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## Effect of various treatments on physico-chemical composition of Indian gooseberry (*Emblica officinalis*) candy during storage

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### Abstract

The present investigation was carried out during 2015-16, at Department of Horticulture, V.N.M.K.V., Parbhani, Maharashtra. The aonla candy was made up by using different treatments i.e. sugar syrup solutions, dipping time and drying methods. There are two levels of sugar syrup concentrations i.e. 60% and 70%, three dipping times i.e. 36 hrs. 42 hrs. and 48 hrs. and 3 drying methods i.e. sun drying, solar drying and cabinet drying. 18 treatment combinations were taken in FCRD with 3 replications. The aonla candy was evaluated for various physico-chemical constituents after preparation of candy and during storage of candy. During storage period there is significant decrease was observed in TA, ascorbic acid and tannins whereas TSS was increased in aonla candy. A gradual and significant decrease in TA of aonla candy was observed during 120 days storage period. The decrease in TA during storage might be due to the co-polymerization of organic acids with sugars and amino acids and loss of volatile acids during storage. The decrease in ascorbic acid during storage was observed during storage, the possible reason of reduction in vitamin C could be due to oxidation by oxygen, which resulted in formation of Dehydroascorbic acid. Gradual decline in tannin during storage might be due to their condensation into brown pigments. A significant increase in TSS was observed during storage period.

**Keywords:** Aonla, sugar, candy, titratable acidity, ascorbic acid

### Introduction

Aonla is one of the oldest Indian fruits and considered as “Wonder fruit for health” (Ganachari *et al.*, 2010) [5]. Aonla (*Phyllanthus emblica* L.), a member of family Euphorbiaceae and sub family Phyllanthoides, is native to India, Ceylon, Malaya and China (Mishra *et al.*, 2010) [10]. It is also known as Indian gooseberry, Amalaki, Amla, Amllet, Amolphal, Aovla, Aurna, Chukna, Dhatriphala, Emblic myrobalan, Nelli and Sobju in different parts of world (Agarwal and Chopra, 2004) [2]. India ranks first in the world in area and production of aonla crop (Priya and Khatkar, 2013) [14, 17]. Its cultivation is concentrated mainly in Uttar Pradesh, Maharashtra, Gujarat, Rajasthan, Andhra Pradesh, Karnataka and Tamil Nadu (Sahu *et al.*, 2010) [20]. The main cultivated varieties are Banarasi, Bansi Red, Chakaiya, Desi, Krishna, Kanchan (NA-4), Francis (Hathijool), NA-6, NA-7, NA-8, NA-9, NA-10 and Pink tinged (Pathak *et al.*, 2002; Alam and Singh, 2010) [13, 4]. The area under aonla cultivation in India is about 1, 08,060 hectares with an annual production of 12, 66,460 tonnes (NHB, 2012-13). Aonla is one of the most important medicinal fruits available in North India and is, therefore, used as a major constituent in several Ayurvedic preparations (Rajkumar *et al.*, 2001) [18]. It is a rich source of vitamin C and its content of ascorbic acid is next to that of Barbados cherry (Ganachari *et al.*, 2010) [5]. About 600–900 mg of vitamin C is found in 100 g of aonla pulp (Pokharkar, 2005) [16]. The edible fruit tissues of aonla contain about 3 times more protein and 160 times more vitamin C as compared to apple. The fruits contain leucoanthocyanin or polyphenols which retard the oxidation of vitamin C. Tannins contain gallic acid, elagic acid and glucose, which retard the oxidation of vitamin C and render its value as antiscorbutic in fresh as well as dried conditions (Pareek and Kaushik, 2012) [15]. Aonla has been reported to be hepatoprotective and possesses expectorant, purgative, spasmolytic, antibacterial, hypoglycemic and hypolipidemic activities (Mishra *et al.*, 2010) [10]. Aonla fruits are astringent, carminative, digestive, stomachic, laxative, altrant, aphoridiac, diuretic, antipyretic and trichogenous. They are useful in curing many diseases like diabetes, cough, asthma, bronchitis, headache, ophthalmic disorders, dyspepsia, colic, flatulence, skin diseases, leprosy, jaundice, scurvy, diarrhoea and greyness of hair (Ganachari *et al.*, 2010) [5].

### Material and Methods

The details of materials used and methods adopted for the conduct of experiment are presented in this chapter under appropriate headings.

**Experimental details****1. Factor A(S) - Standard of Sugar Syrup**

S1-60%  
S2-70 %

**2. Factor B(T) -Dipping Time**

T1-36hrs  
T2-42hrs

T3-48hrs

**3. Factor C(M) -Drying Methods**

M1-Sun drying  
M2-Solar drying  
M3-Cabinet drying

The details of treatments are given in following Table

**Table 1:** Details of treatments

Treatments	Treatment Combination	Treatment Details
T <sub>1</sub>	S <sub>1</sub> T <sub>1</sub> M <sub>1</sub>	60% Sugar conc.+ dipping for 36 hrs + sun drying
T <sub>2</sub>	S <sub>1</sub> T <sub>1</sub> M <sub>2</sub>	60% Sugar conc.+ dipping for 36 hrs + solar drying
T <sub>3</sub>	S <sub>1</sub> T <sub>1</sub> M <sub>3</sub>	60% Sugar conc. + dipping for 36 hrs + cabinet drying
T <sub>4</sub>	S <sub>1</sub> T <sub>2</sub> M <sub>1</sub>	60% Sugar conc.+ dipping for 42 hrs + sun drying
T <sub>5</sub>	S <sub>1</sub> T <sub>2</sub> M <sub>2</sub>	60% Sugar conc. + dipping for 42 hrs + solar drying
T <sub>6</sub>	S <sub>1</sub> T <sub>2</sub> M <sub>3</sub>	60% Sugar conc. + dipping for 42hrs + cabinet drying
T <sub>7</sub>	S <sub>1</sub> T <sub>3</sub> M <sub>1</sub>	60% Sugar conc. + dipping for 48hrs + sun drying
T <sub>8</sub>	S <sub>1</sub> T <sub>3</sub> M <sub>2</sub>	60% Sugar conc. + dipping for 48hrs + solar drying
T <sub>9</sub>	S <sub>1</sub> T <sub>3</sub> M <sub>3</sub>	60% Sugar conc. + dipping for 48hrs + cabinet drying
T <sub>10</sub>	S <sub>2</sub> T <sub>1</sub> M <sub>1</sub>	70% Sugar conc. + dipping for 36 hrs + sun drying
T <sub>11</sub>	S <sub>2</sub> T <sub>1</sub> M <sub>2</sub>	70% Sugar conc. + dipping for 36 hrs + solar drying
T <sub>12</sub>	S <sub>2</sub> T <sub>1</sub> M <sub>3</sub>	70% Sugar conc. + dipping for 36 hrs + cabinet drying
T <sub>13</sub>	S <sub>2</sub> T <sub>2</sub> M <sub>1</sub>	70% Sugar conc. + dipping for 42 hrs + sun drying
T <sub>14</sub>	S <sub>2</sub> T <sub>2</sub> M <sub>2</sub>	70% Sugar conc. + dipping for 42 hrs + solar drying
T <sub>15</sub>	S <sub>2</sub> T <sub>2</sub> M <sub>3</sub>	70% Sugar conc. + dipping for 42 hrs + cabinet drying
T <sub>16</sub>	S <sub>2</sub> T <sub>3</sub> M <sub>1</sub>	70% Sugar conc. + dipping for 48 hrs + sun drying
T <sub>17</sub>	S <sub>2</sub> T <sub>3</sub> M <sub>2</sub>	70% Sugar conc. + dipping for 48 hrs + solar drying
T <sub>18</sub>	S <sub>2</sub> T <sub>3</sub> M <sub>3</sub>	70% Sugar conc. + dipping for 48 hrs + cabinet drying

**Physico-chemical and organoleptic quality of aonla candy****Total soluble solids**

Total soluble solids (TSS) were measured by using hand refractometer (Erma, Japan) and the results were expressed as percent (<sup>0</sup>B) according to standard procedure as given in (Ranganna, 2003) [13]. The refractometer was calibrated with distilled water before use.

**Titrateable acidity**

Titrateable acidity was determined by titrating a known quantity of sample (10ml) against standard solution of 0.1 N Sodium hydroxide to a faint pink colour using phenolphthalein as an indicator. The results were expressed using per cent citric acid (A.O.A.C. 1990) [1].

$$\text{Titrateable acidity \%} = \frac{\text{titer} \times \text{normality of alkali used} \times \text{equivalent wt. of acid}}{\text{volume of sample} \times 100} \times 1000$$

$$\text{Titrateable acidity \%} = \frac{\text{Vol. of aliquot (ml)} \times \text{Wt. or volume of sample (g or ml)}}{\text{sample (g or ml)}} \times 1000$$

**Ascorbic acid**

Determination of ascorbic acid was done by 2, 6 - dichlorophenolindophenol dye method of Johnson (1948) as described by Ranganna (1997). A known quantity of sample was blended with 3 per cent metaphosphoric acid (HPO<sub>3</sub>) to make the final volume of 100 ml and then filtered. A known quantity of aliquot was titrated against 0.025 per cent 2, 6 - dichlorophenolindophenol dye to a pink color end point. The ascorbic acid content of the sample was calculated taking into consideration the dye factor and expressed as mg Ascorbic acid per 100g (Association of Vitamin Chemist, 1966).

$$\text{Ascorbic acid (mg/100g)} = \frac{(\text{Titre} \times \text{Dye factor} \times \text{Volume made up})}{\text{Aliquot of extract taken} \times \text{weight of sample}} \times 100$$

**Tannin**

Total phenols (expressed as tannins) were estimated by the

method of A.O.A.C. (1990) [1]. Reagents 1. Folin Ciocalteu reagents (2N) 1:1. 2. Sodium carbonate 35 g Volume 1 L. 3. Tannic acid 100 mg Volume 1 L Preparation of standard curve Aliquots (0 to 10 ml) of standard tannic acid solution were pipetted into 100 ml volumetric flask and 75 ml of distilled water was added to it. After this, five ml of Folin Ciocalteu reagent and 10 ml of sodium carbonate solution was added. Its volume was then made to 100 ml with water, mixed well and the absorbance was taken after 30 minutes at 760 nm, and standard curve was prepared. Preparation of sample 5 grams of candy powder was boiled for 30 minutes with 400 ml of water, cooled and transferred to a 500 ml volumetric flask and diluted to 500 ml, mixed well and filtered. Estimation One ml of above aliquot was taken and followed the same procedure as for the preparation of standard curve and estimated tannic acid (mg) from the standard curve.

$$\text{Tannins as tannic acid (mg/100g)} = \frac{\text{mg of tannic acid} \times \text{dilution}}{\text{ml of sample taken for colour development} \times \text{weight of sample}} \times 100$$

**Results and Discussion****Effect of various treatments on titrateable acidity of aonla candy during storage**

The data regarding effect of various treatments *viz* concentration of sugar syrup, dipping time and drying methods on titrateable acidity of aonla candy is presented in Table 2. A significant difference was observed in different treatments (S, T and M) during storage of aonla candy.

The data presented in table 2 revealed that combination of sugar solution concentration and dipping time had significant effect on titrateable acidity of aonla candy. The mean values were ranged from 0.17 to 0.35. The minimum value of titrateable acidity (0.12%) was observed in treatment combination S<sub>1</sub>T<sub>1</sub> and S<sub>2</sub>T<sub>2</sub> on 120 days of storage followed by in treatment

combination S<sub>2</sub>T<sub>1</sub> (0.14%), while at initially maximum value (0.40%) was found in S<sub>1</sub>T<sub>2</sub>.

The data presented in table 2 showed that combination of sugar solution concentration and drying methods had significant effect on titrable acidity of aonla candy. The mean values were ranged from 0.13 to 0.40. For treatment combination S<sub>2</sub>M<sub>3</sub> on 120 days of storage showed minimum value (0.13%) followed by S<sub>2</sub>M<sub>2</sub> on 120 days of storage (0.15%), while at initially S<sub>1</sub>M<sub>1</sub> showed maximum value (0.40%).

**Table 2:** Effect of various treatments on titrable acidity (%) of aonla candy during storage

Treatments	Storage period (days)				
	Initial	30	60	90	120
<b>Factor A: Sugar syrup concentration (S)</b>					
S1	0.36	0.32	0.28	0.24	0.19
S2	0.34	0.30	0.24	0.19	0.14
SE	0.003	0.004	0.002	0.003	0.003
CD	0.009	0.012	0.007	0.009	0.009
<b>Factor B: Dipping time (T)</b>					
T1	0.33	0.27	0.22	0.17	0.13
T2	0.34	0.32	0.26	0.22	0.17
T3	0.38	0.34	0.31	0.27	0.21
SE	0.04	0.005	0.003	0.004	0.004
CD	0.01	0.014	0.009	0.011	0.011
<b>Factor C: Drying method (M)</b>					
M1	0.35	0.34	0.29	0.24	0.18
M2	0.33	0.29	0.24	0.20	0.15
M3	0.37	0.31	0.26	0.22	0.17
SE	0.004	0.005	0.003	0.004	0.004
CD	0.011	0.014	0.009	0.011	0.011
<b>Interaction Sx TxM</b>					
S <sub>1</sub> T <sub>1</sub> M <sub>1</sub>	0.32	0.28	0.21	0.15	0.09
S <sub>1</sub> T <sub>1</sub> M <sub>2</sub>	0.25	0.20	0.16	0.11	0.08
S <sub>1</sub> T <sub>1</sub> M <sub>3</sub>	0.38	0.34	0.30	0.26	0.20
S <sub>1</sub> T <sub>2</sub> M <sub>1</sub>	0.44	0.41	0.38	0.31	0.26
S <sub>1</sub> T <sub>2</sub> M <sub>2</sub>	0.38	0.34	0.28	0.24	0.20
S <sub>1</sub> T <sub>2</sub> M <sub>3</sub>	0.38	0.34	0.29	0.27	0.23
S <sub>1</sub> T <sub>3</sub> M <sub>1</sub>	0.34	0.42	0.39	0.36	0.30
S <sub>1</sub> T <sub>3</sub> M <sub>2</sub>	0.38	0.32	0.30	0.27	0.20
S <sub>1</sub> T <sub>3</sub> M <sub>3</sub>	0.32	0.29	0.27	0.27	0.22
S <sub>2</sub> T <sub>1</sub> M <sub>1</sub>	0.35	0.34	0.30	0.24	0.19
S <sub>2</sub> T <sub>1</sub> M <sub>2</sub>	0.32	0.27	0.21	0.18	0.14
S <sub>2</sub> T <sub>1</sub> M <sub>3</sub>	0.38	0.20	0.16	0.11	0.09
S <sub>2</sub> T <sub>2</sub> M <sub>1</sub>	0.25	0.28	0.21	0.18	0.12
S <sub>2</sub> T <sub>2</sub> M <sub>2</sub>	0.32	0.27	0.21	0.17	0.14
S <sub>2</sub> T <sub>2</sub> M <sub>3</sub>	0.32	0.28	0.20	0.16	0.11
S <sub>2</sub> T <sub>3</sub> M <sub>1</sub>	0.32	0.31	0.28	0.21	0.16
S <sub>2</sub> T <sub>3</sub> M <sub>2</sub>	0.38	0.34	0.30	0.24	0.18
S <sub>2</sub> T <sub>3</sub> M <sub>3</sub>	0.44	0.41	0.34	0.28	0.21
SE	0.010	0.0132	0.0080	0.001	0.010
CD @ 5%	0.028	0.036	0.022	0.027	0.028

The data presented in table 2 revealed that, combination of

dipping time and drying methods had significant effect on titrable acidity at initially and at 60 days while there is non-significant effect at 30, 90 and 120 days. The mean values were ranged from 0.17 to 0.35. The minimum value (0.11%) was found in treatment combination T<sub>1</sub>M<sub>2</sub> on 120 days of storage followed by T<sub>1</sub>M<sub>1</sub> on 120 days of storage (0.14%), while at initially maximum value (0.38%) was observed in treatment combination. T<sub>1</sub>M<sub>3</sub>.

The data presented in table 2 showed that, combination of sugar syrup concentration, dipping time and drying methods had significant effect on titrable acidity. The mean values were ranged from 0.17 to 0.34%. For treatment combination S<sub>1</sub>T<sub>1</sub>M<sub>2</sub> on 120 days of storage showed minimum value (0.08%) followed by S<sub>1</sub>T<sub>1</sub>M<sub>1</sub> on 120 days of storage (0.09%), while at initially S<sub>2</sub>T<sub>3</sub>M<sub>3</sub> showed maximum value (0.44%).

Similar results have also been reported by Krishnaveni *et al.*, (2001). Nayak *et al.* (2011) [12], Verma *et al.*, (2006) [22] and Kumar *et al.*, (2015) [15] reported similar observations in aonla segments in syrup, Darunj peel candy and papaya leather respectively. For treatment combination S<sub>1</sub>T<sub>1</sub>M<sub>2</sub> on 120 days of storage showed minimum value (0.08%) followed by S<sub>1</sub>T<sub>1</sub>M<sub>1</sub> on 120 days of storage (0.09%), while S<sub>2</sub>T<sub>3</sub>M<sub>3</sub> on 0 day of storage showed maximum value (0.44%). During storage period (120 days) candy showed significant decrease in titrable acidity content and the mean value of treatments decreased from 0.34 to 0.17.

#### Effect of various treatments on ascorbic acid of aonla candy during storage

The data revealed that, combination of sugar solution concentration and dipping time had significant effect on ascorbic acid of aonla candy. The mean values were ranged from 246.75 to 271.08 mg/100g. For treatment combination S<sub>2</sub>T<sub>3</sub> at initial of storage showed maximum value (320.0 mg/100g) followed by S<sub>1</sub>T<sub>3</sub> on 30 days of storage (313.6mg/100g), while S<sub>1</sub>T<sub>2</sub> on 120 days of storage showed minimum value (164.6 mg/100g).

The data presented in table 3 showed that, combination of sugar solution concentration and drying methods had significant effect on ascorbic acid of aonla candy. The mean values were ranged from 246.75 to 271.08mg/100g. For treatment combination S<sub>2</sub>M<sub>2</sub> at initial day of storage showed maximum value (320.0mg/100g) followed by S<sub>2</sub>M<sub>2</sub> on 60 days of storage (315.3 mg/100g), while S<sub>1</sub>M<sub>1</sub> on 120 days of storage showed minimum value (196.6mg/100g).

The data presented in table 3 showed that, combination of dipping time and drying methods had significant effect on ascorbic acid of aonla candy. The mean values were ranged from 246.77 to 271.11mg/100g. For treatment combination T<sub>3</sub>M<sub>3</sub> at initial day of storage showed maximum value (400.0mg/100g) followed by T<sub>3</sub>M<sub>3</sub> on 30 days of storage (391.0mg/100g), while T<sub>1</sub>M<sub>1</sub> on 120 days of storage showed minimum value (169.5mg/100g).

**Table 3:** Effect of various treatments on ascorbic acid (mg/100g) content of aonla candy during storage

Treatments	Storage period (days)				
	Initial	30	60	90	120
<b>Factor A: Sugar syrup concentration (S)</b>					
S1	248.89	242.78	238.22	232.67	226.11
S2	293.33	287.67	283.00	276.67	267.44
SE	2.48	4.63	4.77	4.64	4.46
CD	6.87	12.8	13.21	12.85	12.35
<b>Factor B: Dipping time (T)</b>					
T1	253.33	248.50	243.00	237.17	228.83
T2	240.00	234.00	231.00	226.50	219.00

T3	320.00	313.17	307.83	300.33	292.50
SE	3.04	56.76	5.84	5.68	5.46
CD	8.42	15.70	16.18	15.74	15.13
<b>Factor C: Drying methods (M)</b>					
M1	240.0	233.83	228.17	221.17	213.33
M2	266.6	262.17	259.50	253.67	246.33
M3	306.6	299.67	294.17	289.17	280.67
SE	3.04	5.67	5.84	5.68	5.46
CD	8.42	15.70	16.18	15.74	15.13
<b>Interaction SxTxM</b>					
S <sub>1</sub> T <sub>1</sub> M <sub>1</sub>	160.0	155.0	151.0	145.0	139.0
S <sub>1</sub> T <sub>1</sub> M <sub>2</sub>	240.0	236.0	230.0	226.0	220.0
S <sub>1</sub> T <sub>1</sub> M <sub>3</sub>	320.0	312.0	309.0	304.0	294.0
S <sub>1</sub> T <sub>2</sub> M <sub>1</sub>	240.0	232.0	226.0	221.0	216.0
S <sub>1</sub> T <sub>2</sub> M <sub>2</sub>	160.0	155.0	150.0	146.0	140.0
S <sub>1</sub> T <sub>2</sub> M <sub>3</sub>	160.0	154.0	151.0	145.0	138.0
S <sub>1</sub> T <sub>3</sub> M <sub>1</sub>	240.0	235.0	230.0	224.0	220.0
S <sub>1</sub> T <sub>3</sub> M <sub>2</sub>	240.0	236.0	231.0	224.0	218.0
S <sub>1</sub> T <sub>3</sub> M <sub>3</sub>	480.0	470.0	466.0	459.0	450.0
S <sub>2</sub> T <sub>1</sub> M <sub>1</sub>	240.0	232.0	222.0	215.0	200.0
S <sub>2</sub> T <sub>1</sub> M <sub>2</sub>	320.0	322.0	318.0	314.0	309.0
S <sub>2</sub> T <sub>1</sub> M <sub>3</sub>	240.0	234.0	228.0	219.0	211.0
S <sub>2</sub> T <sub>2</sub> M <sub>1</sub>	240.0	235.0	229.0	221.0	216.0
S <sub>2</sub> T <sub>2</sub> M <sub>2</sub>	320.0	312.0	319.0	312.0	300.0
S <sub>2</sub> T <sub>2</sub> M <sub>3</sub>	320.0	316.0	311.0	314.0	304.0
S <sub>2</sub> T <sub>3</sub> M <sub>1</sub>	320.0	314.0	311.0	301.0	289.0
S <sub>2</sub> T <sub>3</sub> M <sub>2</sub>	320.0	312.0	309.0	300.0	291.0
S <sub>2</sub> T <sub>3</sub> M <sub>3</sub>	320.0	312.0	300.0	294.0	287.0
SE	7.45	1.39	14.3	13.9	13.3
CD @ 5%	20.6	38.4	39.6	38.5	37.0

The data reveals that combination of sugar syrup concentration, dipping time and drying methods had significant effect on ascorbic acid. The mean values of ascorbic acid content were ranged from 246.77 to 271.11mg/100g. For treatment combination S<sub>1</sub>T<sub>3</sub>M<sub>3</sub> at initial day of storage showed maximum value (480.0mg/100g) followed by S<sub>1</sub>T<sub>3</sub>M<sub>3</sub> on 30days of storage (470.0mg/100g), while S<sub>1</sub>T<sub>2</sub>M<sub>3</sub> on 120 days of storage showed minimum value (138.0mg/100g).

Rani and Bhatia (1986) <sup>[19]</sup> and Ayub *et al.*, (2005) also reported a declining trend in ascorbic acid in intermediate moisture baguighosa and sweetened guava slices, respectively during storage. A declining trend in ascorbic acid content of aonla preserve (Tripathi *et al.*, 1988) <sup>[21]</sup>, ber candy (Kaikadi *et al.*, 2006), karonda candy (Manivasagan *et al.*, 2006), Sethi (1980) in aonla preserve, Rani and Bhatia (1985) in pear candy, Kumar and Singh (2001) <sup>[7]</sup> Rani and Bhatia (1986) <sup>[19]</sup> and Ayub *et al.*, (2005) also reported a declining trend in ascorbic acid in intermediate moisture baguighosa and sweetened guava slices, respectively during storage. Similar results were also reported from Priya and Khatkar (2013) <sup>[14, 17]</sup> during study on effect of processing methods on keeping quality of aonla preserve, Nayak *et al.*, (2012) <sup>[11]</sup> during study on changes in nutritional and organoleptic quality of flavored candy prepared from aonla during storage.

#### Effect of various treatments on total soluble solids (%) of aonla candy during storage.

The data presented in table 4 revealed that combination of

sugar solution concentration and dipping time had significant effect on TSS at initially and 30 days of storage of aonla candy, while there was non-significant effect at 60, 90 and 120 days of storage. The mean values were ranged from 83.2 to 85.1%. The maximum value (85.5%) was recorded in treatment combination S<sub>1</sub>T<sub>2</sub> at 120days of storage followed by in treatment S<sub>1</sub>T<sub>1</sub> (85.4%), while at 30days of storage of aonla candy, minimum value (82.6%) was recorded in treatment S<sub>2</sub>T<sub>1</sub>.

The data presented in table 4 revealed that combination of sugar solution concentration and drying methods on 0, 30, 90 and 120 days of storage period had significant effect on TSS while non-significant effect was observed at 60 days of storage of aonla candy. The mean values were ranged from 83.18 to 83.85%. The maximum value (84.4%) of TSS was recorded in treatment combination S<sub>1</sub>M<sub>1</sub> on 120 days of storage of aonla candy followed by in treatment S<sub>1</sub>M<sub>3</sub> (83.3%), while at initially minimum value (82.3%) of TSS was recorded in treatment S<sub>1</sub>M<sub>3</sub>.

The data presented in table 4 showed that combination of dipping time and drying methods had significant effect on TSS at 30 days of storage, while non-significant effect was observed on 0, 60, 90 and 120 days of storage of aonla candy. The mean values were ranged from 83.20 to 85.8%. At 120day of storage of aonla candy, maximum value (85.8%) of TSS was observed in treatment combination T<sub>2</sub>M<sub>1</sub> followed by T<sub>2</sub>M<sub>3</sub> (85.1%), while minimum value (82.9%) was observed in treatment T<sub>1</sub>M<sub>1</sub> at 30days of storage of aonla candy.

**Table 4:** Effect of various treatments on total soluble solids (%) of aonla candy during storage

Treatments	Storage period (days)				
	Initial		Initial		Initial
<b>Factor A: Sugar syrup concentration (S)</b>					
S1	83.0	83.34	83.96	84.61	85.33
S2	83.4	83.12	83.71	84.36	84.91
SE	0.107	0.065	0.32	0.16	0.11
CD	0.297	0.180	NS	NS	0.31
<b>Factor B: Dipping time (T)</b>					
T1	83.16	83.05	83.73	84.35	85.01
T2	83.50	83.51	84.10	84.78	85.31
T3	83.00	83.13	83.68	84.33	85.03
SE	0.13	0.079	0.40	0.19	0.14
CD	0.36	0.22	NS	NS	NS
<b>Factor C: Drying methods (M)</b>					
M1	83.33	83.50	83.98	84.73	85.31
M2	83.33	83.28	83.86	84.45	85.05
M3	83.0	82.91	83.66	84.28	85.00
SE	0.13	0.079	0.40	0.19	0.14
CD	NS	0.22	NS	NS	NS
<b>Interaction SxTxM</b>					
S <sub>1</sub> T <sub>1</sub> M <sub>1</sub>	83.0	83.6	84.1	84.9	85.4
S <sub>1</sub> T <sub>1</sub> M <sub>2</sub>	84.0	84.2	84.9	85.3	85.9
S <sub>1</sub> T <sub>1</sub> M <sub>3</sub>	82.0	82.6	83.4	84.1	85.0
S <sub>1</sub> T <sub>2</sub> M <sub>1</sub>	84.0	84.5	85.0	85.9	86.4
S <sub>1</sub> T <sub>2</sub> M <sub>2</sub>	83.0	83.7	84.0	84.6	85.0
S <sub>1</sub> T <sub>2</sub> M <sub>3</sub>	82.0	82.9	83.6	84.2	85.2
S <sub>1</sub> T <sub>3</sub> M <sub>1</sub>	83.0	83.7	84.1	84.9	85.6
S <sub>1</sub> T <sub>3</sub> M <sub>2</sub>	83.0	82.9	83.6	84.1	85.4
S <sub>1</sub> T <sub>3</sub> M <sub>3</sub>	83.0	82.0	83.0	83.5	84.1
S <sub>2</sub> T <sub>1</sub> M <sub>1</sub>	83.0	82.2	83.0	83.9	84.6
S <sub>2</sub> T <sub>1</sub> M <sub>2</sub>	83.0	82.3	83.1	83.7	84.2
S <sub>2</sub> T <sub>1</sub> M <sub>3</sub>	84.0	83.4	83.9	84.2	85.0
S <sub>2</sub> T <sub>2</sub> M <sub>1</sub>	84.0	83.6	84.0	84.6	85.3
S <sub>2</sub> T <sub>2</sub> M <sub>2</sub>	84.0	83.4	84.0	84.7	84.9
S <sub>2</sub> T <sub>2</sub> M <sub>3</sub>	84.0	83.0	84.0	84.7	85.1
S <sub>2</sub> T <sub>3</sub> M <sub>1</sub>	83.0	83.4	83.7	84.2	84.6
S <sub>2</sub> T <sub>3</sub> M <sub>2</sub>	83.0	83.2	83.6	84.3	84.9
S <sub>2</sub> T <sub>3</sub> M <sub>3</sub>	83.0	83.6	84.1	85.0	85.6
SE	0.32	0.19	0.98	0.48	0.34
CD @ 5%	0.89	0.54	NS	NS	NS

The data presented in table 4 that combination of sugar syrup concentration, dipping time and drying methods had significant effect on TSS at 0 and 30 days of storage of aonla candy and non-significant effect was observed at 60, 90 and 120 days of storage. The mean values was ranged from 83.22 to 85.12%. At 120days of storage of aonla candy, maximum value (86.4%) of TSS was found in treatment combination S<sub>1</sub>T<sub>2</sub>M<sub>1</sub> followed by in treatment S<sub>1</sub>T<sub>1</sub>M<sub>2</sub> (85.9%), while at 30 days of storage of aonla candy, minimum value (82.0%) of TSS was found in treatment S<sub>1</sub>T<sub>3</sub>M<sub>3</sub>.

The stored IM Baguighosa suffered loss of moisture, which correspondingly increased TSS in the product (Rani and Bhatia, 1986)<sup>[19]</sup>. Ayub *et al.*, (2005) also showed an increase in TSS of intermediate moisture sweetened guava slices during storage. The data reveals that combination of sugar syrup concentration, dipping time and drying methods had significant effect on TSS on 0 and 30 days of storage and non-significant effect was observed on 60, 90 and 120 days of storage. The mean values ranged from 83.22 to 85.12. For treatment combination S<sub>1</sub>T<sub>2</sub>M<sub>1</sub> on 120 days of storage showed maximum value (86.4) followed by S<sub>1</sub>T<sub>1</sub>M<sub>2</sub> on 120 days of storage (85.9), while S<sub>1</sub>T<sub>3</sub>M<sub>3</sub> on 30 days of storage showed minimum value (82.0). Similar results were also found by Kumar *et al.* (2015)<sup>[15]</sup> during study on development of value added product from citrus peel and Aggrawal and Michael

(2014)<sup>[3]</sup> during study on effect of replacing sucrose with fructose on the physico-chemical sensory characteristics of kinnow candy.

#### Effect of various treatments on tannin content of aonla candy during storage

The data presented in table 5 revealed that combination of sugar solution concentration and dipping time had significant effect on tannin at 30 and 60 days of storage of aonla candy, while there is non-significant effect at 0, 90 and 120 days of storage of aonla candy. The mean values were ranged from 0.37 to 1.15mg/100g. At 120 days of storage of aonla candy lowest tannin (0.27mg/100g) content was found in treatment combination S<sub>2</sub>T<sub>3</sub> followed in treatment by S<sub>2</sub>T<sub>2</sub> (0.33mg/100g), while at initially highest tannin content (1.23mg/100g) was found in treatment combination S<sub>2</sub>T<sub>1</sub>.

The data presented in table 5 showed that combination of sugar solution concentration and drying methods at 0, 30, 90 and 120 days of storage of aonla candy had non-significant effect on tannin content while significant effect was observed at 60 days of storage of aonla candy. The mean values were ranged from 0.37 to 1.15mg/100g. The highest tannin content (0.30mg/100g) was recorded in treatment combination S<sub>2</sub>M<sub>1</sub> at 120 days of storage of aonla candy followed by in treatment combination S<sub>2</sub>M<sub>2</sub> (0.33mg/100g), while at initially,

the highest tannin content (1.19mg/100g) was recorded in treatment combination S<sub>2</sub>M<sub>3</sub>.

In interaction effect of dipping time and drying methods on tannin content had significant at 90 days of storage of aonla candy, while non-significant effect was observed at 0, 30, 60 and 120 days of storage of aonla candy. The mean values were ranged from 0.38 to 1.15mg/100g. At 120 day of storage the highest tannin content (0.31mg/100g) was observed in treatment combination T<sub>3</sub>M<sub>2</sub> followed by in treatment T<sub>2</sub>M<sub>3</sub> (0.33mg/100g), while at initially the highest tannin content was observed in treatment T<sub>1</sub>M<sub>1</sub> (1.22mg/100g).

**Table 5:** Effect of various treatments on tannin (mg/100g) content of aonla candy during storage

Treatments	Storage period (days)				
	Initial		Initial		Initial
<b>Factor A: Sugar syrup concentration (S)</b>					
S1	1.13	0.98	0.89	0.64	0.43
S2	1.17	0.92	0.73	0.48	0.33
SE	0.015	0.019	0.008	0.020	0.018
CD	NS	0.053	0.022	0.056	0.049
<b>Factor B: Dipping time (T)</b>					
T1	1.21	0.94	0.82	0.59	0.44
T2	1.15	1.03	0.84	0.60	0.35
T3	1.09	0.89	0.77	0.49	0.35
SE	0.018	0.023	0.010	0.025	0.022
CD	0.051	0.065	0.027	0.069	0.061
<b>Factor C: Drying methods (M)</b>					
M1	1.14	0.96	0.80	0.62	0.37
M2	1.14	0.95	0.82	0.52	0.38
M3	1.16	0.94	0.80	0.53	0.38
SE	0.018	0.023	0.010	0.025	0.022
CD	NS	NS	NS	0.069	NS
<b>Interaction S<sub>i</sub>T<sub>j</sub>M<sub>k</sub></b>					
S <sub>1</sub> T <sub>1</sub> M <sub>1</sub>	1.20	0.09	0.86	0.65	0.48
S <sub>1</sub> T <sub>1</sub> M <sub>2</sub>	1.19	0.95	0.85	0.66	0.57
S <sub>1</sub> T <sub>1</sub> M <sub>3</sub>	1.20	0.93	0.90	0.78	0.43
S <sub>1</sub> T <sub>2</sub> M <sub>1</sub>	1.13	1.10	0.92	0.84	0.38
S <sub>1</sub> T <sub>2</sub> M <sub>2</sub>	1.12	1.10	0.91	0.65	0.39
S <sub>1</sub> T <sub>2</sub> M <sub>3</sub>	1.15	1.12	0.97	0.61	0.34
S <sub>1</sub> T <sub>3</sub> M <sub>1</sub>	1.05	0.91	0.87	0.60	0.47
S <sub>1</sub> T <sub>3</sub> M <sub>2</sub>	1.08	0.92	0.86	0.53	0.35
S <sub>1</sub> T <sub>3</sub> M <sub>3</sub>	1.09	0.94	0.88	0.46	0.46
S <sub>2</sub> T <sub>1</sub> M <sub>1</sub>	1.24	1.00	0.76	0.51	0.33
S <sub>2</sub> T <sub>1</sub> M <sub>2</sub>	1.20	0.91	0.86	0.48	0.39
S <sub>2</sub> T <sub>1</sub> M <sub>3</sub>	1.25	0.95	0.70	0.50	0.47
S <sub>2</sub> T <sub>2</sub> M <sub>1</sub>	1.17	0.99	0.78	0.67	0.33
S <sub>2</sub> T <sub>2</sub> M <sub>2</sub>	1.18	0.97	0.75	0.42	0.34
S <sub>2</sub> T <sub>2</sub> M <sub>3</sub>	1.20	0.90	0.72	0.41	0.33
S <sub>2</sub> T <sub>3</sub> M <sub>1</sub>	1.10	0.89	0.64	0.50	0.25
S <sub>2</sub> T <sub>3</sub> M <sub>2</sub>	1.11	0.86	0.69	0.43	0.28
S <sub>2</sub> T <sub>3</sub> M <sub>3</sub>	1.12	0.85	0.68	0.42	0.30
SE	0.045	0.058	0.024	0.061	0.054
CD @ 5%	NS	NS	NS	NS	NS

In interaction effect of sugar syrup concentration, dipping time and drying methods on tannin content during storage of aonla candy had non-significant. The mean values were ranged from 0.38 to 1.15mg/100g. At 120 days of storage of aonla candy the highest tannin content was recorded in treatment combination S<sub>2</sub>T<sub>3</sub>M<sub>1</sub> (0.25mg/100g) followed by in treatment S<sub>1</sub>T<sub>3</sub>M<sub>2</sub> (0.28mg/100g), while at initially the highest tannin content was recorded in treatment combination S<sub>1</sub>T<sub>1</sub>M<sub>1</sub> (1.24mg/100g).

Similar results were reported by Priya and Khatkar (2013)<sup>[14, 17]</sup> in aonla preserves and by Nayak *et al.*, (2012)<sup>[11]</sup> during

study on changes of nutritional and organoleptic quality of flavoured candy prepared from aonla during storage. Candy showed significant decrease in tannins during storage period and the mean value decreased from 1.15 to 0.38. For treatment combination S<sub>2</sub>T<sub>3</sub>M<sub>1</sub> on 120 days of storage showed minimum value (0.25) followed by S<sub>1</sub>T<sub>3</sub>M<sub>2</sub> on 120 days of storage (0.28), while S<sub>1</sub>T<sub>1</sub>M<sub>1</sub> on 0 day of storage showed maximum value (1.24).

## References

1. AOAC. Official Methods of Analysis. Association of Official Analytical Chemists. Washington, D.C. 15th edition, 1990.
2. Agarwal S, Chopra CS. Studies on changes in ascorbic acid and total phenol in making aonla products. Beverage & Food World. 2004; 31(5):32-34.
3. Aggarwal P, Michael M. Effect of Replacing Sucrose with Fructose on the Physico-chemical Sensory Characteristics of Kinnow Candy. Czech J Food Sci. 2014; 32(2):158-163.
4. Alam MS, Singh A. Optimum process parameters for development of sweet aonla flakes. International Journal Research and Reviews Applied Sciences. 2010; 3(3):323-333.
5. Ganachari A, Thangavel K, Mazara Ali S, Nidoni U, Ananthacharya. Physical properties of Aonla fruit relevant to the design of processing equipments. International Journal of Engineering Science and Technology. 2010; 2(12):7562-7566. 2010; 1(4):419-425.
6. Kalra CL. The chemistry and technology of aonla a resume, Indian Food Packer. 1988; 38(4):67.
7. Kumar S, Singh IS. Storage studies of aonla fruit products at ambient temperature. Progressive Horticulturae. 2001; 33(2):169-173.
8. Kumar S, Kumar K, Prakash C. Effect of sugar and jaggery on the quality characteristics of papaya leather and shelf life stability at room temperature. South Asian J Food Technol. Environ. 2015; 1(1):79-85.
9. Mishra S, Verma A, Prasad VM, Sheikh S. Development of value added amla candy with rose extract. The Allahabad Farmer. 2011; 66(2):134-136.
10. Mishra V, Mishra P, Rai GK. Process and product standardization for the development of amla bar. Beverage & Food World. 2010; 34(6):58-60. Nhb.gov.in/area%20\_production.html.
11. Nayak P, Tondon DK, Bhatt DK. Study on changes of nutritional and organoleptic quality of flavored candy prepared from aonla (*Emblica officinalis* G) during storage, International Journal of Nutrition and Metabolism. 2012; 4(7):100-106.
12. Nayak P, Tondon DK. Standardization of pretreatment for preparation of aonla segments in syrup. National seminar on production and processing of aonla (*Emblica officinalis* G) Amdavad – Gujrat. 2006; 35:21-23.
13. Pathak RK, Pandey D, Haseeb M, Tondon DK. The Aonla, Technical Bulletin, Lucknow, India, 2003.
14. Priya MD, Khatkar BS. Effect of processing methods on keeping quality of aonla (*Emblica officinalis* Gaertn.) Preserve. International Food Research Journal. 2013; 20(2):617-622.
15. Pareek S, Kaushik RA. Effect of drying methods on quality of Indian gooseberry (*Emblica officinalis* Gaertn.) powder during storage. Journal of Scientific and Industrial Research. 2012; 71(11):727-732.

16. Pokharkar SM. Development and performance evaluation of aonla shredding machine. Beverage & Food World. 2005; 32(3):52-53.
17. Priya MD, Khatkar BS. Effect of processing methods on keeping quality of aonla (*Emblica officinalis* Gaertn.) preserve. International Food Research Journal. 2013; 20(2):617-622.
18. Rajkumar NV, Theres M, Kuttan R. Emblica officinalis fruits afford protection against experimental gastric ulcers in rats. Pharmaceutical Biology. 2001; 39(5):375-380.
19. Rani U, Bhatia BS. Studies on bagugosha and pear for preserves and a ready to eat product. Indian Food Packer. 1986; 40(3):25-31.
20. Sahu GD, Singh P, Singh AK. Studies on the physico-chemical changes in Aonla preserve (Murabba) of three cultivars during storage. Research Journal of Agriculture Science, 2010.
21. Tripathi VK, Singh MB, Singh S. Studies on comparative compositional changes in different preserved product of aonla (*Emblica officinalis* Gaertn.) var. Banarasi. Indian Food Packer. 1988; 42(4):60-66.
22. Verma S, Srivastava PK, Durrani A. Organoleptic characteristics of honey based amlamurabba. 40th ISAE Annual Convention, Tamil Nadu Agricultural University, Coimbatore, 2006.