

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2019; 8(4): 1581-1585 Received: 13-05-2019 Accepted: 17-06-2019

Gayatri Khangjarakpam

ICAR - Research Complex for NEH Region, Lamphelpat, Imphal, Manipur, India

Leishangthem Jeebit Singh

Directorate of Horticulture and Soil Conservation, Manipur, India

Soumen Maitra

Department of Floriculture, Medicinal & Aromatic Plants, Faculty of Horticulture, UBKV, Behar, West Bengal, India

Somnath Mandal

Department of Biochemistry, Faculty of Agriculture, UBKV, Behar, West Bengal, India

Correspondence Gayatri Khangjarakpam ICAR - Research Complex for NEH Region, Lamphelpat, Imphal, Manipur, India

Influence of foliar application of Gibberellic acid on growth, development, yield and biochemical constituents of African marigold cv. 'Pusa Narangi Gainda'

Gayatri Khangjarakpam, Leishangthem Jeebit Singh, Soumen Maitra and Somnath Mandal

Abstract

An experiment was carried out on African marigold cv. 'Pusa Narangi Gainda' to study the influence of foliar application of Gibberellic acid (GA₃) on growth, development, yield and biochemical constituents at UBKV, West Bengal. The treatments consist of ten different concentrations of GA₃ with a control. GA₃ treatment at 250 ppm resulted in significantly better performance in relation to plant height (87.87cm), number of branches (13.37), number of leaves (36.01), plant spread (40.92 cm), whole biomass (447.10 g), fresh shoot weight (355.69 g), fresh root weight (92.10 g), days to flower bud initiation (52.93 days), flower longevity on plants (6.67 days), duration of flowering (24.4 days), number of flowers/ plant (53.12), diameter of flowers (6.79 cm) and expected flower yield (52.28 MT/ha). The same treatment was also effective in improving biochemical constituents like leaf chlorophyll content (54.60 SPAD), protein content (1390.5 μ g/g), carotenoids (370.8 mg/Kg), carbohydrate (288.9 μ g/g), reducing sugar content (146.1 μ g/g) and non-reducing sugar content (142.8 μ g/g) in petals.

Keywords: African marigold, Pusa Narangi Gainda, Gibberellic acid, foliar application

Introduction

African marigold (*Tagetes erecta*) belonging to the family Asteraceae is a flowering ornamental annual with profuse branching and flowering habit. Some of the major problems associated with cultivation of this crop are apical dominance (Sunitha *et al.*, 2007) ^[1], delay in flowering (Sharma *et al.*, 2006) ^[2] and development of long and weak stems (Gawle *et al.*, 2012) ^[3]. These in turn resulted in poor yield/economic returns. However, provision of ambient climatic conditions through selection of site, judicious cultural operations, balanced nutrition and physiological manipulations by pinching or application of plant growth regulators may improve the yield of the crop.

The African marigold cultivar 'Pusa Narangi Gainda' was developed by pedigree hybridization by crossing between two exotic varieties 'Cracker Jack' x 'Golden Jubilee'. The plant of this variety is of medium stature, vigorous and uniform with dark green foliage which grows to a height of 80-85 cm. It flowers in 125-136 days after sowing. It is most suitable for garland making, religious offerings and carotenoid extraction (Kanwar and Khandelwal, 2014)^[4].

Now-a-days, application of plant growth regulators is gaining its popularity in various crop cultivation practices. Plant growth regulators are the organic chemical compounds which influence the plant growth, crop yield and flower quality by modification or regulating the plant physiological processes when applied in very minute quantity (Iqbal *et al.*, 2011; Fernández *et al.*, 2011)^{[5] [6]}.

Therefore, an experiment was conducted to study the effect of foliar application of gibberellic acid on growth, development, yield and biochemical constituents of African Marigold cv. 'Pusa Narangi Gainda' in order to overcome the various inherent problems in cultivation.

Materials and Methods

The experiment was conducted on African marigold cv. 'Pusa Narangi Gainda' during winter season of 2014-2015 and 2015-2016 at Uttar Banga Krishi Viswavidyalaya, Pundibari, West Bengal. The treatments consisted of ten different doses of gibberellic acid (GA₃) at 50 ppm, 100 ppm, 150 ppm, 200 ppm, 250 ppm, 300ppm, 350 ppm, 400 ppm, 450 ppm and 500 ppm with a control. The experimental design followed was Randomised Block Design (RBD) with three replications per treatments. Each plot was planted with 40 plants at a spacing of 40 cm x

40 cm. Sowing was done on first fortnight of December and transplanting was carried out 30 days after sowing and standard cultural practices like hoeing, irrigation, weeding and fertilizer application were followed as per the requirement of the crop. Each treatment was applied twice at 3 weeks and 6 weeks after transplanting. Ten plants were selected randomly and tagged for different treatments to record various vegetative and reproductive parameters. Chlorophyll content of leaves was estimated using chlorophyll meter SPAD 502 at 10:00 hours. Estimation of protein content of flower petals was carried out as per Lowry et al., (1951)^[7]. Carotenoid content of the petals was estimated by spectrophotometric method as devised by Ranganna (2002)^[8]. Estimation of total carbohydrate content $(\mu g/g)$ of the flower petal was carried out as per Hodge et al., (1962)^[9], reducing sugar content (μ g/g) and non-reducing sugar content (μ g/g) of the flower petal as per Somogyi (1952)^[10].

Data on various growth, flowering, yield and biochemical parameters were recorded and statistically analysed by applying the technique of analysis of variance. Data was analysed within the ANOVA of the SPSS software program, version 6.0 and level of significance for t-test was kept at 5% (P=0.05).

Results and Discussion

Vegetative growth parameters

Among different concentrations of foliar application of GA_3 on African marigold cv. 'Pusa Narangi Gainda', the treatment 250 ppm $GA_3(T_5)$ resulted in significantly maximum plant

height (87.87cm), number of branches (13.37), number of leaves per plant (36.01) and plant spread (40.92 cm) as evident from Table 1. The effect of gibberellins in increasing various vegetative parameters like plant height, number of branches, plant spread and number of leaves per plantmay be due to increased auxin level of tissues or enhance conversion of tryptophane to IAA which results inenhanced cell division, cell enlargement, promotion of protein synthesis and diversion of plant metabolites from vertical growth to horizontal growth (Nelissen et al., 2012)^[11]. Similar results of enhanced vegetative growth due to the effect of GA₃ have already been reported by Dhekney et al., (2000) ^[12] in rose; Kumar et al., (2003) ^[13] in China aster; Mohariya et al., (2003)^[14] in chrysanthemum; and Ramdevputra (2009)^[15] in marigold. The higher concentrations of exogenously applied GA₃ were found to reduce the vegetative parameters under study as GA₃ worked with its full potential upto certain the optimum concentration and feedback inhibition occurred beyond such concentrations.

The plants treated with 250 ppm GA₃ (T₅) also resulted in significantly maximum gain in whole biomass (447.10 g), fresh shoot weight (355.69g) and fresh root weight (92.10 g) compared to other treatments as shown in Table 1. Application of GA₃ had been found to increase the photosynthetic rate in leaves of plant through changes in plastid development and chloroplast structure (Jiang *et al.*, 2012) ^[16] which resulted in increase in whole biomass, fresh shoot and root weight.

 Table 1: Influence of different treatments of Gibberellic acid on various vegetative parameters of African marigold cv. 'Pusa Narangi Gainda' (data are mean of two season)

Treatments	Plant height (cm)	Number of main branches	Number of leaves	Plant spread (cm)	Biomass per plant (g)	Fresh weight of shoot (g)	Fresh weight of root(g)
T ₀ (control)	51.73	7.40	20.07	30.42	224.43	197.04	27.39
T ₁ (GA ₃ @ 50ppm)	80.38	9.93	30.20	33.48	310.94	264.18	46.76
T ₂ (GA ₃ @ 100ppm)	82.32	10.00	32.07	34.12	318.71	266.04	52.68
T ₃ (GA ₃ @ 150ppm)	83.96	11.27	32.80	34.76	363.09	299.83	63.26
T ₄ (GA ₃ @ 200ppm)	84.43	12.43	33.00	36.30	414.68	330.69	83.99
T5 (GA3@ 250ppm)	87.87	13.37	36.01	40.92	447.10	355.69	92.10
T ₆ (GA ₃ @ 300ppm)	83.73	11.33	32.87	39.43	376.48	301.42	75.05
T7 (GA3@ 350ppm)	82.37	10.80	31.54	35.58	347.37	287.32	60.05
T ₈ (GA ₃ @ 400ppm)	81.37	10.33	31.26	34.33	327.86	274.82	53.04
T9 (GA3@ 450ppm)	80.53	10.47	29.98	33.53	329.24	278.54	50.69
T ₁₀ (GA ₃ @ 500ppm)	76.21	10.01	27.51	33.18	308.65	266.30	42.34
S. Em (±)	1.392	0.174	0.487	0.614	5.625	4.503	1.056
C. D. at 5%	4.056	0.544	1.564	1.790	17.551	14.474	3.077

Flowering parameters

Among the foliar application of different doses, treatment with 250 ppm GA₃also resulted in minimum days to flower bud initiation (52.93 days after transplanting), maximum flower longevity on plant (6.67 days) and maximum duration of flowering (24.3 days) as presented in Table 2. Earliest flower bud initiation and with maximum flowering duration observed with the application of 250 ppm GA₃ was probably due to the reduction in juvenile period through reduction of Sphase in the interphase of cell cycle inducing the shoot apical meristem instead of producing leaves and branches to start producing buds.GA₃ resulted earliness in flowering and improvement in qualitative characters of flowers like early maturity, increased duration of flowering (Dutta and Seemanthini, 1998)^[17]. Increase in duration of flowering might be due to advanced bud formation and stimulating flowering in GA₃ treated plants are in conformity with the findings of Kumar et al., (2010) [18]. Similar results were observed by Nair *et al.*, $(2002)^{[19]}$ in gerbera and Porwal *et al.*, $(2002)^{[20]}$ in Damask Rose. Kumar *et al.*, $(2009)^{[21]}$ also concluded that GA₃ at 200 ppm caused earliest appearance of initial spike and opening of first floret in tuberose.

Plants treated with GA₃ at 250 ppm (T₅) were effective to produce maximum number of flowers per plant (53.12) and maximum diameter of flowers (6.79 cm). The same treatment was also effective to produce maximum expected flower yield (52.28 MT/ha) while control produce a yield of 23.30 MT/ha. Increased loose flower yield in plants treated with GA₃ 250 ppm may be probably due to the production of flowers with greater diameter and increased fresh weight. Greater diameter also might be induced through increased number of florets as a result of better nutrition during reproductive phase. All those factors in turn might be attributed to the better partitioning of photosynthates to reproductive sinks under the control of GA₃ (Morris, 1996) ^[22]. Singh and Kumar (2009) ^[23] recorded maximum flower size and flower stalk length in chrysanthemum using GA₃ at 150 ppm. GA₃ mediated yield increment were also reported by Tyagi and Singh (2006) ^[24], Swaroop *et al.* (2007) ^[25] and Maharnor *et al.*, (2012) ^[26] in

marigold; by Kumar *et al.*, (2003) ^[27] in China aster; by Rakesh and Beniwal (2003) ^[28], Shinde *et al.*, (2010) ^[29] in chrysanthemum.

 Table 2: Influence of different treatments of gibberellic acid on various reproductive parameters of African marigold cv. 'Pusa Narangi Gainda' (data are mean of two season)

Treatments	Days required for flower bud initiation (days)	Flower longevity on plant (days)	Flowering duration (days)	Number of flowers per plant	Flower diameter (cm)	Expected yield (MT/Ha)
T ₀ (control)	60.73	4.67	20.0	34.60	4.01	23.30
T ₁ (GA ₃ @ 50ppm)	59.47	5.67	21.4	40.21	5.25	30.22
T ₂ (GA ₃ @ 100ppm)	58.60	5.77	22.3	42.93	5.77	31.43
T ₃ (GA ₃ @ 150ppm)	58.13	6.13	23.4	43.87	5.94	30.92
T4 (GA3@ 200ppm)	55.47	6.21	23.5	46.57	6.46	35.92
T5 (GA3@ 250ppm)	52.93	6.67	24.4	53.12	6.79	52.28
T ₆ (GA ₃ @ 300ppm)	53.53	6.07	23.6	50.13	6.25	39.02
T7 (GA3@ 350ppm)	57.93	5.67	23.4	46.27	5.81	40.78
T ₈ (GA ₃ @ 400ppm)	58.33	6.00	22.3	43.08	5.63	32.22
T9 (GA3@ 450ppm)	58.67	5.67	22.1	41.23	4.89	29.37
T ₁₀ (GA ₃ @ 500ppm)	59.60	5.67	20.3	37.46	4.75	27.62
S. Em (±)	1.008	0.089	0.378	0.763	0.089	3.69
C. D. at 5%	2.937	0.287	1.101	2.223	0.285	11.86

Biochemical analysis

Treatment with GA3 at 250 ppmresulted in maximum leaf chlorophyll content (54.60 SPAD) and protein content $(1390.5 \ \mu g/g)$ as presented in Table 3. GA₃ had a stimulatory role to decrease the activity of particular enzyme chlorophyllase (Jacob-Wilk et al., 1999) [30], which retards chlorophyll degradation leading to the enhancement of rate of photosynthesis and leaf surface area. Increase in leaf chlorophyll content through application of GA3 was also observed by Mynett et al., (2001) [31] in Freesia; Gad et al., (2016)^[32] in *Ixora coccinea* and Yaghoubi et al., (2013)^[33] in Bellis perennis. Gibberellic acid also checked the degradation of protein and chlorophyll as reported by Faraji et al., (2011) ^[34] during a study in corms of gladiolus cv. White Prosperity. Total carotenoid content in African marigold flowers was found increasing at a steady rate with exogenous application of GA₃ upto 250 ppm but thereafter started to decline sharply upto 500 ppm concentration. The maximum carotenoid

concentration (370.8 mg/1000 g) was observed in GA3 at 250

ppm (T₅) and all the concentrations of GA₃ showed

significantly higher carotenoid concentration over control (148.8 mg/1000 g). Light stimulated chlorophyll biosynthesis increases the carotenoid content (Giuliano *et al.*, 1993) ^[35]. GA₃ mediated enhancement in carotenoid especially lutein biosynthesis in African marigold was also observed by Valadon and Mummery (1967) ^[36].

Exogenous treatment with GA₃ at 250 ppm also resulted in highest concentration of carbohydrate (288.9 µg/g), reducing sugar content (146.1 µg/g) and non-reducing sugar content (142.8 µg/g) in petals of African marigold as depicted in Fig 1. Increased number of leaves per plant resulted in higher photosynthesis than the rest producing greater amount of photosynthates which were channelized to different parts of the plant helping all round development during the vegetative phase reflected by higher content of carbohydrates. The GA₃ mediated enhancement in growth, biomass production and carbohydrate content in crop plants was observed by Demura and Ye (2010) ^[37]. The above results are in close conformity with the findings of Singh and Bijimol (2001) ^[38] in tuberose and Sujatha *et al.*, (2002) ^[39] in gerbera.

 Table 3: Influence of different treatments of Gibberellic acid on chlorophyll content of leaves, protein of petals and total carotenoids of petals of African marigold cv. 'Pusa Narangi Gainda' (data are mean of two season)

Treatments	Chlorophyll (SPAD)	Protein (µg/g)	Total carotenoid (mg/1000 g)
T ₀ (control)	48.8	626.2	148.8
T ₁ (GA ₃ @ 50ppm)	50.8	790.5	197.4
T ₂ (GA ₃ @ 100ppm)	51.3	914.3	204.1
T ₃ (GA ₃ @ 150ppm)	51.9	983.3	219.0
T ₄ (GA ₃ @ 200ppm)	53.3	1159.5	351.5
T5 (GA3@ 250ppm)	54.6	1390.5	370.8
T ₆ (GA ₃ @ 300ppm)	53.9	1342.9	361.1
T7 (GA3@ 350ppm)	52.9	1052.4	241.6
T ₈ (GA ₃ @ 400ppm)	51.5	916.7	213.0
T ₉ (GA ₃ @ 450ppm)	50.8	809.5	198.8
T ₁₀ (GA ₃ @ 500ppm)	50.8	631.0	190.3
S. Em (±)	0.824	16.892	4.007
C. D. at 5%	2.648	49.223	12.501

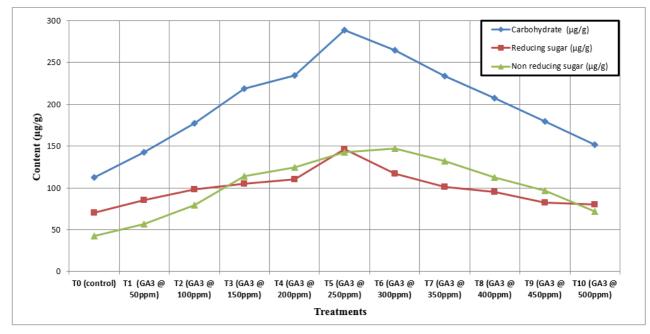


Fig 1: Influence of different treatments of Gibberellic acid on total carbohydrate content ($\mu g/g$), reducing sugar content ($\mu g/g$) and non-reducing sugar content ($\mu g/g$) of petal of African marigold cv. 'Pusa Narangi Gainda' (data are mean of two season)

References

- 1. Sunitha HM, Hunje R, Vyakaranahal BS, Bablad HB. Effect of pinching and growth regulators on plant growth, flowering and seed yield in African marigold (*Tagetes erecta* Linn.). Journal of Ornamental Horticulture. 2007; 10(2):91-95.
- 2. Sharma DP, Patel M, Gupta N. Influence of nitrogen, phosphorus and pinching on vegetative growth and floral attributes in African marigold (*Tagetes erecta* Linn.). Journal of Ornamental Horticulture. 2006; 9(1):25-28.
- 3. Gawle SK, Chaturvedi MK, Yadav KN. Adoption pattern of improved marigold production technologies by the farmers in Bilaspur district of Chhattisgarh. Agriculture Update. 2012; 7(3/4):323-329.
- 4. Kanwar J, Khandelwal SK. Effect of plant growth regulators on growth and yield of African Marigold (*Tagetes erecta* Linn.). Madras Agricultural Journal. 2014; 101(1):45-47.
- Iqbal N, Nazar R, Khan M, Masood A, Khan NA. Role of gibberellins in regulation of source-sink relations under optimal and limiting environmental conditions. Current Science (Bangalore). 2011; 100(7):998-1007.
- Fernández M, Sanz LM, Lewis DR, Muday GK, Lorenzo O. Nitric oxide causes root apical meristem defects and growth inhibition while reducing PIN-FORMED 1 (PIN1)-dependent acropetal auxin transport. Proceedings of the National Academy of Sciences. 2011: 108(45):18506-18511.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Estimation of protein. Journal of Biological Chemistry. 1951; 193:265.
- 8. Ranganna S. Handbook of analysis and quality control for fruit and vegetable products. Tata McGraw Hill Publishing Company Limited, New Delhi, 2002.
- 9. Hodge JE, Hofreiter BT. In: Methods in Carbohydrate Chemistry, (Eds. Whistler RL and Be Miller JN), Academic Press, New York, 1962.
- Somogyi M. Note on sugar estimation. Journal of Biological Chemistry. 1952; 200:245
- 11. Nelissen H, Rymen B, Jikumaru Y, Demuynck K, Van Lijsebettens M, Kamiya Y, Inze D, Beemster GT. A local

maximum in gibberellin levels regulates maize leaf growth by spatial control of cell division. Current Biology. 2012; 22(13):1183-1187.

- Dhekney SA, Ashok AD, Rengasamy P. Effect of some growth regulating chemicals on growth and flowering of rose cv. 'First Red' under greenhouse conditions. Journal of Ornamental Horticulture. 2000; 3(1):51-53.
- Kumar P, Raghava SPS, Misra RL, Singh KP. Effect of GA₃ on growth and yield of China aster. Journal of Ornamental Horticulture. 2003; 6(2):110-112
- Mohariya AD, Patil BN, Wankhede SG, Band PE, Kartikeyan R. Effect of GA₃ and TIBA on growth, flowering and yield of different varieties of Chrysanthemum. Adv. Plant Sci. 2003; 16(1):143-146.
- 15. Ramdevputra MV, Deshmukh HN, Butani AM, Savaliya JJ, Pansuriya AG, Kanzaria DR. Effect of different Gibberellic acid (GA₃) concentrations on growth, flowering and yield of African marigold. The Asian Journal of Horticulture. 2009; 4(1):82-85.
- Jiang X, Li H, Wang T, Peng C, Wang H, Wu H, Wang X. Gibberellin indirectly promotes chloroplast biogenesis as a means to maintain the chloroplast population of expanded cells. The Plant Journal. 2012; 72(5):768-780.
- 17. Dutta JP, Seemanthini R. Growth and flowering response of chrysanthemum (*Dendranthema grandiflora* Tzelev.) to growth regulator treatments. The Orissa Journal of Horticulture. 1998; 26(1):70-75.
- Kumar R, Ram M, Gaur G S. Effect of GA3 and ethrel on growth and flowering of African marigold cv. Pusa Narangi Gainda. Indian Journal of Horticulture. 2010; 67(4):362-366.
- Nair, Sujatha A, Singh, Vijai, Sharma TVRS. Effect of plant growth regulators on yield and quality of gerbera under Bay Island conditions. Indian Journal of Horticulture. 2002; 59(1):100-105
- Porwal R, Nagda CL, Pundir JPS. Influence of plant growth regulators on vegetative growth and flower earliness of damask rose. South Indian Horticulture. 2002; 50(1-3):119-123.
- 21. Kumar J, Singh AK, Pal K. Effect of GA3 and urea on growth and flowering Intuberose (*Polianthes tuberosa* L.)

cv. Pearl Double. Annals of Horticulture. 2009; 2(2):201-203.

- Morris DA. Hormonal regulation of source-sink relationships: an overview of potential control mechanisms. Photoassimilate distribution in plants and crops. Source-sink relationships. Zamski, E. & Schaffer, AA (Ed.). Marcel Dekker, New York, 1996, 441-466.
- 23. Singh JR, Kumar K. Effect of GA3, CCC and inorganic fertilizers on growth and flowering in Chrysanthemum (*Dendranthema grandiflora* Ramet.) cv. Birbal Shahni. Annal Horticulture. 2009; 2 (2):232-233.
- 24. Tyagi AK, Singh. Effect of gibberellic acid and vermicompost on vegetative growth and flowering in African marigold (*Tagetes erecta* Linn.). Journal of Ornamental Horticulture. 2006; 9(2):150-151.
- 25. Swaroop, Kishan, Singh KP, Raju DVS. Vegetative growth, flowering and seed characters of African marigold (*Tagetes erecta* Linn.) as influenced by different growth substances during mild off seasons. Journal of Ornamental Horticulture. 2007; 10:268-70.
- 26. Maharnor SI, Chopde N, Thakre S, Raut PD. Flower yield and quality of African marigold as influenced by nitrogen and pinching. Journal of Soils and Crops. 2012; 22(2):375-378.
- 27. Kumar P, Raghava SPS, Misra RL, Singh KP. Effect of GA₃ on growth and yield of China aster. Journal of Ornamental Horticulture. 2003; 6(2):110-112.
- 28. Rakesh SRS, Beniwal BS. Effect of GA₃ and pinching on growth and yield in chrysanthemum. Haryana Journal of Horticultural Sciences. 2003; 32(1-2):61-63.
- 29. Shinde KH, Parekh NS, Upadhyay NV, Patel HC. Investigation of different levels of Gibberellic acid (GA₃) and pinching treatments on growth, flowering and yield of chrysanthemum (*Chrysanthemum morofolium* Ramat.) cv. 'IIHR-6' under middle Gujarat conditions. The Asian Journal of Horticulture. 2010; 5(2):416-419.
- 30. Jacob-Wilk D, Holland D, Goldschmidt EE, Riov J, Eyal Y. Chlorophyll breakdown by chlorophyllase: isolation and functional expression of the Chlase1 gene from ethylene treated Citrus fruit and its regulation during development. The Plant Journal. 1999; 20(6):653-661
- Mynett K, Startek L, Zurawik P, Ploszaj B. The Effect of Gibrescol and Flordimexon the Emergence and Growth of *Freesia* Rocz.AR w Poznaniu CCCXXXII, Ogrodn 33: 2001, 103-110 (In Polish)
- 32. Gad MM, Abdul-Hafeez EY, Ibrahim OHM. Foliar application of salicylic acid and gibberellic acid enhances growth and flowering of *Ixora coccinea* L. plants. International Journal of Plant Production. 2016; 7(1):85-91.
- 33. Yaghoubi L, Hatamzadeh A, Bakhshi A. Effect of gibberellic acid and mateljasmonat on Growth and Photosynthetic pigments of *Bellis perennis* plants. Proceedings 8 red Congress Sciences Horticulture hemadan Branch, Ab Ali Sina university, Iran, 2013, 3100-3096.
- 34. Faraji S, Naderi R, Ibadli OV, Basaki T, Gasimov SN, Hosseinova S. Effects of post harvesting on biochemical changes in gladiolus cut flowers cultivars. Middle East Journal of Scientific Research. 2011; 9(5):572-577.
- 35. Giuliano G, Bartley GE, Scolnik PA. Regulation of carotenoid biosynthesis during tomato development. The Plant Cell. 1993; 5(4):379-387
- 36. Valadon LT, Mummery RS. Carotenoids of certain compositae flowers. Phytochemistry. 1967; 6(7):983-988.

- Demura T, Ye ZH. Regulation of plant biomass production. Current Opinion in Plant Biology. 2010; 13(3):298-303.
- Singh AK, Bijimol G. Influence of growth-regulating chemicals on growth, flowering and bulb production in tuberose (*Polianthus tuberose* L.). Indian Perfumer. 2001; 45(1):31-34
- Sujatha K, Gowda JVN, Khan MM. Effects of different fertigation levels on gerbera under low cost greenhouse. Journal of Ornamental Horticulture New Series. 2002; 5(1):54-59.