



E-ISSN: 2278-4136
P-ISSN: 2349-8234
JPP 2019; 8(3): 596-599
Received: 07-03-2019
Accepted: 09-04-2019

Satish Sharma
Department of Plant Pathology,
College of Agriculture, R.V.S.K
V.V. Gwalior, Madhya Pradesh,
India

Reeti Singh
Department of Plant Pathology,
College of Agriculture, R.V.S.K
V.V. Gwalior, Madhya Pradesh,
India

Ajay Kumar
Department of Plant Pathology,
College of Agriculture, R.V.S.K
V.V. Gwalior, Madhya Pradesh,
India

Screening of cluster bean genotypes against *Alternaria* blight (*Alternaria Cucumerina* var. *cyamopsidis*) in field condition

Satish Sharma, Reeti Singh and Ajay Kumar

Abstract

Cluster bean has become popular not only for consumption as vegetable but also as good source of useful industrial "Guar gum". Cluster bean commonly recognized as a summer, annual, drought-tolerant, well adapted to arid and semi-arid climates with hot temperatures legume crop. It can grow in sub humid conditions, Total twenty four genotypes were screened. Among different genotypes, 18 were found moderately resistant whereas 6 were moderately susceptible towards *A. blight* of Cluster bean. No genotype was found immune, HR, S and HS against disease. The maximum yield (14.58 q/ha) of guar crop was obtained with the genotype HG 563. The resistant varieties are cheapest source to manage this disease so that more entries should be screened out to find out resistant or immune lines of Cluster bean.

Keywords: Cluster bean, *Alternaria* blight *Alternaria Cucumerina* var. *cyamopsidis*

Introduction

Cluster bean [*Cyamopsis tetragonoloba* (L.) Taub.] is being grown in India since ancient time. Although believed to be of African origin (Vavilov, 1951) [25], it was domesticated centuries ago in the north-western region of the Indo Pakistan sub-continent (Hymowitz, 1972) [4]. Tender green guar pods are important source of nutrition to human being and animals. Primary methods of controlling *Alternaria* leaf blight include preventing long periods of wetness on the leaf surface, cultural scouting, sanitation, and development of the host plant resistant with the application of fungicides (Kirk *et al.*, 2001 and Namanda *et al.*, 2004) [8, 15]. In recent years, an increasing consciousness about environmental pollution due to pesticides and development of fungicide-resistant strain in plant pathogens has challenged the plant pathologists to search for non-toxic fungicides for substituting the recommended chemicals. This is the most cost effective and eco-friendly management strategy selecting the genotypes possessing the resistant/tolerant reaction against the disease. Studies conducted on Blight of guar and its various aspects indicated that there are lot of gaps in the understanding of the disease, the pathogen and the control aspects and very meagre information is available on these aspects. Hence, a thorough investigation is required. Therefore,

Materials and Methods

The experiment was conducted in *Kharif* 2016 and 2017 in the experimental field of Department of Plant Pathology. Five plants from each genotype were tagged for taking the observations. The intensity of the disease was recorded 45 days after sowing using 0-5 scale (Table 1, Plate 1).

Cluster bean materials consisting of 24 lines were evaluated against *Alternaria* blight under artificial inoculated conditions by adopting a field screening technique (Mc. Kinney 1923) [10]. The experiment was laid out in RBD with two replications. In the experiment the spacing between row to row was 45 cm and plant to plant 10 cm. The nitrogen and phosphorous was applied @ 25:50 kg/ha respectively.

After germination, the crop was regularly watched for first appearance of disease. The observation on disease intensity was recorded using a progressive 0-5 scale, (McKinney, 1923) [10] in the Table 1. Numerical rating grade was given on the basis of percentage of area covered by pathogen on the leaves.

$$PDI = \frac{\text{Sum of all individual rating}}{\text{Total no. of leaf assessed} \times \text{Maximum rating}} \times 100$$

Correspondence

Satish Sharma
Department of Plant Pathology,
College of Agriculture, R.V.S.K
V.V. Gwalior, Madhya Pradesh,
India

Table 1: Disease rating scale for Alternaria blight

Rating Scale	Disease Reaction	Per cent Disease Intensity	Description
0	Near immune/ Resistant reaction (I)	0	No symptoms
1	Resistant (R)	1-10	1 - 10 % leaf area infected
2	Moderately Resistant (MR)	11-25	11 – 25 % leaf area infected
3	Moderately Susceptible (MS)	26-50	26 - 50 % leaf area infected
4	Susceptible (S)	51-75	51 – 75 % leaf area infected
5	Highly Susceptible (HS)	76-100	76 % above leaf area Infected

Symptomology; Disease rating for alternaria blight

(No symptoms)

(1 – 10 % leaf area infected)



(11 – 25 % leaf area infected)

(26 – 50 % leaf area infected)



(51 – 75 % leaf area infected)

(75 % above leaf area infected)

Plate 1: Alternaria blight disease rating scale on Cluster bean**Plate 2:** Screening of the cluster bean genotypes against Alternaria blight under artificial inoculation conditions.**Results and Discussion**

Twenty four genotypes of Cluster bean were screened under artificial inoculation field conditions against Alternaria blight disease. The observations on disease intensity on various genotypes were recorded and were categorized as per their reaction. The rating scale 0-5 was used for recording the observation on leaf. Table 16 and Figure 7 revealed that none of Cluster bean entries were found completely free from the disease. Among different genotypes, 18 were found moderately resistant RGC1033, HG2-20, RGr16-2, CAZG-15-7, GAUG1304, RGr16 -7, RGr-17-2, RGR16-10, x-10, HG2-20, GC1033, RGR16-4, CAZG15-3, GC1066, RGr16-3, AUG1305, HG563, GAUG1304 whereas 6 were moderately susceptible GAUG-1501, CAZG-15-5, RGr-17-4, GAUG-1502, CAZG15-4, M-83 (control). None of the genotypes were found immune. The moderately resistant variety HG563 showed the highest yield 14.58 q/ha with 21.48 per cent disease intensity of 21.48 whereas the variety GAUG-1502 showed the minimum yield 6.18 q/ha with 28.88 per cent disease intensity.

Table 2: Field evaluation of Cluster bean genotypes against Alternaria blight on leaf during 2016 and 2017.

S.N.	Genotypes	Per cent Disease Intensity		Mean	Disease Reaction	Yield (q/ha)
		2016	2017			
1.	RGC 1066	16.3 (23.8)	21.10 (27.4)	18.75	MR	11.2
2.	X-10	15.55 (23.2)	20.20 (26.7)	17.93	MR	11.55
3.	RGC 1033	20.2 (26.6)	23.60 (28.9)	21.85	MR	10.45
4.	HG2-20	18.65 (25.5)	24.20 (29.5)	21.50	MR	9.76
5.	RGr16-2	21.15 (27.3)	25.10 (30.2)	23.25	MR	10.25
6.	CAZG 15-7	20.6 (26.9)	24.30 (29.4)	22.40	MR	11.22
7.	GAUG1304	23.4 (28.9)	26.50 (30.9)	24.90	MR	8.80
8.	GAUG-1501	24.65 (29.7)	26.10 (30.8)	25.45	MS	7.95
9.	RGr16 -7	21.7 (27.7)	26.50 (30.9)	24.05	MR	8.25
10.	RGr-17-2	24.25 (29.4)	25.40 (30.2)	24.80	MR	8.54
11.	CAZG-15-5	25.55 (30.3)	27.10 (31.4)	26.43	MS	7.86
12.	RGr-17-4	25.45 (30.2)	26.30 (30.7)	25.83	MS	8.2
13.	GAUG- 1502	28.45 (32.2)	29.40 (32.7)	28.88	MS	6.18

14.	RGr16-10	20.2 (26.6)	24.30 (29.5)	22.30	MR	10.96
15.	HG2-20	22.3 (28.1)	25.30 (30.2)	23.83	MR	11.56
16.	CAZG15-4	24.3 (29.5)	26.50 (30.9)	25.38	MS	9.12
17.	RGC1033	17.6 (24.7)	26.80 (31.0)	22.13	MR	10.32
18.	RGr16-4	20.5 (26.9)	29.30 (32.7)	24.93	MR	11.34
19.	CAZG15-3	21.15 (27.3)	26.40 (30.8)	23.73	MR	12.56
20.	GC1066	20.55 (26.9)	25.1 (30.1)	22.88	MR	9.33
21.	RGr16-3	18.5 (25.4)	24.5 (29.5)	21.45	MR	9.88
22.	GAUG1305	19.45 (26.1)	22.4 (28.3)	20.98	MR	8.56
23.	HG563	18.3 (25.3)	24.7 (29.7)	21.48	MR	14.58
24.	M-83 (Control)	34.35 (35.8)	44.1 (41.6)	39.28	MS	7.96
SE (m) ±		0.12	0.32			
CD at 5%		0.35	0.94			

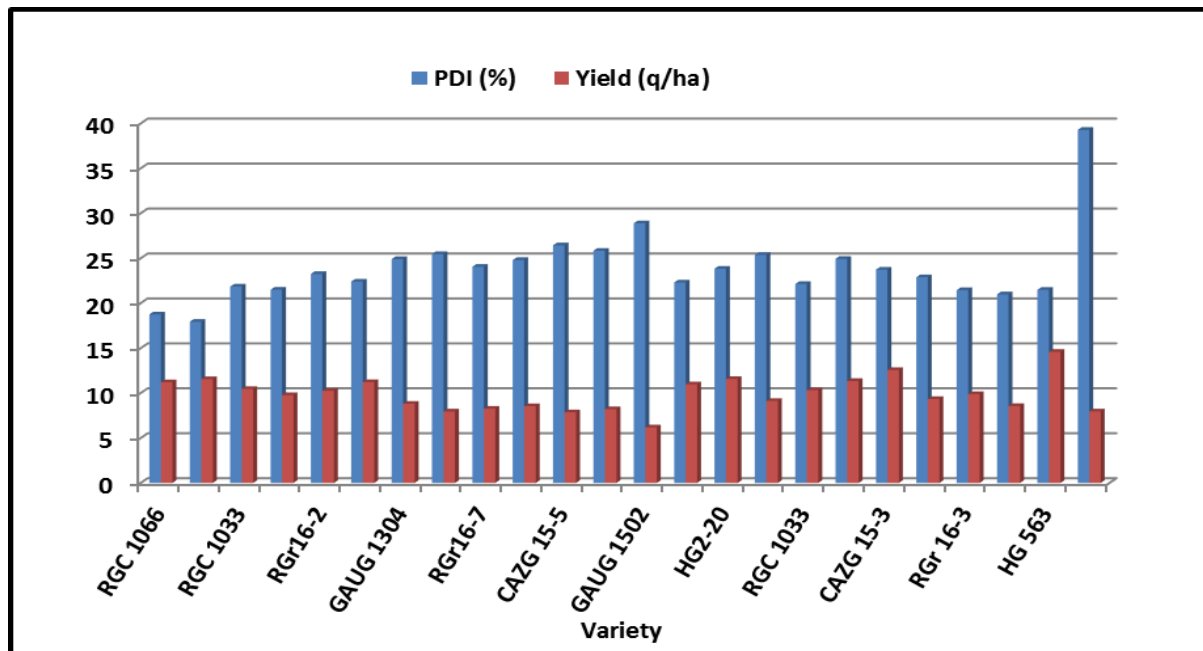


Fig 1: Field evaluation of Cluster bean genotypes against Alternaria blight on leaf.

The use of resistant cultivars is the cheapest, easiest, safest and most effective means of controlling plant diseases in crops for which such cultivars are available. Cultivation of resistant cultivars not only eliminates losses from diseases but also eliminates expenses for sprays and for other ways of disease control thus avoid the contamination of environment with toxic chemicals.

Total 24 genotypes of Cluster bean were evaluated against Alternaria blight (*A. cucumerina* var. *cymopsidis*) under artificial inoculation conditions. The rating scale 0-5 was used for recording observation on leaf. Among different genotypes, 18 were found moderately resistant whereas 6 were moderately susceptible. None of the genotypes of Alternaria blight were found immune, HR, S and HS against disease.

The limited information is available on the reaction of cluster bean varieties to infection by *Alternaria cyamopsidis* (Saini and Paroda, 1983) [17]. RGC-619, RGC-677 and RGC-679 were reported to be least susceptible by Guar and Ahmed (1983) [3]. Shivanna and Shetty (1991) [18] reported HG-812 to be least susceptible to blight in Mysore. Few genotypes of guar viz., HFG-14, HFG-516, HFG-516, HFG-522, HFG-530, HFG-554 (Gandhi *et al.*, 1978) [2] and Guar-279-6, Guar-47 and Gawarani short (Jagtap, 1975) [5] have shown resistance against this disease. Guar-279-6, Guar-47 and Gawarani short (Jagtap, 1975) [5] have shown resistance against this disease. Yejenrappa and Padaganur (1993) [26] showed that Guar plants of all ages were susceptible to infection by leaf blight pathogen.

The possibilities of exploiting cultivars resistant in controlling guar disease are limited. The commercial cultivars either do not exhibit sufficient degree of resistant or the resistant is turned down by new physiological races.

Acknowledgement

The authors are grateful to Dean, college of Agriculture, Gwalior, for providing the field and lab facility for the research work.

References

1. Bharodia PS, Zsaveri PP, Kher HR, Patel MP, Chaudhari DN. GAUG-34 A high gum yielding variety of cluster bean. Indian Farm. 1993; 43(9):31-33.
2. Gandhi SK, Saini ML, Jhorar BS. Screening of clusterbean genotypes for resistance to leaf spot caused by *Alternaria cyamopsidis*. Forage Res. 1978; 4:169-171.
3. Gaur RB, Ahmed SA. Studies on chemical control, source of resistance and survey of Alternaria leaf spot of clusterbean. Forage Res. 1983; 9:179-180.
4. Hymowitz T. The trans-domestication concept as applied to guar. Econ. Bot. 1972; 26:49-60.
5. Jagtap RP. Varietal resistance in clusterbean to *Alternaria cyamopsidis* in Maharashtra state. Res. J Mahatma Phule Agric. Univ. 1975; 6:164.
6. Kamel M, Abraham AN, Kamel SA, El-Fahi AM. Spore germination of *Alternaria tenuis* in juice of susceptible and resistant cotton cultivars to Alternaria leaf spot

- disease. United Arab Republic J Bot. 1971; 14(2):245-254.
7. Kandolo SD, Thompson AH, Calitz FJ, Laurie SM, Truter M, Van Der JE *et al.* Field tolerance of selected varieties and fungicide efficacy against *Alternaria* blight of sweet potato. African Crop Sci. J 2016; 24(3):331-339.
 8. Kirk WW, Flecher DS, Douchs JM, Coombs KM, Hammerschmidt R. Effect of host plant resistance and reduced rates and frequency of fungicides application to control potato late blight. Plant Dis. 2001; 85:1113-1118.
 9. Manjunatha H, Saifulla M. Screening of desi and kabuli chickpea (*Cicer arietinum.*) entries against *Alternaria* blight. Indian J Sci. Res. Tech. 2013; 1(3):66-68.
 10. Mckinney HN. Influence of soil temperature and moisture on infection of wheat seedling by *Helminthosporium sativum.* J Agric, Res. 1923; 26:195-197.
 11. Misra V. Donor for resistance in Chickpea against *Alternaria alternata* (Fr) Keisslar. J Phytol. Res. 2005; 18(2):265-266.
 12. Mogle UP. Efficacy of leaf extracts against the post-harvest fungal pathogens of cowpea. Biosci. Discov. 2013; 4(1):39-42.
 13. Mohapatra A, Mohanty AK, Mohanty NN. Studies on physiology of the sesame leaf blight pathogen, *Alternaria sesami.* Indian Phytotopath. 1977; 30:332-334.
 14. Narula PN, Farooqui O, Singh SP. Screening pigeonpea varieties against leaf blight disease caused by *Alternaria tenuissima.* Indian J Genet. Pl. Breed. 1990; 50(2):171-174.
 15. Namanda S, Olanya OM, Adipala E, Hakiza JJ, Bedewy RE. Fungicide application and host resistance for potato late blight management: benefits assessment from on-farm studies in S.W. Uganda. Crop Protect. 2004; 23:1075-1083.
 16. Pathak R. Clusterbean: physiology, genetics and cultivation, 2015; pp. 87-106.
 17. Saini ML, Paroda RS. Guar cultivation in Haryana. In: Bulletin, Dept. of Pl. Breeding, Haryana, Agric. Univ., Hisar. 1983; pp. 25.
 18. Shivanna MB, Shetty HS. Occurrence of fungal diseases and its relationship with growth stages in clusterbean during different season. Inter. J Pl. Dis. 1991; 9:10-12.
 19. Saxena D, Prasad R. Biochemical changes in resistant and susceptible cultivars of mustard for disease resistance against *Alternaria* blight. Indian Phytopath. 2003; 56:313-314.
 20. Singh HK, Kavita Singh RB, Maurya KN. Management of *Alternaria* blight of Indian mustard through resistance inducing chemicals. J Agric Res. 2014; 1(2):108-111.
 21. Singh HV. Biochemical basis of resistance in *Brassica* species against downy mildew and white rust of mustard. Pl. Dis. Res. 2000; 15(1):75-77.
 22. Singh V, Lal M, Kumar S, Mohd A, Singh J Management of *Alternaria* blight of linseed with sowing dates and host resistance. Universe Emerg. Technol. Sci. 2015; 2(4):1-4.
 23. Tomar DS. Studies on *Alternaria* blight of Guar caused by *Alternaria cyamopsidis.* M.Sc. Thesis, JNKVV, College of Agriculture, Jabalpur, 2000.
 24. Tomar DS, Singh R. Screening of clusterbean germplasm against foliar diseases in Grid region of Madhya Pradesh. JNKVV Res. J. 2004; 38(2):97-98.
 25. Vavilov NI. The origin, variation, immunity and breeding of cultivated plants. Chronica Bot. 1951; 13:2-6.
 26. Yenjerappa ST, Padaganur GM. Relation of clusterbean varieties and plant age to infection by *Alternaria cyamopsidis* Karnataka J Agri. Sci. 1993; 6:305-307.