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# Responses of different light intensities and continue light during dark period on rice (*Oryza sativa* L.) seed germination and seedling development

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#### Abstract

Temperature, humidity and moisture content are the important abiotic component regulating seedling establishment in plants including rice. Light factor intensity and duration are also important environmental factors regulating rice growth and development. In the growth and development of rice crop germination followed by seedling establishment is the foremost and very important growth stages. Light component such as intensity, direction and duration are the regulating factor for several physiological, biochemical and molecular processes in plants. To consider these facts, in the present piece of work rice seed of HUR-105 grown under different light regimes, from T1 (2000 lux for 12 h during day time + 12 h dark period) (lower light intensity), T<sub>2</sub> (4000 lux for 12 h during day time+ 12 h dark period), T<sub>3</sub> (6000 lux for 12 h during day time + 12 h dark period) (moderate), T<sub>4</sub> (9000 lux for 12 h during day time + 12 h dark period) (optimum) and to T<sub>5</sub> (9000 lux for 12h during day time + 200 lux for 12 h during night time). Germination, seedling growth and biochemical parameters were observed at different time intervals. It was observed that germination %, germination index (GI), germination rate index (GRI), coefficient of velocity of germination (CVG), mean germination rate (MGR), seedling vigour (SV),  $\alpha$ -amylase activity and soluble sugar content significantly reduced in both the treatments T<sub>1</sub> and T<sub>5</sub>. Further, the mean time germination and insoluble sugar content were increased in  $T_1$  and  $T_5$ treatment. The present experiment concluded that both lower light intensity  $(T_1)$  and addition of low light during dark period (considered as night light pollution) causes stress condition and reduce germination and seedling establishment potential of rice crop.

**Keywords:** Light intensity, seed germination, seedling growth and development, seedling vigor, night interruption, fluorescent light

#### Introduction

Rice is the most important cereal crop and staple food for the half of the world population. To secure food security in near future, we need to double the rice production and productivity (Dawe 2012)<sup>[6]</sup>. Unfortunately, abiotic and biotic stresses and climate change are the major challenges in front of agriculturist to achieve this goal (Godfray et al. 2010; Singhal et al. 2017b) <sup>[9, 21]</sup>. In the atmospheric factors, light is considered as foremost component in regulation of diverse plant processes. Light regulates the seed germination, seedling development, metabolism, photosynthesis, Photomorphogenesis, Photoperiodism and circadian rhythms in plants (Bose & Srivastava 1980; Smith 1982; Jiao et al. 2007; Bose et al. 2018) <sup>[5, 23, 11, 4]</sup>. Different light component such as light intensity, duration, and direction have important role in regulation of plant growth and development (Singhal et al. 2017a)<sup>[20]</sup>. First important component of light is light intensity, which effects plant growth and development. Nearly 80% of the grown rice area in Eastern & North East India is rain-fed and exposed to several abiotic stresses, among them lower light intensity is the most prevalent across the entire region. Lower light intensity affects the rice growth and development by influencing physiological traits (leaf area, tiller number, plant height, and total dry matter production), biochemical parameters such as (chlorophyll content, starch content), antioxidant production, growth parameters (crop growth rate, relative growth rate) and yield attributes (Liu et al., 2014; Nag 2017) <sup>[15, 16]</sup>. Most disaster effects of low intensity were observed at the time of reproductive stage, which is responsible for huge penalty in yield potential (Nag 2017; Dutta et al. 2017) [16, 8].

Plants are adapted to the world of day and night (diurnal) changes in light components and develop pigment system and photoreceptors accordingly to sense these environmental changes from the million years. Second important component of light is light duration, which control the photoperiodic response, circadian rhythm and flowering phenomenon in plant [Song *et al.*]

2012]<sup>[24]</sup>. Interruption of night light period change the natural diurnal phenomenon in plants and showed the drastic effects on plant from germination to yield (Singhal et al. 2019)<sup>[22]</sup>. Continuation of dark period by the very low light flux to high light cause modulation in leaf number, leaf length, vegetative growth, flowering in long day plant, photosynthesis (Blanchard & Runkle 2010; Kim et al. 2011; Kim et al. 2015) <sup>[3, 12, 13]</sup>. Furthermore, in the modern period human development and settlement processes produces excess light during night time cause photo pollution or night light pollution. Effect of night light pollution are enormous such as proximity to street lights was found to promote vegetative growth, delay in development and flowering and decrease vield in soybean (Palmer et al. 2017)<sup>[17]</sup>. It also act as a repressor of photosynthesis and growth by inducing oxidative damage in chloroplasts (Kwak et al. 2017) [14]. To consider these above facts an experiment on rice crop by using different intensity of artificial light and using continuation of light during night time was conducted. Various seed germination and seedling establishment parameter were measured, which are described followed.

## **Material and Method**

The present piece of work was undertaken during the years 2016-17. This experiment repeated three times during the same year. Details of the materials and methods used in this experiment have been described as follows.

## **Experimental site**

This experiment was conducted in the Seed Priming and Seed Physiology Laboratory of the Department of Plant Physiology, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, India. During day time temperature was maintained at 25±2 °C to 30±2 °C C and night temperature maintained at 18±2 °C to 20±2 °C. A light condition in the laboratory was maintained by the T5 28 W, 6500 K fluorescent lamp. Light intensity was measured by the help of CHY 332 light Meter and measured light intensity was in case of T<sub>1</sub> (2000 lux for 12 h during day time + 12 h dark period) (lower light intensity), T<sub>2</sub> (4000 lux for 12 h during day time+ 12 h dark period),  $T_3$  (6000 lux for 12 h during day time + 12 h dark period) (moderate), T<sub>4</sub> (9000 lux for 12 h during day time + 12 h dark period) (optimum) and to  $T_5$  (9000 lux for 12h during day time + 200 lux for 12 h during night time). Rice (Oryza Sativa var. HUR-105 (Malviya Sugandhit Dhan) was selected as the plant material to be used in the experiment. This variety developed by the Department of Genetics and Plant Breeding, Institute of Agriculture Sciences, Banaras Hindu University, India.

## Layout of the experiment

The experiment was laid down in a completely randomized design (CRD) with three replications. Each treatment in the experiment was repeated thrice for each replication. Differences between the means were calculated at standard deviation between replications.

## In vitro seed sterilization

Seeds of uniform size were selected, and surface sterilized before using for the experiment. They were first washed with tap water for 7 minutes and then sterilized with  $HgCl_2$  (0.1%) for 5 minutes and then washed with sterile distilled water for 4 to 5 times.

## **Germination studies**

Seed were grown in 30cm diameter Petri plates and recorded the observations at various time intervals. plumule, radical and seedling length in cm was calculated by the scale at 2, 3, 5 and 7 days after seed germination, The fresh and dry weight of embryo+ endosperm (in mg) were observed at 2, 3, and 5 days after sowing by electronic weighing machine. Fresh weight of seedling (in mg) were observed at 2, 3, and 5 days after sowing by electronic weighing machine.

## Final germination percentage (FGP in %)

FGP in % was calculated by the formula

 $=\frac{\text{Total no.of seeds germinated on end day}}{\text{No.of seeds present in Petri dish}} \times 100$ 

# Mean time germination (MTG in days)

For determining mean germination time was calculated by the formula given by the Al-Mudaris (1998)<sup>[1]</sup>.

MTG (in days) =  $\Sigma F^* x / \Sigma F$ 

Where F is the number of seeds germinated on day x

## Germination index (GI)

Germination index of wheat seed was calculated by the formula given by Ranal *et al.* (2009) <sup>[18]</sup>.

$$GI=(3 \times n_1) + (2 \times n_2) + \cdots + (1 \times n_4)$$

 $n_1$ = 1 day germinated seed  $n_2$ = second day germinated seed  $n_4$  = fourth day germinated seed

## Germination rate index (GRI)

GRI is calculated by the formula given by the Al-Mudaris (1998) <sup>[1]</sup>.

 $GRI = G1/1 + G2/2 + \cdot \cdot + Gx/x$ 

G1 = Germination percentage  $\times 100$  on the first day after sowing

 $G2 = Germination percentage \times 100$  on the second day after sowing

## Coefficient of velocity of germination (CVG)

CVG was calculated by the formula given by Al-Mudaris (1998)<sup>[1]</sup>.

 $CVG = N1 + N2 + \cdots + Nx/100 \times N1T1 + \cdots + NxTx$ 

N=No. of seeds germinated each day, T=No. of days from seeding corresponding to N

## Mean germination rate (MGR)

MGR was calculated by the formula given by Ranal *et al.*  $(2009)^{[18]}$ .

MGR = CVG/100

## Seedling vigor (SV)

Seedling vigor index measured in 7 day old seedling was calculated by the formula given by Goodi & Sharifzadeh  $(2006)^{[10]}$ . SV= Germination % X Seedling length (in cm)

## α-amylase activity

For the estimation of the activity of  $\alpha$ -amylase (EC3.2.1.1) enzyme in the endosperm of germinating rice seeds was calculated by the method of Bernfeld, P. (1955)<sup>[2]</sup>.

### Soluble and insoluble sugar content

For the estimation of the soluble and insoluble sugar content in the endosperm of germinating rice seeds, the method of Dubois *et al.* (1956) <sup>[7]</sup> was followed.

#### Statistical analysis

Statistical analysis of all above measured parameter done by using software statistical package for the social sciences (SPSS) and window excel.

#### Results

## Seed germination and seedling growth

Fig. 1(a) represents the germination percentage of HUR-105 at different time intervals. At 24 h after sowing highest germination % was noted in treatment  $T_2$  (82) subsequent by  $T_3$  and  $T_4$  (78), while the lower germination percentage observed in case of  $T_1$  (61). At 48 h the highest germination percentage recorded in  $T_4$  (97) subsequent by  $T_3$  (92). The lower germination percentage at 48 observed in  $T_5$  (83). At 72h T4 show higher germination percentage (98) followed by  $T_3$ (95) and lowest germination observed in  $T_5$  (89).



Fig 1a: Represents the effect of low intensity light during night time and different light intensities on germination percentage of rice (*Oryza sativa* L.)

## Plumule and radicle length

Fig. 1(b) and Fig 1(c) represent the plumule length and radicle length at different time interval, both are decreased by night light treatment (T<sub>5</sub>) and low light condition (T<sub>1</sub>). However, the highest plumule and radicle length at  $2^{nd}$  and  $3^{rd}$  day were observed in T<sub>2</sub> and the lowest plumule and radicle length observed in T<sub>5</sub>. At 5<sup>th</sup> day the highest plumule length (5.425 cm) observed in T<sub>3</sub> followed by T<sub>4</sub> (4.844 cm) and lowest plumule length observed in T<sub>1</sub> (4.268 cm). The highest radicle length at 5<sup>th</sup> day observed in T<sub>3</sub> (5.7916 cm) subsequent by T<sub>4</sub> (5.483 cm) and lowest observed in T<sub>5</sub> (4.858 cm). On the 7<sup>th</sup> day, the highest plumule length observed in T<sub>3</sub> (7.558 cm) followed by T<sub>4</sub> (6.34 cm) and lowest plumule length observed in T<sub>1</sub> (4.961 cm). The highest radicle length at 7<sup>th</sup> day observed in T<sub>4</sub> (7.65 cm) subsequent by T<sub>3</sub> (7.1416 cm) and lowest observed in T<sub>5</sub> (5.42 cm). At 10<sup>th</sup> day T<sub>3</sub> again obtained highest plumule (8.63 cm) and radicle length (9.08 cm) subsequent by T<sub>2</sub> (7.94 cm and 8.5 cm) and lowest plumule length observed in T<sub>1</sub> (6.65 cm) but lowest radicle length observed in T<sub>5</sub> (7.03 cm).



Fig 1b: Represents the effect of low intensity light during night time and different light intensities on plumule length of rice (Oryza sativa L.)



Fig 1c: Effect of low intensity light during night time and different light intensities on radical length of rice (Oryza sativa L.).

## Seedling length (cm)

Fig. 2(a) represent the seedling length of rice at different time interval. At  $2^{nd}$  day the highest seedling length were observed in T<sub>2</sub> (1.89 cm) subsequent by T<sub>4</sub> (1.73) and lowest seedling length were observed in T<sub>5</sub> (1.47). At  $3^{rd}$  day the highest seedling length were observed in T<sub>2</sub> (4.25) followed by T<sub>5</sub> (3.64) and lowest seedling length were observed in T<sub>1</sub> (3.39). At  $5^{th}$  day the highest length was observed in T<sub>3</sub> (11.21 cm)

subsequent by  $T_4$  (10.32) and lowest seedling length were observed in  $T_5$  (9.28 cm). At 7<sup>th</sup> day the highest seedling length was observed in  $T_3$  (14.7 cm) subsequent by  $T_4$  (13.96 cm) and lowest seedling was observed in  $T_1$  (10.49 cm). At 10<sup>th</sup> day the highest seedling length were observed in  $T_3$ (17.72 cm) subsequent by  $T_2$  (16.51 cm) and lowest seedling length was observed in  $T_1$  (13.72 cm).



Fig 2a: Represents the effect of low intensity light during night time and different light intensities on Seedling Length of rice (Oryza sativa L.)

## Seedling vigor I

Fig. 2(b) represent the seedling vigor of rice at different time interval. At  $2^{nd}$  day the highest seedling vigour found in  $T_2$  (1.724) subsequent by  $T_4$  (1.68) and lowest observed in  $T_5$  (1.22). At  $3^{rd}$  days the highest seedling vigour observed in  $T_2$  (3.995) followed by  $T_4$  (3.55) and lowest observed in  $T_1$  (3.22). At  $5^{th}$  day  $T_3$  showed highest vigour (10.65)

subsequent by  $T_4$  (10.12) and lowest observed with  $T_5$  (8.26). The highest vigour at 7<sup>th</sup> day found in  $T_3$  (13.96) subsequent by  $T_4$  (13.68) and lowest vigour observed in  $T_1$  (9.96). At 10<sup>th</sup> day the highest vigour was observed in  $T_3$  (16.83) subsequent by  $T_2$  (15.52) and lowest seedling vigour observed in  $T_5$  (12.51).



Fig 2b: Represents the effect of low intensity light during night time and different light intensities on seedling vigor of rice (Oryza sativa L.)

#### Fresh weight of seedling

Fig. 2(c) represent the fresh weight of seedling of rice at different time interval. At  $2^{nd}$  and  $3^{rd}$  day the highest seedling fresh weight was observed in T<sub>3</sub> (58.33mg and 215 mg)

subsequent by  $T_4$  (35 mg and 160 mg) and lowest was observed in  $T_1$  (21.66 and 55 mg). At 5<sup>th</sup> day the highest seedling fresh weight was observed in  $T_3$  (410 mg) followed by  $T_2$  (370 mg) and lowest was observed in  $T_1$  (255 mg).



Fig 2c: Represents the effect of low intensity light during night time and different light intensities on fresh weight Seedling of rice (*Oryza sativa* L.)

## Final germination percentage (FGP)

Table 1 represents the final germination percentage (FGP) of rice at different time interval. The highest FGP was observed in  $T_3$  (98) subsequent by  $T_1$  and  $T_4$  (95), and lowest FGP observed in  $T_5$  (89).

## Mean time germination (MTG in days)

Table 1 represents the mean time germination (MTG in days). The highest MGT was observed in  $T_1$  (1.44) subsequent by  $T_5$  (1.28) and lowest MGT observed in  $T_2$  (1.16).

#### Germination index (GI)

Table 1 represents the germination index (GI). Higher germination index were observed in  $T_3$  (526) subsequent by  $T_2$  (522) and lower GI observed in  $T_1$  (452).

#### Germination rate index (GRI)

Table 1 represents the germination rate index (GRI). Highest GRI were observed in  $T_3$  (87.83) subsequent by  $T_2$  (87.50) and the lowest GRI was observed in  $T_1$  (76.67).

## Coefficient of velocity of germination (CVG)

Table 1 represents the coefficient of velocity of germination (CVG). The highest CVG observed in  $T_2$  (86.24) subsequent by  $T_4$  (82.61) and lowest CVG observed in  $T_1$  (69.34).

### Mean germination rate (MGR)

**Table 1:** Represents the mean germination rate. The highest MGRobserved in  $T_2$  (0.86) subsequently by  $T_4$  (0.83) and lowest observedin  $T_1$  (0.69).

Treatment	FGP (%)	MTG (day)	CVG	GI	GRI (%/day)	MGR
T1	95	1.44	69.34	452.00	76.67	0.69
T2	94	1.16	86.24	522.00	87.50	0.86
T3	98	1.21	82.35	526.00	87.83	0.82
T4	95	1.21	82.61	513.00	86.00	0.83
T5	89	1.28	78.07	465.00	78.50	0.78

Table 1 represents the final germination %, mean time germination, coefficient of velocity of germination, germination index, germination rate index and mean germination rate of rice variety HUR-105 under different light regimes.

#### α-amylase activity

Fig. 3(a) represents the alpha amylase activity at different time interval. The highest induction of  $\alpha$ -amylase activity was observed in T<sub>4</sub> than T<sub>3</sub> at all days and lowest induction was observed in T<sub>1</sub>. T5 also showed lower induction of  $\alpha$ -amylase activity.  $\alpha$ -amylase activity was increased steadily up to 72 h and then decline at 96 h in all conditions. Mean value of  $\alpha$ -amylase activity of 96 h clearly showed the highest enzyme induction in T<sub>4</sub> and lowest in T<sub>1</sub> and T<sub>5</sub>.



Fig 3a: Represents the effect of low intensity light during night time and different light intensities on α-amylase activity of rice (Oryza sativa L.)

#### Soluble sugar

Fig. 3(b) represents the soluble sugar content at different time intervals. The soluble sugar content highest in  $T_4$  than T3 at 24 h, 48hrs and 72h and at 96 h  $T_3$  was more soluble sugar content than  $T_4$ . The increment of soluble sugar content at 96

h in all condition/treatments due to induction/synthesis of beta amylase activity. The soluble sugar content also significant in  $T_2$ . The lowest soluble sugar content were observed in  $T_1$  than  $T_5$  at different time intervals.



Fig 3b: Effect of low intensity light during night time and different light intensities on soluble sugar content of rice (Oryza sativa L.)

## Insoluble sugar

Fig. 3(c) represents the non-soluble sugar content at different time intervals. The highest non-sugar content was observed in  $T_1$  than  $T_2$  and lowest insoluble sugar content observed in  $T_3$ 

at 24hrs. After 24 h the highest insoluble sugar content observed  $T_1$  than  $T_5$  and least insoluble sugar content was observed in  $T_3$  at 48 to 96 h time intervals.



Fig 3c: Represents the effect of low intensity light during night time and different light intensities on insoluble sugar of rice (Oryza sativa L.)

## Fresh weight of embryo + endosperm (mg)

Table 2 represents the fresh weight of embryo + endosperm in (mg) at different time intervals. At  $2^{nd}$  day the highest fresh weight of embryo + endosperm was observed in T<sub>1</sub> (111.33) and lowest value observed in case of T<sub>3</sub> (110.00). At  $3^{rd}$  day the highest value observed in T<sub>5</sub> (107.33) followed by T<sub>1</sub> (106.33) and lowest value observed in T<sub>4</sub> (100.00). At  $5^{th}$  day the highest weight of embryo + endosperm observed in T<sub>1</sub> (104.67) subsequent by T<sub>5</sub> (102.67) and lowest fresh weight of embryo + endosperm observed in T<sub>4</sub> (99.33).

## Dry weight of embryo+ endosperm (mg)

Table 2 represent the dry weight of embryo + endosperm at different time intervals. At  $2^{nd}$  day the highest dry weight of embryo + endosperm was observed in T<sub>1</sub> (90.99) subsequent by T<sub>5</sub> (90.01±0.46) and lowest value observed in T<sub>3</sub> (88.00). At  $3^{rd}$  day the highest dry weight of embryo + endosperm observed in T<sub>1</sub> (89.45) followed by T<sub>5</sub> (87.22) and lowest value was observed in T<sub>4</sub> (80.04). At  $5^{th}$  day the highest dry weight of embryo + endosperm weight of embryo + endosperm was observed in T<sub>1</sub> (87.00) subsequent by T<sub>5</sub> (85.02) and lowest dry weight of embryo + endosperm was observed in T<sub>1</sub> (87.00) subsequent by T<sub>5</sub> (85.02) and lowest dry weight of embryo + endosperm was observed in T<sub>3</sub> (78.41).

Table 2: Represents the fresh and dry weight of embryo + endosperm of rice variety HUR-105 under different light regimes

	Fresh weight of embryo + endosperm (mg)			Dry weight of embryo+ endosperm (mg)		
Treatments	2 day	3 day	5 day	2 day	3 day	5 day
T1	111.33±1.25 <sup>b</sup>	106.33±0.94 <sup>b</sup>	104.67±0.94°	90.99±0.46 <sup>e</sup>	89.45±0.33e	87.00±0.33 <sup>d</sup>
T2	106.33±0.94 <sup>a</sup>	101.33±1.25 <sup>a</sup>	100.67±0.94 <sup>a</sup>	89.00±0.20°	84.10±0.27°	80.02±0.23 <sup>b</sup>
T3	$110.00 \pm 0.82^{b}$	101.67±0.94 <sup>a</sup>	99.67±0.41 <sup>a</sup>	88.00±0.34 <sup>b</sup>	83.22±0.15 <sup>b</sup>	78.41±0.13 <sup>a</sup>

T4	104.67±0.47 <sup>a</sup>	100.00±0.82ª	99.33±0.47 <sup>a</sup>	84.00±0.33ª	$80.04 \pm 0.22^{a}$	79.99±0.36 <sup>b</sup>
T5	111.00±0.82 <sup>b</sup>	107.33±1.25 <sup>b</sup>	102.67±0.47 <sup>a</sup>	$90.01 \pm 0.46^{d}$	87.22±0.23 <sup>d</sup>	85.02±0.07°

## Discussion

Present study is done to observe the effect of different light intensities and dark period added with low light intensity on seed germination and seedling establishment. Various seed germination and seedling establishment parameters were observed in different light regimes. Most of the observed parameters were lower in case of treatment  $T_1$  and  $T_5$ , represent the lower seedling establishment and development. Germination % is the potential of seed to emerge from the dormant stage and it was observed that both light condition i.e. low light intensity and dark period added with low light intensity (night light pollution) as stress factor during germination. Mean time germination indicates the speed of germination capacity of seed and lowest speed of MTG represents the lower level of shoot and seedling growth. Mean germination rate represents the rate of seed germination, higher value represents the good seedling growth. MGR data correlated with the seedling length. CVG value gives the indication of the rapidity of germination, this value correlated with the FGP. The GRI value indicates the percentage of germination on each day of the germination period, which was lowest in T<sub>1</sub>. All these measured data lower in case of treatment  $T_1$  and  $T_5$ , because these treatment have lower germination percentage.

Seed stored the necessary food reserve for the seedling growth. During initial stage food reserve material used by the seedling. Therefore, measuring regarding fresh weight of seedling represents the biomass transferred from seed to seedling, which was lower in case of  $T_1$  and  $T_5$  treatments. Which might be representing a cause for lower seedling growth in both the treatments. This data correlate with the fresh and dry weight of embryo and endosperm  $\alpha$ -amylase activity, non-soluble and sugar content of seed. A higher embryo+ endosperm weight represents the lower rate of translocation of food reserve from endosperm/ seed towards newly emerging seedlings. It is well established that  $\alpha$ amylase is a foremost important enzyme in starch breakdown in cereals. In the present case also  $T_2$  to  $T_4$  treatment are showing higher activities of  $\alpha$ -amylase in respect to T<sub>1</sub> and T<sub>5</sub>; among these treatment T<sub>4</sub> represented the highest activity which is clearly reflected from the mean days observation of this study Sananda and Bandana (2012)<sup>[19]</sup> and it may proof that high light intensity during day time is better as compared to the same light followed by a continuous low light during night time  $(T_5)$ . Therefore, the night light may disturb the rhythmicity of this enzyme.

A higher induction of  $\alpha$ -amylase activity is responsible for the conversion of insoluble sugar to soluble sugar. Which is in agreement with the present study also, where the soluble sugar content was maximum in T<sub>4</sub> treatment in all the days among others, whereas the content of insoluble sugar was more in  $T_1$  and  $T_5$  (Fig. 3c), and that was well correlated with the activity of amylase and soluble sugar content. Therefore, the present study clearly concludes that the different light intensities have influenced effects not only on the germination of seed but it interferes with its sugar metabolism and translocation of the food material from seed to seedling. Further, it can be suggested that the presence of continuous even in very low intensity has a negative impact on the seedling growth and development. Lastly, it is to be presumed that with the increased industrialization and increment in various kind of artificial light in the roadside and farming field may affect not only the crop yield but also biodiversity (Singhal *et al.* 2019)<sup>[22]</sup>.

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## References

- 1. Al-Mudaris M. Notes on various parameters recording the speed of seed germination. Der Tropenlandwirt. 1998; 99:147-54.
- 2. Bernfeld P. Amylases alpha and beta. Meth Enzymol. 1955; 1:149.
- 3. Blanchard MG, Runkle ES. Intermittent light from a rotating high-pressure sodium lamp promotes flowering of long-day plants. Hort. Science. 2010; 45(2):236-241.
- 4. Bose B, Pant B, Singhal RK, Kumar M, Mondal S. Phytochrome: physiology, molecular aspects, and sustainable crop production. Emerging Trends of Plant Physiology for Sustainable Crop Production, 2018, 25.
- 5. Bose B, Srivastava HS. Proteolytic activity and nitrogen transfer in maize seeds during imbibition. Biologia Plantarum. 1980; 22(6):414-419.
- 6. Dawe D. The rice crisis: Markets, policies and food security. Routledge, 2012.
- 7. Dubois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. Colorimetric method for determination of sugars and related substances. Anal. Chem. 1956; 28:350-356.
- Dutta SS, Pale G, Pattanayak A, Aochen C, Pandey A, Rai M. Effect of low light intensity on key traits and genotypes of hilly rice (*Oryza sativa*) germplasm, 2017. http://krishi.icar.gov.in/PDF/ICAR\_Data\_Use\_Licence.p df.
- Godfray HCJ, Beddington JR, Crute IR, Haddad L, Lawrence D, Muir JF *et al.* Food security: the challenge of feeding 9 billion people. Science. 2010; 327(5967):812-818.
- Goodi M, Sharifzadeh F. Evaluation effect of hydropriming in barley difference cultivars. Mag biaban. 2006; 11:99-109.
- 11. Jiao Y, Lau OS, Deng XW. Light-regulated transcriptional networks in higher plants. Nature Reviews Genetics. 2007; 8(3):217.
- 12. Kim YJ, Lee HJ, Kim KS. Night interruption promotes vegetative growth and flowering of Cymbidium. Scientia horticulturae. 2011; 130(4):887-893.
- 13. Kim YJ, Yu DJ, Rho H, Runkle ES, Lee HJ, Kim KS. Photosynthetic changes in Cymbidium orchids grown under different intensities of night interruption lighting. Scientia Horticulturae. 2015; 186:124-128.
- Kwak MJ, Lee SH, Khaine I, Je SM, Lee TY, You HN *et al.* Stomatal movements depend on interactions between external night light cue and internal signals activated by rhythmic starch turnover and abscisic acid (ABA) levels at dawn and dusk. Acta Physiologiae Plantarum. 2017; 39(8):162.
- 15. Liu QH, Xiu WU, Chen BC, Jie GAO. Effects of low light on agronomic and physiological characteristics of rice including grain yield and quality. Rice Science. 2014; 21(5):243-251.

- 16. Nag B. Physiological evaluation for low light tolerance on rice (*Oryza sativa* L.) (Doctoral dissertation, Indira Gandhi Krishi Vishwavidhyalaya, Raipur), 2017.
- Palmer M, Gibbons R, Bhagavathula R, Davidson D, Holshouser D. Roadway Lighting's Impact on Altering Soybean Growth: Illinois Center for Transportation/Illinois Department of Transportation, 2017, 1.
- Ranal MA, Santana DGD, Ferreira WR, Rodrigues MC. Calculating germination measurements and organizing spreadsheets. Brazilian Journal of Botany. 2009; 32(4):849-855.
- Sananda M, Bandana B. Kinetics studies on α-amylase extracted from germinating wheat endosperm of primed and non-primed seeds. Indian Journal of Agricultural Biochemistry. 2012; 25(2):137-141.
- 20. Singhal RK, Kumar V, Kumar S, Choudhary BL. High light stress response and tolerance mechanism in plant. Interdisciplinary journal of Contemporary Research, 2017, 4(1). ISSN: 23938358
- Singhal RK, Sodani R, Chauhan J, Sharma MK, Yashu BR. Physiological Adaptation and Tolerance Mechanism of Rice (*Oryza sativa* L.) in Multiple Abiotic Stresses. Int. J Pure App. Bio Sci. 2017; 5(3):459-66.
- 22. Singhal RK, Kumar M, Bose B. Ecophysiological Responses of Artificial Night Light Pollution in Plants. Russian Journal of Plant Physiology, 2019, 1-13.
- 23. Smith H. Light quality Photoperception, and plant strategy. Annual review of plant physiology. 1982; 33(1):481-518.
- 24. Song Y, Gao Z, Luan W. Interaction between temperature and photoperiod in regulation of flowering time in rice. Science China Life Sciences. 2012; 55(3):241-249.