



E-ISSN: 2278-4136
P-ISSN: 2349-8234
www.phytojournal.com
JPP 2020; 9(1): 2001-2007
Received: 18-11-2019
Accepted: 20-12-2019

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In vitro evaluation of botanicals against ralstonia solanacearum causing bacterial wilt of potato

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Abstract

Bacterial wilt or brown rot of potato, caused by *Ralstonia solanacearum*, is a limiting factor of potato production worldwide. The management of the disease is difficult, and the chemicals are least effective and hazardous. Hence, it is needed to develop alternative methods to control this deadly disease, which is effective and eco-friendly. Aqueous and ethanolic extracts of eleven plants were evaluated for the antibacterial activity against *Ralstonia solanacearum* in *in-vitro* by well diffusion method. Out of the eleven crude aqueous extracts tested, maximum inhibition zone was observed in bulb extract of *Allium sativum* (30.21mm) followed by *Psidium guajava* (25.11mm), *Curcuma longa* (17.77mm), *Tagetes erecta* (16.21mm), *Carica papaya* (14.55mm) and *Azadirachta indica* (13.77mm). Whereas on evaluation of eleven crude ethanolic extracts, maximum inhibition zone was observed in bulb extract of *Allium sativum* (29.77mm) followed by *Psidium guajava* (22.88mm), *Tagetes erecta* (19.77mm), *Carica papaya* (15.11mm), *Azadirachta indica* (14.88mm), *Ocimum gratissimum* (14.10) and *Curcuma longa* (11.33mm). After screening, four aqueous extracts of botanicals (*Allium sativum*, *Psidium guajava*, *Curcuma longa* and *Tagetes erecta*) and four ethanol extracts of botanicals (*Allium sativum*, *Psidium guajava*, *Tagetes erecta* and *Carica papaya*) which showed inhibition zone ≥ 15 mm were evaluated against *R. solanacearum* at 1%, 5%, 10% and 15% concentration. Among them, garlic showed superiority over all the plants extracts at all the levels of concentration.

Keywords: botanicals, ralstonia, solanacearum, potato

Introduction

Potato (*Solanum tuberosum* L.) is one of the four major and important food crops after wheat, maize and rice around the world (Hawkes, 1992) [10]. It is a rich source of nutrients. Bacterial wilt or brown rot, caused by *Ralstonia solanacearum*, limit potato production throughout the world. It causes huge crop losses in tropical, subtropical, and warm temperate regions (Hayward, 1994; Elphinstone, 2005) [9, 3]. Verma and Shekhawat (1990) [17] reported nearly 37% of potato loss due to brown rot disease in potato in Mukteshwar region of Uttarakhand, 13.8-55% in Kumaun hills and 75% in Karnataka (Gadewar *et al.*, 1991) [4].

The management of bacterial wilt disease caused by *R. solanacearum* is difficult due to its broad host range with the ability to survive in the soil and roots of non-host plants including several weeds. The available conventional bactericides and fumigants such as methyl bromide did not provide effective control of this disease on soil treatment (Chellemi *et al.*, 1997; Enfinger *et al.*, 1979) [1, 2]. By the use of plant metabolites bacterial wilt diseases of potato can be managed which is eco-friendly, non-hazardous and bio-degradable. India is a land of biodiversity in terms of plant species. Various plants have been mentioned in Ayurveda, an ancient Indian Sanskrit literature, for their therapeutic advantages (Kaushik, 1988) [11]. There are many reports by the different researcher that plant metabolites or plant-based pesticides can be used as a good alternative to manage the plant diseases as they have a minimal environmental impact and also non-hazardous in contrast to chemicals (Harborne, 1998; Varma and Dubey, 1999; Gottlieb *et al.*, 2002) [8, 16, 6]. The natural plant products derived from plant species have the capacity to control diseases caused by viruses, bacteria and fungal pathogens (Guleria and Tiku, 2009) [7]. Therefore, under this scenario, botanical extracts seem to be an ideal candidates to be exploited in the management of bacterial wilt of potato because of the safety, renewable nature, cost-effective and high target specificity. The objective of this study was to determine the efficacy of aqueous and ethanolic extract of various plants for controlling *R. solanacearum* causing bacterial wilt of potato under *in vitro* condition.

Materials and Methods

Isolation and purification of *R. solanacearum* strains from wilted potato plants Potato plants showing typical symptoms viz., wilting, stunting and yellowing of plants infected by bacterial

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wilt were collected and diagnosis of the disease was done by ooze test. The pathogen was cultured on TZC (tetrazolium chloride) agar medium and well isolated typical colonies of an isolate of *R. solanacearum* were picked and streaked separately on TZC medium. The pure cultures of the isolates were preserved on CPG slants and kept at 40C for future use.

Sample Collection: Eleven local plants were used in the study. The parts of the plants that were used in this study are the most popular and most readily available. They are shown in Table 1.

Method of Extraction: Aqueous and ethanolic extraction was followed to extract the antimicrobial components contained in eleven different plant species to screen for their antimicrobial property against potato bacterial wilt pathogen.

Protocol for aqueous extract: The crude aqueous extracts of garlic, guava, marigold, turmeric, lemongrass, ginger, onion, neem, tulsi, papaya and datura were prepared according to the method described by Raghu *et al.* 2013^[14].

Protocol for ethanol extract: The crude ethanolic extracts of garlic, guava, marigold, turmeric, lemongrass, ginger, onion, neem, tulsi, papaya and datura were prepared according to the method described by Kumar *et al.* 2017^[12].

In vitro evaluation of botanicals extracts: Both water and ethanolic plant extracts were tested for their antibacterial property by agar well diffusion method (Gopalakrishnan *et al.*, 2014)^[5]. The commercial antibiotic, Streptomycin 500ppm was used as standards for comparison. The well which was loaded with sterile distilled water was used as control. The experiment was performed in triplicate under aseptic conditions. The Petri plates were then placed in the lowermost shelf of a refrigerator (5 °C) for half an hour, thus allowing plant extracts and test chemicals to diffuse into the medium. The plates were then shifted to the incubator at 28 ±10C. The inhibition zone was measured in mm with the help of a scale after 24 h of incubation.

Evaluation of screened aqueous and ethanolic extracts of botanicals against *R. solanacearum* at different concentration. Further four aqueous plant extracts (garlic, guava, turmeric and marigold) and four ethanolic extracts (garlic, guava, marigold and papaya) producing an inhibition zone ≥15mm in diameter were made to different concentrations viz., 1 percent, 5 percent, 10 percent, 15 percent, by adding sterilized distilled water proportionately and screened to determine minimum inhibitory concentration.

Results and Discussion

Screening of crude aqueous extract of different botanicals against *R. solanacearum*. Aqueous extracts of different botanicals were selected to evaluate their antibacterial property against *Ralstonia solanacearum*. Data presented in table 2 showed that out of 11 botanicals extract, 6 botanicals extract (garlic, guava, marigold, turmeric, neem and papaya) exhibited antibacterial property whereas 5 botanicals extract (datura, onion, ginger, tulsi and lemongrass) failed to inhibit the growth of *Ralstonia solanacearum in vitro* (Plate-1). Out of the six botanicals extract, garlic bulb extract was highly inhibitory to the growth of the pathogen showing the highest mean inhibition zone of 30.21mm, followed by guava leaf extract with an inhibition zone of 25.11mm. The next best botanicals extract were turmeric, marigold, papaya and neem

extracts with inhibition zones of 17.77mm, 16.21mm, 14.55 mm and 13.77mm, respectively (Fig.-1).

Several workers reported the inhibitory effects of botanicals against *R. solanacearum in vitro*. Sinha, K. (2016)^[15] demonstrated *in vitro* evaluation of botanicals against *R. solanacearum* in which he had taken rhizome, bulb and leaf aqueous extracts. Among them, the aqueous extract of garlic (*Allium sativum*) at 100% concentration found to be very effective and showed an inhibition zone of about 21mm & 24mm in diameter at 24 & 48 hours after incubation. Evaluation of aqueous extract of different botanicals at different concentration against *R. solanacearum* under *in vitro* conditions

Data presented in table 3 showed that out of the four botanicals extract tested, garlic bulb extract recorded maximum inhibition zone (9.06 mm) at 1% concentration whereas other botanicals extract failed to inhibit the growth of *R. solanacearum*. At 5% concentration, garlic extract exhibited the highest inhibition zone (11mm) followed by guava (8.38 mm) and turmeric (8.10mm) respectively; whereas marigold extract failed to inhibit the growth of *R. solanacearum*. At 10% concentration, garlic extract exhibited the highest inhibition zone (12mm) followed by marigold (10.10mm), guava (8.99mm), and turmeric (8.36) extracts respectively. At 15% concentration garlic extract exhibited highest inhibition zone (13.11) followed by extracts of marigold (11.33mm), guava (11.32mm), and turmeric (8.77mm) (Plate-2 and Fig.-2). The statistical analyses also showed that garlic showed superiority over all the botanicals extract at all the levels of concentration followed by marigold and guava which is statistically at par at 10 and 15% concentration. Gopalakrishnan *et al.*, (2014)^[5] demonstrated *in vitro* evaluation of botanicals against *R. solanacearum* in which they had taken 23 botanicals. Among them, garlic extract showed a higher zone of inhibition of about 49mm diameter at 10% concentration.

Screening of crude ethanol extracts of botanicals against *R. solanacearum*. Ethanol extracts of different botanicals were evaluated for their antibacterial property against *Ralstonia solanacearum*. Extracts were taken at 100% concentration. Observations were taken in the form of inhibition zone (mm) after 24 h of incubation. The results revealed that out of 11 botanicals extract tested, 7 botanicals extract (garlic, guava, marigold, turmeric, neem, papaya and tulsi) exhibited significant antibacterial property whereas 4 botanicals extract (datura, onion, ginger and lemongrass) failed to inhibit the growth of *R. solanacearum in vitro*. The result presented in table- 4 and plate-3.

Out of the seven botanicals extract garlic bulb extract exhibited maximum zone of inhibition (29.77mm) followed by guava leaf extract (22.88mm), marigold (19.77mm), papaya (15.11mm), neem (14.88mm), tulsi (14.10mm) and turmeric extracts (11.33mm) (Fig.-3). Similar findings were made by Owoseni and Sangoyomi (2014), who reported that *A. Sativum* ethanolic extracts had the widest zone of inhibition (12.5mm) out of 10 ethanolic extracts at 100% concentration.

Evaluation of ethanolic extract of botanicals at different concentration against *R. solanacearum*

Out of the four botanicals extract tested only garlic bulb exhibited antibacterial activity at 1% concentration and the inhibition zone produced by garlic extracts was 8.27 mm against *R. solanacearum* (Fig.-4). At 5 and 10% concentration garlic showed maximum inhibition zone (9.05 mm and 11.72 mm) and guava showed (8.05mm and 8.76mm) inhibition

zone respectively, whereas marigold and papaya extract failed to inhibit the growth of *R. solanacearum* respectively. At 15% concentration marigold and papaya also showed antibacterial activity against *R. solanacearum* but the maximum inhibition zone was recorded in garlic (13mm) followed by guava (10.94), marigold (8.66mm) and papaya (7.88mm). Result presented in table 5 and fig. 4. The statistical analysis also showed that garlic showed superiority over all the plant extracts at all levels of concentration followed by guava. However, marigold and papaya statistically at par at 15% concentration.

Conclusion

Eleven aqueous and ethanol extracts of botanicals were studied *in vitro* against *R. solanacearum*. Among aqueous extract, garlic extract (*Allium sativum*) at 100% concentration found to be very effective and showed the highest inhibition zone of about 30.21mm. After screening, four aqueous extracts of botanicals (garlic, guava, marigold and turmeric) which showed inhibition zone ≥ 15 mm was evaluated against *R. solanacearum* at 1%, 5%, 10% and 15% concentration. Among them, garlic showed superiority over all the botanicals extract at all the levels of concentration. In ethanol extract, garlic extract (*Allium sativum*) at 100% concentration produced maximum inhibition zone 29.77mm. After screening, four ethanol extracts of botanicals (garlic, guava,

marigold and papaya) which showed inhibition zone ≥ 15 mm was evaluated against *R. solanacearum* at 1%, 5%, 10% and 15% concentration. Among them, garlic showed superiority over all the botanicals extract at all the levels of concentration.

Table 1: List of botanicals used for the preparation of extracts

S. No.	Local name	Scientific name	Family	Plant parts used
1	Ginger	<i>Zingiber officinale</i>	Zingiberaceae	Rhizome
2	Neem	<i>Azadirachta indica</i>	Meliaceae	Leaf
3	Tulsi	<i>Ocimum gratissimum</i>	Lamiaceae	Leaf
4	Garlic	<i>Allium sativum</i>	Amaryllidaceae	Bulb
5	Onion	<i>Allium cepa</i>	Amaryllidaceae	Bulb
6	Guava	<i>Psidium guajava</i>	Myrtaceae	Leaf
7	Papaya	<i>Carica papaya</i>	Caricaceae	Leaf
8	Turmeric	<i>Curcuma longa</i>	Zingiberaceae	Rhizome
9	Lemon grass	<i>Cymbopogon citratus</i>	Poaceae	Leaf
10	Marigold	<i>Tagetes erecta</i>	Asteraceae	Leaf
11	Datura	<i>Datura stramonium</i>	Solanaceae	Leaf

Table 2: Effect of crude aqueous extract of different botanicals on *R. solanacearum* under *in vitro* condition

S. No.	Botanicals	Inhibition zone (mm) against <i>R. solanacearum</i>
1.	Neem	13.77
2.	Tulsi	0
3.	Garlic	30.21
4.	Onion	0
5.	Guava	25.11
6.	Papaya	14.55
7.	Turmeric	17.77
8.	Lemon Grass	0
9.	Datura	0
10.	Marigold	16.21
11.	Ginger	0
12.	Streptomycin(500ppm)	11.72
13.	Control	0
SEm \pm		0.66
CD at 5%		1.93

Table 3: Effect of aqueous extract of different botanicals at the different concentrations on *R. solanacearum* under *in vitro* conditions.

S. No.	Botanicals	Inhibition zone (mm) against <i>R. solanacearum</i>			
		1%	5%	10%	15%
1.	Guava	0	8.38	8.99	11.32
2.	Garlic	9.06	11	12	13.11
3.	Marigold	0	0	10.10	11.33
4.	Turmeric	0	8.10	8.36	8.77
5.	Streptomycin 500ppm	11.72			
6.	Control	0			
Factors		CD at 5%		SEm \pm	
Botanicals (A)		0.88		0.30	
Concentration (B)		0.72		0.25	
Interaction (A \times B)		1.76		0.61	

Table 4: Effect of crude ethanol extract of different botanicals on *R. solanacearum* under *in vitro* conditions

S. No.	Botanicals	Inhibition zone (mm) against <i>R. solanacearum</i>
1.	Neem	14.88
2.	Tulsi	14.10
3.	Garlic	29.77
4.	Onion	0
5.	Guava	22.88

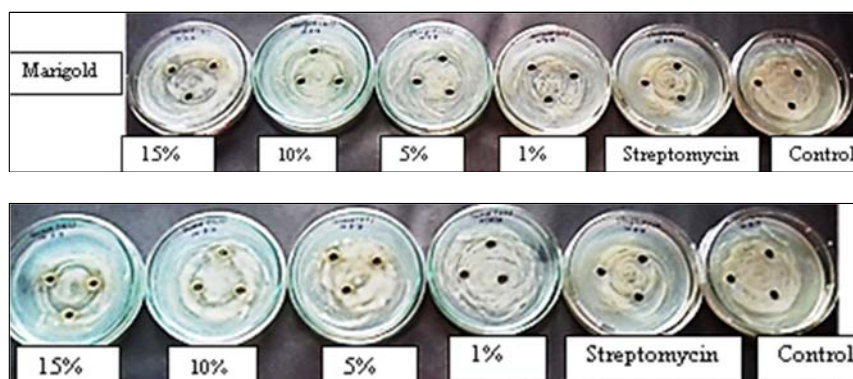
6.	Papaya	15.11
7.	Turmeric	11.33
8.	Lemon Grass	0
9.	Datura	0
10.	Marigold	19.77
11.	Ginger	0
12.	Streptomycin(500ppm)	11.72
13.	Control	0
SEm±		0.47
CD at 5%		1.38

Table 5: Effect of ethanol extract of different botanicals at the different concentrations on *R. solanacearum* under *in vitro* conditions.

S. No.	Botanicals	Inhibition zone (mm) against <i>R. solanacearum</i>			
		1%	5%	10%	15%
1.	Guava	0	8.05	8.76	10.94
2.	Garlic	8.27	9.05	11.72	13
3.	Marigold	0	0	0	8.66
4.	papaya	0	0	0	7.88
5.	Streptomycin 500ppm	11.72			
6.	Control	0			
Factors		CD at 5%		SEm±	
Botanicals (A)		0.87		0.30	
Concentration (B)		0.71		0.25	
Interaction (A×B)		1.75		0.61	



Plate 1: Inhibition zone produced by aqueous extracts of different botanicals against *R. solanacearum* at 100% concentrations



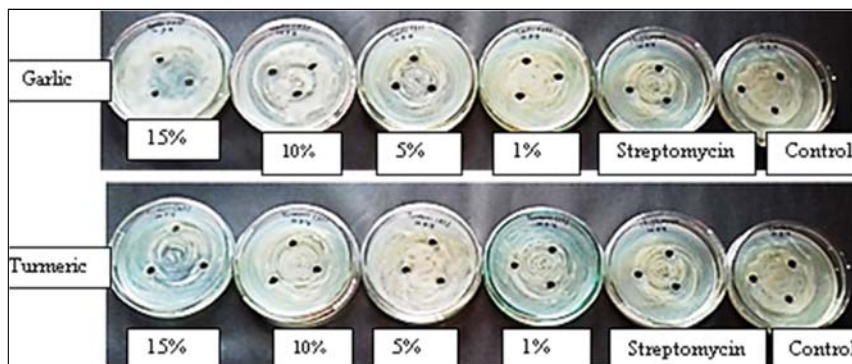


Plate 2: Inhibition zone produced by aqueous extracts of different botanicals against *R. solanacearum* at 1%, 5%, 10% and 15% concentrations

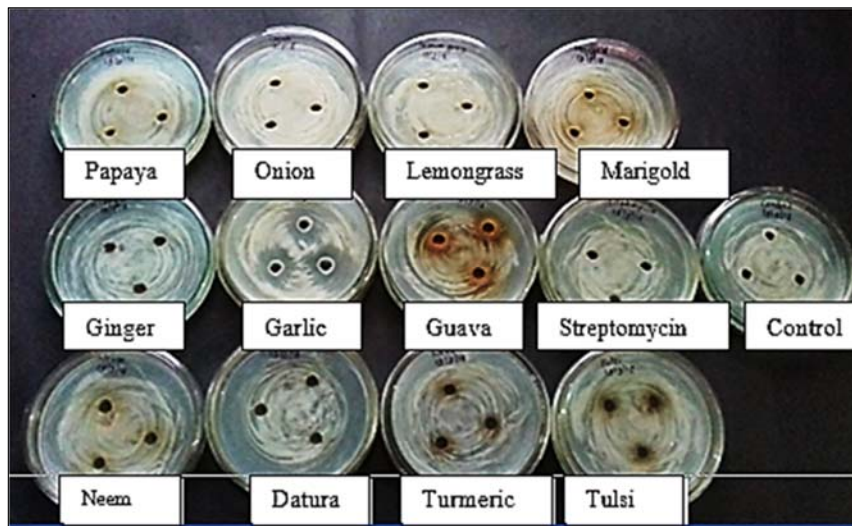


Plate 3: Inhibition zone produced by ethanol extracts of different botanicals against *R. solanacearum* at 100% concentrations



Plate 4: Inhibition zone produced by ethanol extracts of different botanicals against *R. solanacearum* at 1%, 5%, 10% and 15% concentrations

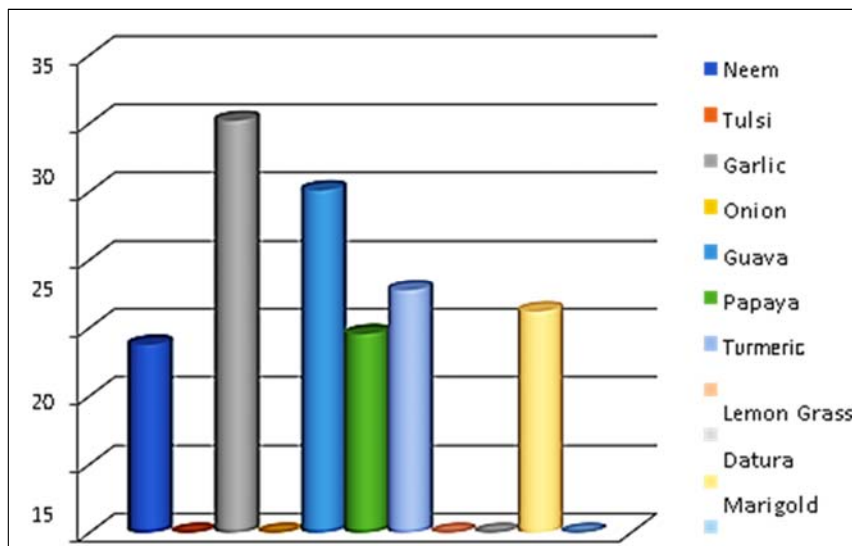


Fig 1: Inhibition zone produced by aqueous extracts of different botanicals against *R. solanacearum* at 100% concentrations under *in vitro* conditions

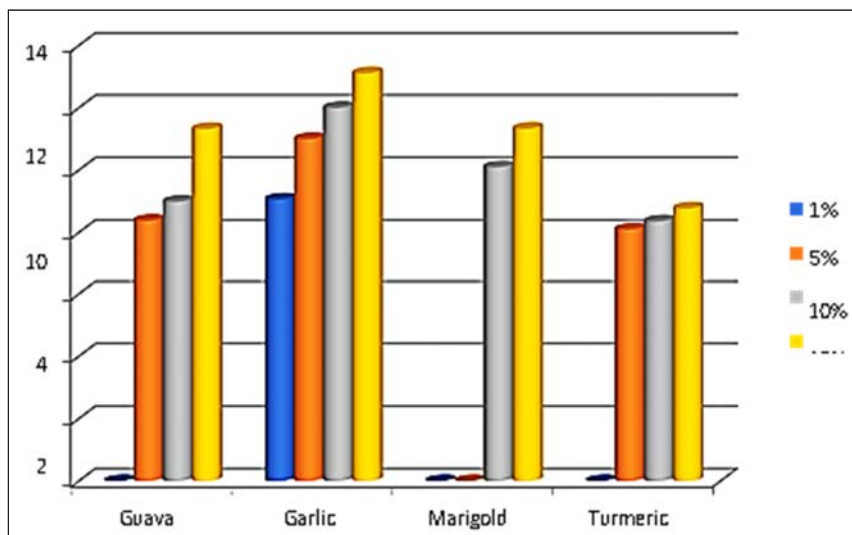


Fig 2: Inhibition zone produced by aqueous extracts of different botanicals against *R. solanacearum* at 1%, 5%, 10% and 15% concentrations under *in vitro* conditions

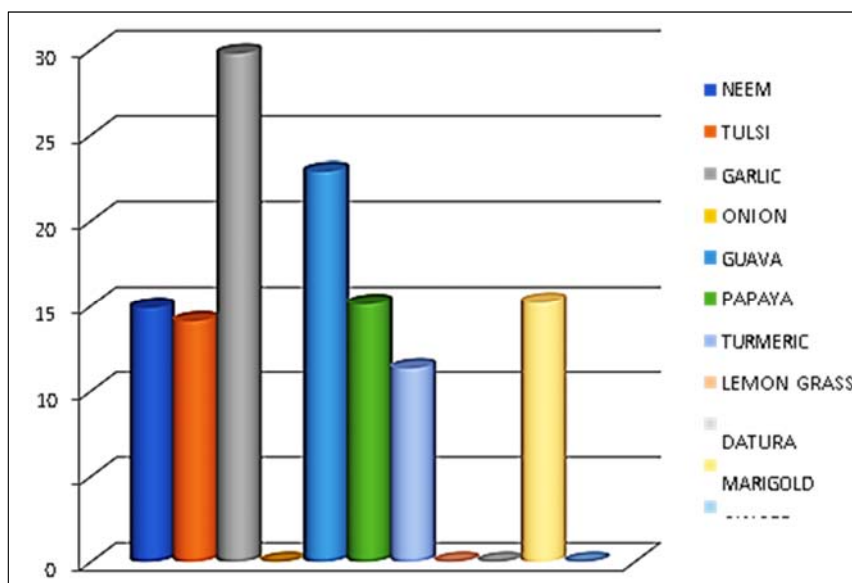


Fig 3: Inhibition zone produced by ethanol extracts of different botanicals against *R. solanacearum* at 100% concentrations under *in vitro* conditions

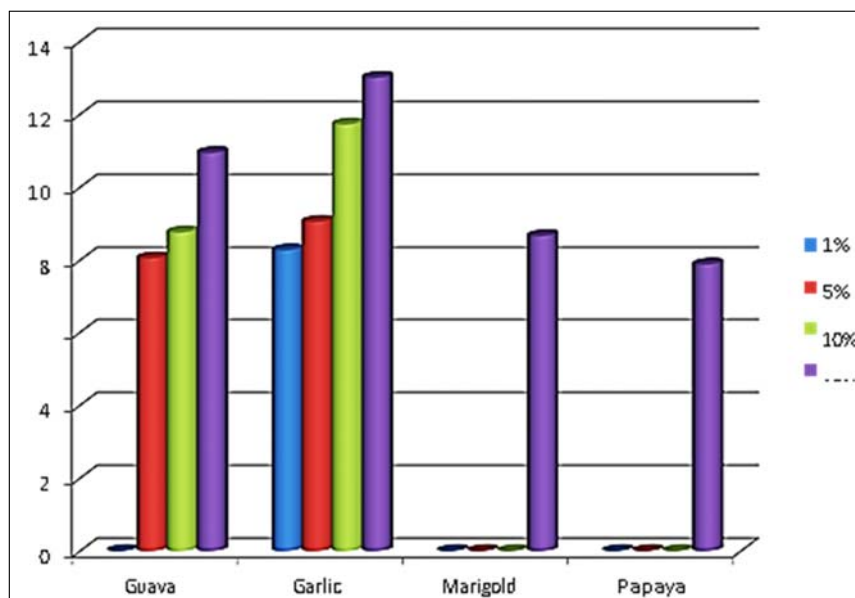


Fig 4: Inhibition zone produced by ethanol extracts of different botanicals against *R. solanacearum* at 1%, 5%, 10% and 15% concentrations under *in vitro* conditions

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