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Standardization of agro-techniques for establishment of cashew (*Anacardium occidentale* L.) plantations in strongly saline sandy loam coastal soils

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Abstract

Good rhizosphere soil environment is fundamental to establishment and growth of tree crops in coastal degraded soils. Experiments were conducted at Northern Coastal Tamil Nadu, India, to determine the influence of agronomic management methods on the survival and growth of cashew (*Anacardium occidentale* L.) in strongly saline sandy loam coastal soils. To identify the suitable and sustainable agro-technology for tree establishment two methods of planting viz., pit method and auger hole method and combination of ameliorative amendments such as FYM, composted coir pith and pressmud along with gypsum, ZnSO₄ and biofertilizers were used. After three years of experimentation plant survival and growth parameters were statistically analyzed for knowing its feasibility in coastal degraded environment. The results showed that a significantly beneficial improvement in planting methods and soils with and without ameliorative amendments. Accordingly the study justified that, auger hole method of planting in combination with composted coir pith @ 25 kg pit⁻¹, gypsum @ 500 g pit⁻¹, ZnSO₄ @ 60 g pit⁻¹, *Azospirillum* @ 50 g pit⁻¹ and Phosphobacteria @ 50 g pit⁻¹ was significantly registered the maximum establishment percentage of 88.25, plant height of 231.72 cm, girth of 14.98 cm, RGR of 2.75 (cm month⁻¹) and taproot length of 118.36 cm.

Keywords: Ameliorative amendments, degraded lands, planting methods, site preparation, waste lands

Introduction

The cashew (*Anacardium occidentale* L.) family Anacardiaceae is a multipurpose tree native of Brazil and has become naturalized in coastal regions of tropical countries of the world including India. Cashew plantations are considerable importance for the social economy of developing nations, since it is very well adapted to wide range of agro-climatic conditions and degraded/waste land soils in tropical and subtropical regions and generate revenue for small and marginal farmers and landless labours hence popularly known as 'gold mine of wasteland'. The cashew kernel is rich in protein (21%), carbohydrate (22%) and fat (47%) and a kilogram of nut yields about 6000 calories of energy (Nambiar *et al.*, 1990) [23]. Cashew kernel lipids are rich in unsaturated fatty acids such as oleic acid (73.7%), linoleic acid (7.67%) and stearic acid (11.2%) (Saroj *et al.*, 2014) [35]. The cashew apple is basically intended for the manufacturing industry consists of alcoholic beverage, synthesise pectin, sweets and protein-enriched animal feed (Apine and Jadhav, 2015; Bhoomika and Sudha Rani, 2018) [2, 4]. The bark is used in tanning, insecticide, adhesive, as substitute for gum Arabic and making ink. The cashew nut shell contains 25-30 per cent dark reddish brown viscous liquid known as cashew nut shell liquid (CNSL). CNSL is a versatile industrial raw material with high technological potential due to its phenolic constitution including anacardic acid, cardanol and cardol. It is used in the cosmetic industry, pharmaceutical industry, textile industry, paper industry, ink making, plastic, varnish, insulating material, paint, lubricant, petro chemical and preservation coatings for wood (Raghavendra Prasada, 2014; Telascrea *et al.*, 2014; Taiwo, 2015; Leite *et al.*, 2016; Shi *et al.*, 2019) [30, 40, 39, 18, 37].

The production of cashew kernels in the world exhibited an increasing trend from past three decades. The total world production of cashew was around 11.75 lakh tonnes during 1994 which rose to 48.98 lakh tonnes by 2016 (FAO, 2017) [9]. India is the largest producer, processor, consumer and exporter of cashew in the world (Elakkiya *et al.*, 2017) [7]. Area under cashew nuts in India has increased from 464 to 1062 thousand ha and production has increased from 185 to 817 thousand tonnes with the productivity of 399 to 769 kg ha⁻¹ during 1980-81 and 2017-18, respectively (Nayak and Paled, 2018) [25]. Similarly the production of cashew nuts in Tamilnadu has also increased considerably during the years and the present production

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is about 67.65 thousand metric tonnes from an area of 141.33 thousand ha (HSD, 2017)^[14]. India's cashew kernels export decreased from 118, 540 metric tonnes in 2006-07 to 82, 302 metric tonnes in 2016-17. In the same time the import of raw kernels increased markedly and more than one third of the overall processing need of the country is met by the imports. Around 18-27 per cent of the shell weight is with CNSL and about 1.25 lakh tones of CNSL are produced annually worldwide. Though India processes more than two million tonnes of raw cashew nut annually, the production of its shell liquid is limited, as all processing units do not produce it. Currently less than 60, 000 tonnes of CNSL is produced in the country as against the potential of 160, 000 tonnes.

In spite of its potentialities, the Indian cashew industry with a processing capacity of two million tonnes annum⁻¹ has been depending on other producing countries for almost half of its requirement for several decades (Nair, 2018)^[22]. Further, the production growth of raw cashew nuts in India (3.1%) is far behind the world (6.3%) growth levels. In this situation, due to the world market demand, if other countries have establishing their own processing units in a big way, the availability of raw cashew for import is steadily declining. Hence, there is a need to expand the production to meet the requirement of the processing industries. As a multipurpose tree having wider agro-climatic adaptability and grows very well on wide range of soils, there is still much scope for area expansion in waste /degraded lands (Rex Immanuel and Ganapathy, 2019a)^[33]. According to people's perception *A. occidentale* is flourishing well under stress situations in coastal habitat's of Tamilnadu (Rex Immanuel *et al.*, 2018a)^[31]. However, its survival and establishment in degraded lands is often affected due to the condition of the soil is far worse now than the native condition in which local vegetation once thrived (Topper *et al.*, 2001; Hammed and Olaniyan, 2012; Nayak and Paled, 2018; Olubode *et al.*, 2018; Rex Immanuel and Ganapathy, 2019b)^[41, 13, 25, 26, 34]. Systematic management of degraded coastal agroecosystem soils with appropriate agro-techniques can tackle these complicated problems faced by the cashew farming communities. With the above background a field experiment was conducted to assess the effectiveness of agronomic management strategies on the performance of *Anacardium occidentale* trees planted in strongly saline sandy loam coastal soils.

Materials and Methods

The geographical situation of study area is 12°12' N Latitude and 79°50' E Longitude with an altitude of +7.2 m mean sea level. The site experienced semi arid to sub-humid climate with the mean annual rainfall of 1350 mm of which 80 per cent is received during North-East monsoon (Oct. – Dec.) and the remaining is received South West monsoon and summer showers. The potential evapotranspiration varied from 1700 to 1900 mm resulting in an annual water deficit of 350 – 550 mm. The length of the crop growing period varied from 80 to 120 days. The mean annual maximum and minimum temperatures are 33.5° C and 23.5° C, respectively. The pH and EC (dsm⁻¹) of the study area were 8.24 and 9.78, respectively. The nutrient status is low in organic carbon (0.27%), N (68.83 kg ha⁻¹), P₂O₅ (7.34 kg ha⁻¹) and K₂O (164.13 kg ha⁻¹).

The treatment consisted of two land management methods (M₁ - Pit method and M₂ - Auger hole method) in main plots and four ameliorative amendments in sub plots (S₁ - FYM @ 30 kg pit⁻¹, S₂ - Pressmud @ 35 kg pit⁻¹, S₃ - Composted coir pith @ 25 kg pit⁻¹ and S₄ - Control (without amendments)).

Along with the organic amendments, gypsum @ 500 g pit⁻¹, ZnSO₄ @ 60 g pit⁻¹, *Azospirillum* @ 50 g pit⁻¹ and Phosphobacteria @ 50 g pit⁻¹ were mixed thoroughly and used. The experiments were laid out in split plot design with three replications. The cashew seedlings were planted under high density planting with the spacing of 5 x 5 m and the plot size of 250 m². The field was fenced, cleaned and ploughed once and leveled. Then the planting pits were prepared according to the treatment schedule. The normal pits were formed with a dimension of 0.60 m³. The auger holes were prepared with a dimension of 0.20 m and at a depth of 0.90 m by using mechanical auger. The pits and holes were filled with mixture of original soil and amendments and the seedling was placed in the center. The observations such as establishment per cent, plant height and girth at 36 months (cm) after planting and relative growth rate (cm month⁻¹) (RGR) were recorded. To study the treatment effects on root development pattern, root systems were exposed in the field after one year following wet excavation methodology (Bohm, 1979)^[5].

Results and Discussion

In the present study, the auger hole method of planting along with the integrated application of composted coir pith @ 25 kg pit⁻¹, gypsum @ 500 g pit⁻¹, ZnSO₄ @ 60 g pit⁻¹, *Azospirillum* @ 50 g pit⁻¹ and Phosphobacteria @ 50 g pit⁻¹ (M₂S₃) significantly recorded the maximum establishment percentage of 88.25, plant height of 231.72 cm (36 months), girth of 14.98 cm (36 months) and RGR of 2.75 (cm month⁻¹). Overall effect of site preparation techniques on survival and growth were in the order of auger hole method followed by pit method of planting. Roots of auger hole planted seedlings grew deeper soil layers (118.36 cm in 12 months after planting) and the majority of roots remained in the amended soil and followed a vertical path instead of growing horizontally. In the pit planting method, the whole root system was confined to the pit soil and formed a shallow root system. It can expose the plant to adverse climatic conditions such as moisture stress prevailed in the region. These observations ensure a way for the planting of *A. occidentale* in saline soils in a vertical direction (auger hole) is more essential than in a horizontal direction (pit method). Auger hole method pierces through hard sub soil layers (kanker pan), improved the porosity by loosening the soil and provides larger volume of reclaimed soil for the development of feeder roots during the initial years of growth. It eases the penetration and proliferation, encourages and trains deeper rooting, does not cause roots to coil or face constriction which adversely affect plant growth, improves the penetration of water through the holes thereby inducing better water storage and its availability and ensures deeper reclamation of soil was reported by Gupta *et al.* (1995)^[11], Dagar *et al.* (2001)^[6] and Pazhanivelan *et al.* (2006)^[27].

The synergistic and cumulative effect of composted coir pith with gypsum, ZnSO₄ and biofertilizers could be the reason for better performance of cashew in terms of establishment per cent, plant height, girth RGR and root length. Composted coir pith with ameliorative amendments sustain growth of cashew through better nutrient recycling and improvement in soil physical, chemical and biological properties. The slow mineralization of nutrients in composted coir pith promotes crop growth over a long period and increases soil water holding capacity making the nutrients more readily available to plant. Coir pith has very high moisture retention capacity of 500-600 per cent and can be as high as 1100 per cent of dry weight (Evans *et al.*, 1996)^[8]. It is also reported that coir pith

retained the maximum moisture after 90 days of composting (Mbah and Pdili, 1998) ^[20]. Geetha *et al.* (2005) ^[10] and Natarajan *et al.* (2005) ^[24] observed that addition of composted coir pith reduced the bulk density of the soil and favorably improved the physical properties *viz.*, hydraulic conductivity and water holding capacity of the soil. High CEC enables it to retain large amounts of nutrients and the adsorption complex has high contents of exchangeable K, Na,

Ca and Mg (Evans *et al.*, 1996; Verhagen and Papadopoulos, 1997) ^[8, 42]. Because of its high moisture retention capacity, coir pith compost encouraged the activity of beneficial microorganisms and involved in solubilisation and mobilization of nutrients. All these characteristics make it ideal for use as a soil amendment, especially for dry and sandy areas with low moisture retention capacity.

Table 1: The effect of agro techniques on the establishment per cent and plant height (36 months after planting)

Treatments	Establishment percentage			Plant height (cm)		
	M ₁	M ₂	Mean	M ₁	M ₂	Mean
S ₁	75.36 (59.85)	82.22 (64.22)	78.79 (62.04)	176.92	192.16	184.54
S ₂	73.21 (58.19)	77.86 (62.52)	75.54 (60.34)	191.20	213.82	202.51
S ₃	80.24 (64.69)	88.25 (70.08)	84.25 (67.39)	205.30	231.72	218.51
S ₄	69.74 (55.20)	74.12 (59.83)	71.93 (57.52)	148.02	165.27	156.65
Mean	74.64 (60.73)	80.21 (64.16)		180.36	200.74	
	S. Ed		CD (P = 0.05)	S. Ed		CD (P = 0.05)
M	1.45		3.10	4.23		08.96
S	1.58		3.25	5.21		10.50
M x S	2.30		4.62	5.78		11.85

(Figures in parenthesis indicate arc-sine transformed values)

The ameliorative amendments such as composted coir pith and gypsum initially reduced the soil pH and EC_e of the soil solution around the root zone and also increased the mineralization of native N due to reduction in pH and slow release of macro and micro nutrients. This resulted in steady availability of nutrients to a longer period which created a favorable environment for plant growth. In addition, the presence of organic materials during decomposition, releases organic acids and presence of acid forming SO₄⁻ ion in gypsum reduced the ill effects of salinity. The superiority of these amendments in reducing soil pH and EC_e have been reported by Murali and Gupta (2001) ^[21] and Natarajan *et al.* (2005) ^[24]. Gypsum application markedly increased the growth of plants planted in the saline environments (Anwar *et al.*, 2003; Hamza and Anderson, 2003; Pazhanivelan *et al.*, 2006) ^[3, 12, 27]. Addition of zinc is necessary for root cell membrane integrity and functions of the bio-membranes of

plants and alleviates possible Na and Cl injury by inhibiting Na and/or Cl uptake and translocation of plants (Khoshgoftar *et al.*, 2004 and Abd-El-Hady, 2007) ^[16, 1].

Table 2. The effect of agro techniques on the girth and relative growth rate (36 months after planting)

Treatments	Girth (cm)			RGR (cm month ⁻¹)		
	M ₁	M ₂	Mean	M ₁	M ₂	Mean
S ₁	11.66	12.88	12.27	2.02	2.52	2.27
S ₂	10.40	11.82	11.11	2.24	2.65	2.45
S ₃	13.26	14.98	14.12	2.37	2.75	2.56
S ₄	09.09	09.51	9.3	1.47	1.74	1.61
Mean	11.10	12.30		2.03	2.42	
	S. Ed		CD (P = 0.05)	S. Ed		CD (P = 0.05)
M	0.21		0.44	0.06		0.12
S	0.42		0.85	0.06		0.14
M x S	0.53		1.09	0.04		0.09

Table 3. The effect of agro techniques on the taproot length (cm) (12 months after planting)

Treatments	Taproot length (cm)		
	M ₁	M ₂	Mean
S ₁	89.15	104.62	96.89
S ₂	74.56	089.11	81.84
S ₃	96.11	118.36	107.24
S ₄	48.23	055.17	51.70
Mean	77.01	91.82	
	S. Ed		CD (P = 0.05)
M	3.68		7.42
S	2.94		6.01
M x S	4.63		9.58

Application of *Azospirillum* and Phosphobacteria improved mineralization of available plant nutrients coupled with fixation of atmospheric nitrogen in to the soil which enhanced the growth of seedlings. Application of *Azospirillum* during planting involved in growth promotion through the production of bioactive molecules *viz.*, Abscisic acid, Gibberellic acid, Indole 3-acetic acid, Ethylene and Cadaverine, which facilitating nutrient uptake, accelerating mineralization, reducing plant stress, stimulating nodulation and providing nitrogen fixation (Somers *et al.*, 2005; Perrig *et al.*, 2007 and Mallik and Williams, 2008) ^[38, 28, 19]. In degraded soils, phosphorus might be present in considerable amounts;

however the preferred form for assimilation is usually much depleted owing to adsorption to soil particles or conversion into organic complexes. Lal (2002) ^[17] and Kaleeswari *et al.* (2007) ^[15] reported that Phosphobacteria produce organic acids such as monocarboxylic acid (acetic, formic), monocarboxylic hydroxy (lactic, glucenic, glycolic), monocarboxylic, ketoglucenic, decarboxylic (oxalic, succinic), dicarboxylic hydroxy (malic, maleic) and tricarboxylic hydroxy (citric) acids, which is involved in solubilizing inorganic phosphate compounds which helps in better height, girth, RGR and root length of trees. There is an increasing evidence that phosphobacteria improved the

growth due to biosynthesis of plant growth substances rather than their action in releasing available phosphorous (Ponmurugan and Gopi, 2006) [29]. Shankarappa *et al.* (2017) [36], Rex Immanuel *et al.* (2018b) [32] and Rex Immanuel and Ganapathy (2019b) [34] reported that the collective use of *Azospirillum* and phosphobacteria have more benefit in unproductive and stressful environments and significantly increased the seedling growth of trees. Hence, *Azospirillum* and phosphobacteria considered to be involved in alleviating the ill effects of stress and enhanced the growth of cashew in combination with organic amendments, gypsum and ZnSO₄.

Conclusion

The observations point out that auger hole method of planting with composted coir pith @ 25 kg pit⁻¹, gypsum @ 500 g pit⁻¹, ZnSO₄ @ 60 g pit⁻¹, *Azospirillum* @ 50 g pit⁻¹ and Phosphobacteria @ 50 g pit⁻¹ is the most appropriate and sustainable agro-technique for planting of *Anacardium occidentale* in degraded coastal soils.

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References

1. Abd-El-Hady BA. Effect of zinc application on growth and nutrient uptake of barley plant irrigated with saline water. *The Journal of Applied Sciences Research*. 2007; 3(6):431-436.
2. Apine OA, Jadhav JP. Fermentation of cashew apple (*Anacardium occidentale*) juice into wine by different *Saccharomyces cerevisiae* strains: a comparative study. *Indian Journal of Research*. 2015; 4(3):6-10.
3. Anwar ZM, Mujeeb FG, Sarwar Hassan NM, Hassan G. Agro amelioration of saline sodic soils. *Journal of Biological Sciences*. 2003; 3(3):329-334.
4. Bhoomika HR, Sudha Rani N. Problems and Prospects of Cashew Cultivation in India - An Overview. *International Journal of Current Microbiology and Applied Sciences*. 2018; 7(10):3687-3694
5. Bohm, W. Methods of studying root systems. *Ecological studies*. Springer, New York. 1979, 186
6. Dagar JC, Singh G, Singh NT. Evaluation of forest and fruit trees used for rehabilitation of semi arid alkali – sodic soils in India. *Arid Land Research and Management*. 2001; 15(2):115-113.
7. Elakkiya E, Sivaraj P, Vijayaprabhakar A. Growth and performance of cashew nut production in India- an analysis. *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(6):1817-1823.
8. Evans MR, Konduru S, Stamps RH. Source variation in physical and chemical properties of coconut coir dust. *Horticultural Science*. 1996; 31:965-967.
9. FAO. Crops, 2017; <http://www.fao.org/faostat/en/#data/QC> (Accessed on 20.04.2019).
10. Geetha D, Suharban M, Vijayan M, Pramod R. Coir pith compost for increasing banana crop production efficiency. *Indian Coconut Journal*. 2005; XXXV(9):7-9.
11. Gupta RK, Tomar OS, Minhas PS. Managing salty soils and waters for afforestation. *CSSRI Bulletin No. 7/95*, Central Soil Salinity Research Institute, Karnal, 1995, 23.
12. Hamza MA, Anderson WK. Response of soil EC and grain yields to deep ripping and gypsum application in a compacted loamy sand soil contrasted with a sandy clay loam soil in Western Australia. *Australian Journal of Agricultural Research*. 2003; 54(3):273-282.
13. Hamed LA, Olaniyan AB. Field establishment of cashew (*Anacardium occidentale* L.) transplants as affected by nursery periods, *Journal of Agricultural Science and Technology*, 2012, 1158-1164.
14. HSD. Horticultural Statistics at a Glance. Horticulture Statistics Division, Department of Agriculture, Cooperation & Farmers Welfare Ministry of Agriculture & Farmers Welfare, Government of India, 2017, 414.
15. Kaleeswari RK, Meena S, Latha MR, Sarvanapandian P, Latha N, Indriani R. Impact of organics and P solubilizing micro-organisms on phosphatic fertilizers under wetland ecosystem. *Research Journal of Agriculture and Biological Sciences*. 2007; 3(6):577-580.
16. Khoshgoftar AH, Shariatmadari H, Krishnan N, Kalhasi M, Vander Zee SEATM *et al.* Salinity and Zinc application effects on phyto availability of Cadmium and Zinc. *Soil Science Society of America Journal*. 2004; 68(6):1885-1889.
17. Lal L. Phosphatic biofertilizers. *Agrotech*, Publ. Academy, Udaipur, India, 2002, 224.
18. Leite AS, Islam T, Júnior ALG, Sousa JMC, Alencar MVOB, Paz MFCJ *et al.* Pharmacological properties of cashew (*Anacardium occidentale*). *African Journal of Biotechnology*. 2016 15(35):1855-1863.
19. Mallik MAB, Williams RD. Plant Growth Promoting Rhizobacteria and Mycorrhizal Fungi in Sustainable Agriculture and Forestry. In: Ren Sen Zeng, Azim U. Mallik and Shi Ming Luo (Eds.) *Allelopathy in Sustainable Agriculture and Forestry*, Springer, New York. 2008, 321-345.
20. Mbah BN, Pdili PN. Changes in moisture retention properties of five waste materials during short-term mesophilic composting. *Compost Science and Utilization*. 1998; 6(4):67-73.
21. Murali G, Gupta A. Coir waste for scientific cause. *Indian Coconut Journal*. 2001; XXXI(12):13-15.
22. Nair GK. Raw cashew nut output to double by 2025-26 on improved acreage. 2018; <https://www.thehindubusinessline.com/economy/agri-business/raw-cashew-nut-output-to-noproduction-no-double-by-2025-26-on-improved-acreage/article23446466.ece>
23. Nambiar MC, Rao B, Thankamma EVV, Pillai PK. Cashew. In: Bose, T.K, Mitra, S.K (Eds) *Fruits: Tropical and Subtropical*, Naya Prakash, Calcutta, 1990, 386-419.
24. Natarajan SK, Thirumurugan V, Mahendran S, Dhanalakshmi R, Balasubramanian R. Influence of coirpith on physio – chemical properties under coastal sandy clay loam soils of Tamil Nadu, In: *Nat. Sem. on New Frontiers of Soil Science Res. towards Sustainable Agriculture*, Madurai, 2005, 13.
25. Nayak M, Paled M. Trends in area, production, yield and export-import of cashew in India an economic analysis. *International Journal of Current Microbiology and Applied Sciences*. 2018; 7(12):1088-1098.
26. Olubode OO, Hamed LA, Odeyemi OM, Adekoya FJ, Meroyi FM, Ogunlade OI. Influence of moisture regimes and organic manure on nutrient dynamics and growth of cashew. *Acta Horticulture*. 2018; 1225:125-132; DOI: 10.17660/ActaHortic.2018.1225.16

27. Pazhanivelan S, Mohamed Amanullah M, Vaiyapuri K, Sathyamoorthi K, Alagesan A, Sharmila Rahale C. Influence of planting techniques and amendments on the performance of Neem (*Azadirachta Indica*) and changes in soil properties in rainfed alkali soil. *Research Journal of Agriculture and Biological Sciences*. 2006; 2(6):443-446.
28. Perrig D, Boiero ML, Masciarelli OA, Penna C, Ruiz OA, Cassan FD *et al.* Plant-growth-promoting compounds produced by two agronomically important strains of *Azospirillum brasilense*, and implications for inoculant formulation. *Applied Microbiology and Biotechnology*. 2007; 75(5):1143-1150.
29. Ponmurugan P, Gopi C. *In vitro* production of growth regulators and phosphatase activity by phosphate solubilizing bacteria. *African Journal of Biotechnology*. 2006; 5(4):348-350.
30. Raghavendra Prasada SA. A review on CNSL biodiesel as an alternative fuel for diesel engine. *International Journal of Science and Research*. 2014; 3(7):2028-2038.
31. Rex Immanuel R, Ganapathy M, Thirupathi M. Perception analysis of coastal agro ecosystem degradation, its effect on agricultural production and the performance of multipurpose tree species. *The Research Journal of Social Sciences*. 2018a; 9(12):122-135
32. Rex Immanuel R, Thirupathi M, Mullaivendhan V. Agronomic management systems for rehabilitation and sustained crop production in coastal agro ecosystem of Tamil Nadu, India. *Innovations in Agriculture*. 2018b; 1(2):28-30; doi: 10.25081/ia.2018. v1.i2.1033.
33. Rex Immanuel R, Ganapathy M. Characterization of degraded lands in coastal agro ecosystem of Northern Tamil Nadu, India. *Journal of Emerging Technologies and Innovative Research*. 2019a; 6(2):200-216
34. Rex Immanuel R, Ganapathy M. Agro-techniques for afforestation of degraded coastal agricultural lands with silk cotton (*Ceiba pentandra* (L.) Gaertn.). *Journal of Pharmacognosy and Phytochemistry*. 2019b; 8(2):1587-1590
35. Saroj PL, Nayak MG, Meena RK. Physiology of Flowering, Fruit and Nut Development in Cashew. In Eds. Ravishankar, H., Singh, V.K., Misra, A.K. and Mishra, M., Souvenir, National Seminar-cum-workshop on physiology of flowering in perennial fruit crops, 2014, 105-114.
36. Shankarappa TH, Mushrif SK, Subramanyam B, Sreenatha A, Maruthi Prasad BN, Aswathanarayana Reddy N. Effect of biofertilizers on growth and establishment of cashew grafts under nursery condition. *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(8): 1959-1965
37. Shi Y, Kamer PCJ, Cole-Hamilton DJ. Synthesis of pharmaceutical drugs from cardanol derived from cashew nut shell liquid. *Green Chemistry*. 2019; 21:1043-1053
38. Somers E, Ptacek D, Gysegom P, Srinivasan M, Vanderleyden J. *Azospirillum brasilense* produces the auxin-like phenylacetic acid by using the key enzyme for indole-3-acetic acid biosynthesis. *Applied and Environmental Microbiology*. 2005; 71(11):1803-1810.
39. Taiwo EA. Cashew Nut Shell Oil - A renewable and reliable petrochemical feedstock. In: *Advances in Petrochemicals*. (Ed) Vivek Patel <https://www.intechopen.com/books/advances-in-petrochemicals/cashew-nut-shell-oil-a-renewable-and-reliable-petrochemical-feedstock>. 2015, DOI: 10.5772/61096
40. Telascrea M, Leao AL, Ferreira MZ, Pupo HFF, Cherian BM, Narine S. Use of a cashew nut shell liquid resin as a potential replacement for phenolic resins in the preparation of panels – a review. *Molecular Crystals and Liquid Crystals*. 2014; 604(1):222-232, DOI: 10.1080/15421406.2014.968509
41. Topper CP, Caligari PDS, Camara M, Diaora S, Dyaha A, Coulibay F *et al.* West African Regional Cashew Survey Report (Guinea, Guinea Bissau; Cote d'Ivoire; Ghana and Nigeria. Sustainable Tree Crops Programme. STCP and Biohybrids Agrisystem Ltd. UK. 2001.
42. Verhagen JBG, Papadopoulos AP. CEC and the saturation of the adsorption complex of coir dust. *Acta Horticulture*. 1997; 481:151-155.