidates for development of biopesticides.

Above findings are in agreement with the observations made by Bhuiyan *et al.*, (2012) ^[3] reported that *T. harzianum* isolate Th-18 showed the highest (83.09%) reduction of the radial growth against *S. rolfsii*. This might be due to the production of secondary metabolites and antibiotics production, which diffused into the PDA and air filed spores which showed detrimental effect towards growth of *S. rolfsii* as well as due to higher competitive ability of potential *Trichoderma* mutants. The antagonistic fungus *Trichoderma* spp.

Bandyopodhyay *et al.* (2003) ^[2] reported that *Trichoderma* strains inhibited the growth of *Sclerotium rolfsii* and *Rhizoctonia solani* by 76.6% and 73.3% respectively. Yadub and Shahzad (2005) ^[15] reported that *T. harzianum* and *T. longibrachiatum* restricted the growth of *S. rolfsii* under *in vitro* condition by coiling around mycelium of *S. rolfsii* which leads in lysis of hyphae. Several workers reported that *Trichoderma viride* as an important antagonist inhibiting the growth of *Sclerotium rolfsii* (Kolte and Raut., 2007; Mandhare and Suryawanshi., 2008) ^[8, 9]. Prajapati *et al.* (2015) ^[10] observed that among different species of *Trichoderma* tested against *S. rolfsii* through dual culture technique, *T. asperellum* showed strong antagonistic effect in terms of mycelia growth inhibition *i.e.* 61.48, 75.00 and 73.33 per cent at 4, 6 and 8 days of incubation, respectively.

Table 1: In vitro evaluation of potential Trichoderma mutantsagainst Sclerotium rolfsii (Dual Culture technique, Dennis and
Webster, 1971)

Treatment	Treatment Name	Per cent Inhibition
T_1	BARC mutant	81.5
T_2	M-18	79
T3	M-23	80.5
T_4	M-136	81
T5	Control	-

*Percent inhibition is calculated 96 hours of incubation

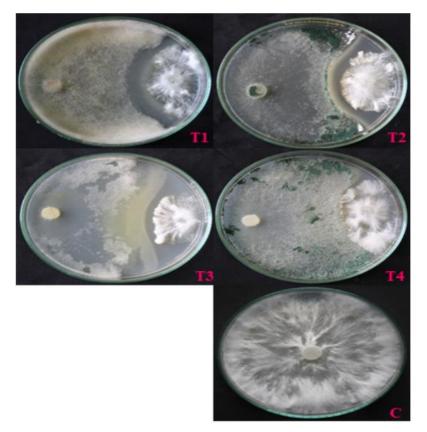


Fig 1: In vitro effect of antagonists against S. rolfsii

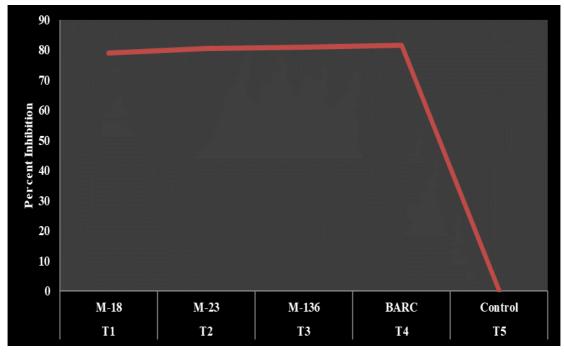


Fig 2: In vitro effect of antagonists against S. rolfsii

References

- 1. Ahsan MS, Kumar M, Upadhya JP. Integrated Approach for the Management of Collar Rot of Chickpea. *Int. J. Curr. Microbiol. App. Sci.* 2018; 7(5):3560-3569.
- 2. Bandyopadhyay S, Sharma ND, Dutta S. Screening of potential *Trichodermai* strains against major root pathogens. *Annals of plant protection sciences*. 2003; 11:163.
- 3. Bhuiyan MA, Rahman MT, Bhuiyan K. *In vitro* screening of fungicides and antagonists against *Sclerotium rolfsii*. African Journal of Biotechnology. 2012; 11(82):14822-14827.
- 4. Eziashi EI, Uma NU, Adekunle AA, Airede CE. Effect of metabolites produced by Trichodermaspeciesagainst Ceratocystis paradoxa in culture medium. *African Journal of Biotechnology*. 2006; 5:703-706.
- 5. Harman GE, Howell CR, Viterbo A, Chet I, Lorito M. *Trichoderma* species-opportunistic, avirulent plant symbionts. *Nat Rev Microbiol.* 2004; 2:43-56.
- Kumari A, Ghatak A. Variability in chickpea rot-causing soil-borne necrotrophs, *Sclerotium rolfsii* and *Macrophomina phaseolina*. Journal of Agri Search. 2018. 5(4):247-253.
- Kulkarni VR. Epidemiology and integrated management of potato wilt caused by Sacc. Ph. D. Thesis, Univ. Agric. Sci., Dharwad, 2007; 191:106.
- 8. Kolte VS, Raut SP. Efficacy of *Trichoderma* spp. against *Sclerotium rolfsii* inciting collar dry rot of orchids. *Annual of Plant Protection Science*. 2007; 15:516-517.
- 9. Mandhare VK, Suryawanshi AV. Efficacy of some botanicals and *Trichoderma* species against soil borne pathogens infecting chickpea. Journal of food legumes, 2008; 21(2):122-124.
- 10. Prajapati BK, Patel JK, Patil RK. Bio-efficacy of *Trichoderma* spp. against *Sclerotium rolfsii* Sacc. An incitant of Collar rot of chickpea *in vitro.The Bioscan*. 2015; 10(4):1745-1748.
- Rawat R, Tewari L. Effect of Abiotic Stress on Phosphate Solublization. *Current Microbiology*. 2011; 62:1521-1526.

- Singh AK, Singh SS, Prakash V, Kumar S, Dwivedi SK. Pulses Production in India: Present Status, Bottleneck and Way Forward. Journal of Agri Search. 2015; 2(2):75-83.
- 13. Vincent JM. Distribution of fungal hyphae in the presence of certain inhibitors. Nature. 1947; 159:850.
- 14. Wavare SH, Gade RM, Shitole AV. Effect of plant extracts, bio agents and fungicides against *Sclerotium rolfsii* causing collar rot in chickpea. Indian Journal of Pharmaceutical Sciences. 2017; 79(4):513-520.
- Yaqub F, Shahzad S. In vitro evaluation of microbial antagonists against Sclerotium rolfsii. Pak. J. Bot. 2005; 37(4):1033-1036.
- 16. Yedidia I, Shoresh M, Kerem Z, Benhamou N, Kapulnik Y, Chet I. Concomitant induction of systemic resistance to *Pseudomonas syringae* pv. *lachrymansin* cucumber by *Trichoderma asperellum* (T-203) and accumulation of phytoalexins. Appl Environ Micro-biol. 2003; 69:7343-7353