



E-ISSN: 2278-4136  
P-ISSN: 2349-8234  
JPP 2019; 8(3): 3193-3198  
Received: 07-03-2019  
Accepted: 09-04-2019

**Ipsita De Pradhan**  
Phytochemistry and  
Pharmacognosy Research  
Laboratory, Centre of Advanced  
Study, Department of Botany,  
University of Calcutta, 35  
Ballygunge Circular Road,  
Kolkata, West Bengal, India

**Bratati De**  
Phytochemistry and  
Pharmacognosy Research  
Laboratory, Centre of Advanced  
Study, Department of Botany,  
University of Calcutta, 35  
Ballygunge Circular Road,  
Kolkata, West Bengal, India

## Varietal variation of hexane soluble chemical components of Indian mango (*Mangifera indica* L.) fruit pulp

Ipsita De Pradhan and Bratati De

### Abstract

Hexane soluble non-polar chemical components of fruit pulp of seven Indian *Mangifera indica* L. varieties were investigated and analyzed by GC-MS. The varieties were 'Amrapali', 'Himsagar', 'Fazli', 'Langra', 'Golapkhaskhas', 'Gopalbhog', 'Mohanbhog'. Twenty one aromatic components were identified and some were unidentified components. Multivariate analysis of the identified components indicated that the mango varieties were different on the basis of these non-polar metabolites. The ten most important contributory metabolites for varietal variation were delta-3-carene, beta-myrcene, 1R-alpha-pinene, beta-pinene, humulene, isocaryophyllene, longifolene, undecane, tetradecane, and alpha pinene.

**Keywords:** Mango, aromatic components, GC-MS

### Introduction

*Mangifera indica* L. is one of the most popular tropical fruits cultivated and consumed worldwide. The fruits are very well known for its taste and distinctive flavors [1]. The fruit has nutritional benefits due to its high fibre, vitamin C and  $\beta$ -carotene content [2]. Several hundreds of cultivars are grown in various parts of the world and are known to vary markedly in their flavour characteristics [1]. This fruit is with green skin which turns yellow or lightly reddish yellow when ripe. The skin is not consumed and underneath is a luscious, edible (flesh) mesocarp which surrounds a central seed. The fruits, which are fairly sweet, are mainly eaten fresh but to extend the shelf life and meet consumer demand, mangoes are processed into juice, puree, jam and dried fruits [3, 2]. Although the aromatic compounds of exotic mangoes have been studied extensively [4] but there is very little data on the most popular Indigenous varieties of India. The aroma of each individual variety are extremely complex due to presence of many components, each may have different polarities and volatilities. Most of the aromatic constituents consists of oxygenated compounds which include terpene hydrocarbons, esters, furanones, lactones, ketones, alcohols aldehydes, acid and other groups [5]. The type of aromatic components of mango depends on cultivars [6], maturity level of the fruit [7], the part of the fruit [8], area of production [9], processing method and solvent used [10]. Most fruits produce significant number of aromatic compounds as indicators of fruit ripening. Many of these compounds are produced in trace amounts, which are below the detection level of the analytical instruments [11]. The aim of the current work was to analyze and identify aroma causing non-polar components of seven indigenous varieties, e.g. 'Amrapali', 'Himsagar', 'Fazli', 'Langra', 'Golapkhaskhas', 'Gopalbhog', 'Mohanbhog' grown in Kolkata, West Bengal using GC-MS, and to find out varietal variations through chemometric analysis.

### Materials and methods

#### Plant materials

Seven indigenous varieties of mango were 'Amrapali', 'Himsagar', 'Fazli', 'Langra', 'Golapkhaskhas', 'Gopalbhog', 'Mohanbhog', they were collected in ripe, ready to consume stage, from University of Calcutta Agricultural field, Baruipur, Kolkata, West Bengal, in the month of April- May 2013.

#### Chemical reagents

HPLC grade hexane and a mixture of n-alkanes, [nonene (C<sub>9</sub>), decane (C<sub>10</sub>), undecane (C<sub>11</sub>), dodecane (C<sub>12</sub>), tridecane (C<sub>13</sub>), tetradecane (C<sub>14</sub>), pentadecane (C<sub>15</sub>), hexadecane (C<sub>16</sub>), heptadecane (C<sub>17</sub>), octadecane (C<sub>18</sub>), nonadecane (C<sub>19</sub>), eicosane (C<sub>20</sub>)] were purchased from Sigma-Aldrich.

#### Correspondence

**Ipsita De Pradhan**  
Phytochemistry and  
Pharmacognosy Research  
Laboratory, Centre of Advanced  
Study, Department of Botany,  
University of Calcutta, 35  
Ballygunge Circular Road,  
Kolkata, West Bengal, India

### Extraction of nonpolar aromatic components

The ripened mango of every variety was surface cleaned with distilled water, and then they were peeled properly. The extraction of the mesocarp was done modifying the previously published methods [12, 13]. The yellowish orange pulp portion of the fruits was separated, crushed to powder with liquid nitrogen. The pulp powder ( $150 \pm 10$  mg each) was then transferred in to micro centrifuge tubes, as soon as possible and n-hexane (500  $\mu$ l) was added. The tubes were capped airtight. They were kept at 25 °C for 30 minutes with frequent shaking. The tubes were centrifuged for 10 minutes at 10000 rpm and the supernatants were collected in separate tubes. Five biological replicas of each variety were prepared in the same way. Each extract was treated with Na<sub>2</sub>SO<sub>4</sub> to remove moisture. The whole process was done minimizing direct exposure to air as much as possible to avoid loss of volatile components. The supernatants were then analyzed by GC-MS directly.

### GC-MS analysis

GC-MS analysis was carried out using (Agilent gas chromatography system) 7890 A GC and 5975C MSD with triple axis detector. The GC-MS was operated on EI mode (70V). HP-5MS capillary column (Agilent J&W GC columns, USA) (length 30 m, diameter 0.25 mm narrow bore, 0.25  $\mu$ m) was used. The analysis was performed as detailed earlier [14] under the oven temperature program: injection at 60 °C (5 min), temperature increasing at the rate 4 °C/min to 220 °C, 10 minutes hold time before cooling. The injection temperature was set at 230 °C. The MSD transfer line was set at 280 °C and ion source at 250 °C. Helium was used as carrier gas at a constant flow rate of 1ml/minute, carrier liner velocity being 36.623 cm/sec. Sample (1  $\mu$ l) was injected via the split mode onto the GC column. Mass spectra ranging from 30 to 500m/z were recorded. Metabolites were identified by comparing the fragmentation patterns of the mass spectra with entries of mass spectra library G1033A NIST. The Arithmetic Indices (AI) were calculated for each of the component identified and matched with those reported by Adams, (2009) [15]. Arithmetic index was calculated from the equation:  $AI(x) = 100 Pz + 100 [(RT(x) - RT(Pz)) / RT(Pz + 1) - RT(Pz)]$  where x: compound; RT: retention time; Pz: alkane before x; Pz+1: alkane after x. n-alkane mixture C<sub>8</sub>-C<sub>20</sub> was used. Other scientific journals were also studied for the identification of components [16, 17, 18].

### Statistical analysis

The data were analyzed by different statistical and multivariate analysis such as Principal Component Analysis (PCA), Partial Least Squares - Discriminant Analysis (PLS-DA) obtained using METABOANALYST 4 software.

### Result and discussion

In this analysis 27 aromatic components were identified from the hexane extract of the seven varieties of mango pulp of by comparing mass spectral fragmentation pattern and AI, RI (Retention Index) [15] and by mass spectral fragmentation pattern and R match value from NIST library (Table 1). The statistical analysis was done with the most confirmed 27 aromatic metabolites. Heat map of the identified metabolites indicated that these seven varieties were different from each other on the basis of non-polar metabolite profile (Fig 1). The data were also analyzed by PCA (Fig. 2), PLS-DA (Fig.3). All the models segregated the varieties distinctly from each other. Also in Ten most influential aromatic compounds responsible for the differences were selected (Fig. 4) from the VIP score. Here delta-3-carene, beta-myrcene, 1R-alpha-pinene, (-)-beta-pinene, are the first four most important non polar aromatic compounds which are responsible for the varietal separation on the basis of PLS-DA. The result clearly shows that the different varieties of mango fruit pulps were aromatically different. Delta-3-Carene, the most important contributory metabolite for distinction of the varieties could be detected in 'Langra', 'Himsagar' and 'Mohagbog'. The next important compound beta-myrcene could be detected in 'Gopalbhog', 'Amrapali' and 'Fazli'. 1R-alpha-pinene was detected in 'Amrapali' in noticeable amount; beta-pinene in 'Amrapali'. Attempts are being made to characterize different polar and non-polar metabolites of mango fruits. Different ripening stages of mango fruits were distinguished on the basis of volatile organic compounds in variety "Tommy Atkins" [19] and different organic acids, sugars, polyphenolic compounds [20]. Varietal variation on the basis of different polar metabolites e.g. sugars, organic acids, amino acids, phenols including mangiferin has been reported [21]. In this work attempt was made to distinguish some Indian varieties of mangoes based on their hexane soluble non-polar metabolites which were identified to be mainly aromatic compounds. Two most important metabolites for varietal variation were delta-3-carene and beta-myrcene. The study may be important for rapid identification of mango varieties.

**Table 1:** Identified non-polar components from the pulp of the seven varieties of *M. indica* fruit pulp

| Metabolites  | Formula                         | MW  | R match | AI   | AI   | Reference | MS Fragmentation  |
|--|---------------------------------|-----|---------|------|------|-----------|---|
| $\alpha$ -Pinene   | C <sub>10</sub> H <sub>16</sub> | 136 | 984     | 933  | 932  | *(15)     | 136, 121, 105, 94, 93 (100), 92, 91, 81, 79, 77, 69, 67, 65,                        |
| Beta-Phellandrene  | C <sub>10</sub> H <sub>16</sub> | 136 | 867     | 1013 | 1025 | *(15)     | 136, 121, 93(100), 77, 65, 53, 41   |
| Bicyclo[3.1.1]heptane, 6,6-dimethyl-2-methylene-, (1S)-/(-)- beta pinene | C <sub>10</sub> H <sub>16</sub> | 136 | 962     | 989  | 986  | *(15)     | 136, 121, 107, 93, 79, 69, 53   |
| alpha Phellandrene   | C <sub>10</sub> H <sub>16</sub> | 136 | 892     | 1004 | 1002 | *(15)     | 136, 93(100), 77(40), 65, 51, 41  |
| 1R- $\alpha$ -Pinene   | C <sub>10</sub> H <sub>16</sub> | 136 | 925     | 932  | 936  | *(17)     | 136, 121, 115, 105, 94, 93(100), 91, 89, 81, 79, 77, 69, 67, 65, 63, 57, 55, 53, 51 |
| $\beta$ -Myrcene   | C <sub>10</sub> H <sub>17</sub> | 136 | 932     | 991  | 986  | *(15)     | 136, 121, 107, 93(100), 79, 69, 53  |
| delta 3-Carene   | C <sub>10</sub> H <sub>16</sub> | 136 | 952     | 1009 | 1008 | *(15)     | 136, 121, 115, 105, 94, 93, 91, 81, 79, 77, 69, 67, 65                              |
| $\gamma$ -Terpinene  | C <sub>10</sub> H <sub>16</sub> | 136 | 892     | 1040 | 1055 | *(15)     | 136, 121, 115, 105, 94, 93, 92, 91, 89, 81, 79, 77, 67                              |
| Limonene   | C <sub>10</sub> H <sub>16</sub> | 136 | 922     | 1029 | 1024 | *(15)     | 136, 121, 107, 93, 91, 86, 79, 68, 65, 58, 53                                       |
| Terpinolene  | C <sub>10</sub> H <sub>16</sub> | 136 | 880     | 1088 | 1089 | *(15)     | 136, 121, 105, 93, 91, 80, 79, 77, 67, 65, 63, 55, 53, 51                           |

|                                       |            |     |     |      |      |       |   |
|---------------------------------------|------------|-----|-----|------|------|-------|---|
| $\beta$ -Caryophyllene                | C15H24     | 204 | 808 | 1420 | 1417 | *(15) | 204, 189, 175, 161, 147, 133, 120, 105, 93, 91, 81, 79, 69, 67, 55              |
| Isocaryophyllene                      | C15H24     | 204 | 828 | 1406 | 1407 | *(16) | 204, 189, 175, 161, 147, 133, 115, 119, 105, 98, 93, 91, 85, 79, 77, 69, 67, 55 |
| gamma Patchoulene                     | C15H24     | 204 | 840 | 1495 | 1502 | *(15) | 204, 189, 186, 175, 161, 147, 133, 121, 105, 91, 81, 67, 55                     |
| Germacrene D                          | C15H24     | 204 | 925 | 1484 | 1482 | *(15) | 204, 161, 147, 133, 119, 107, 105, 95, 91, 81, 79, 77, 67, 55                   |
| Humulene                              | C15H24     | 204 | 992 | 1456 | 1455 | *(15) | 204, 189, 161, 147, 121, 105,   |
| $\gamma$ -Elemene                     | C15H24     | 204 | 965 | 1441 | 1434 | *(15) | 204, 189, 175, 161, 147, 133, 121, 107, 93, 79, 67, 53                          |
| $\beta$ -Copaene                      | C15H25     | 204 | 982 | 1435 | 1430 | *(15) | 204, 161, 147, 139, 133, 119, 105, 95, 91, 81, 79, 71, 67, 59, 55               |
| Longifolene                           | C15H24     | 204 | 935 | 1406 | 1407 | *(15) | 204, 189, 175, 161, 147, 133, 119, 107, 105, 91, 79, 67, 57, 55                 |
| Longicyclene (L)                      | C15H24     | 204 | 956 | 1372 | 1373 | *(15) | 204, 189, 161, 147, 133, 119, 105, 94, 91, 79, 69, 65, 59, 55                   |
| Undecane                              | C11H24     | 156 | 865 | 1102 | 1100 | *•    | 156, 127, 113, 85, 71, 57(100), 43  |
| Tetradecane                           | C13H28     | 184 | 912 | 1401 | 1400 | *•    | 141,127,85,71,57(100),43  |
| Dodecane, 2,6,11-trimethyl-           | C15H32     | 212 | 836 | 1280 | 1275 | *     | 212, 196, 182, 169, 154, 140, 126, 112, 99, 85, 71, 69, 57                      |
| Tridecane, 3-methyl-                  | C14H30     | 198 | 843 | 1371 | 1371 | (18)  | 198,169,113,99,85,71,57(100),56(50),55(50)                                      |
| 2,6-Dimethyl-6-trifluoroacetoxyoctane | C12H21F3O2 | 254 | 765 | 1079 | 1067 | *     | 140, 125, 111, 97, 85, 83, 71, 70, 69, 67, 58, 57(100), 55                      |
| 2-Butyloctanol                        | C12H26O    | 186 | 807 | 1274 | 1277 | *     | 186, 154, 140, 125, 111, 97, 85, 72, 69, 67, 58, 57                             |
| 1-Chlorotetradecane                   |            | 232 | 717 | 1646 | 1659 | *     | 232, 204, 189, 119, 105, 91, 71(50), 57(100), 55                                |
| 1-Iodo-2-methylundecane               | C12H25I    | 296 | 841 | 1555 | 1564 | *     | 296, 113, 99, 97, 85, 83, 71, 69, 57, 55, 53                                    |

• Authentic; \* NIST Library; MW: Molecular weight

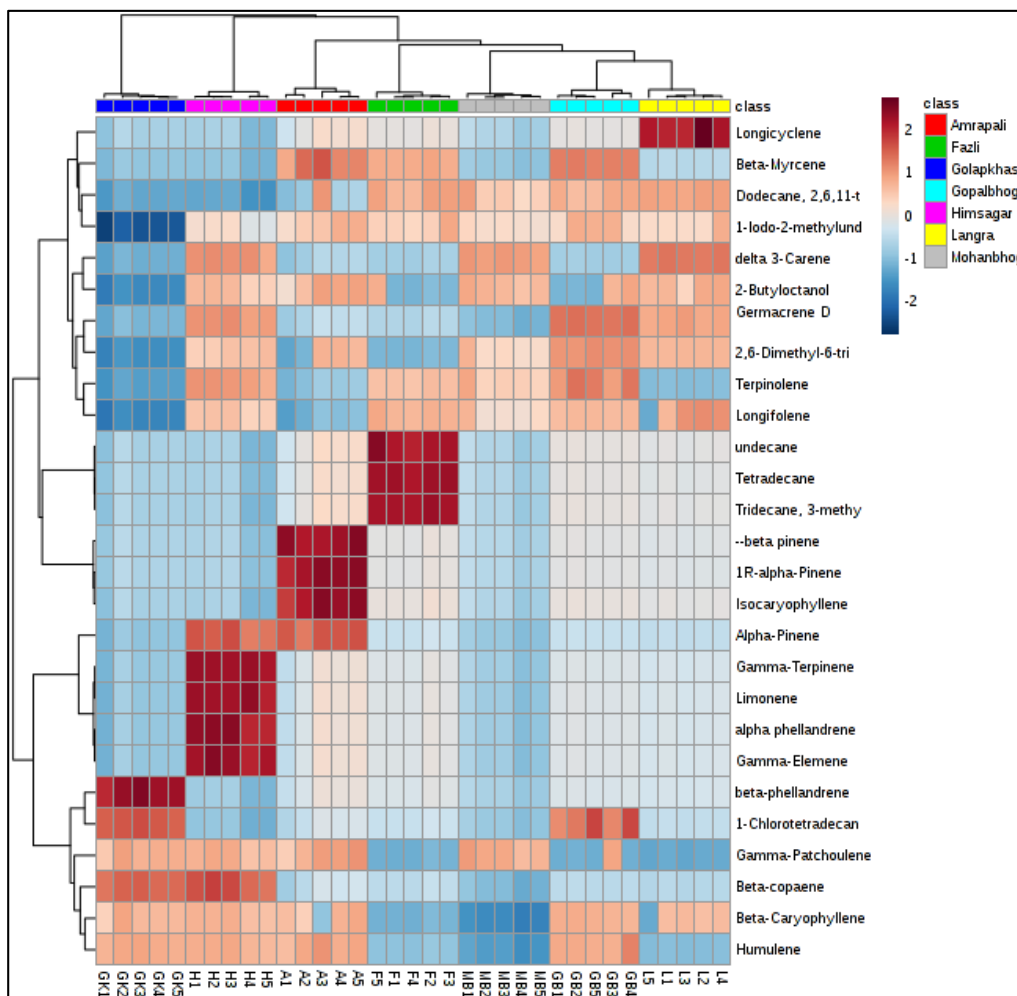


Fig 1: Heatmap showing differences in relative response ratios of identified non-polar components from the pulp of the seven varieties of *M. indica*

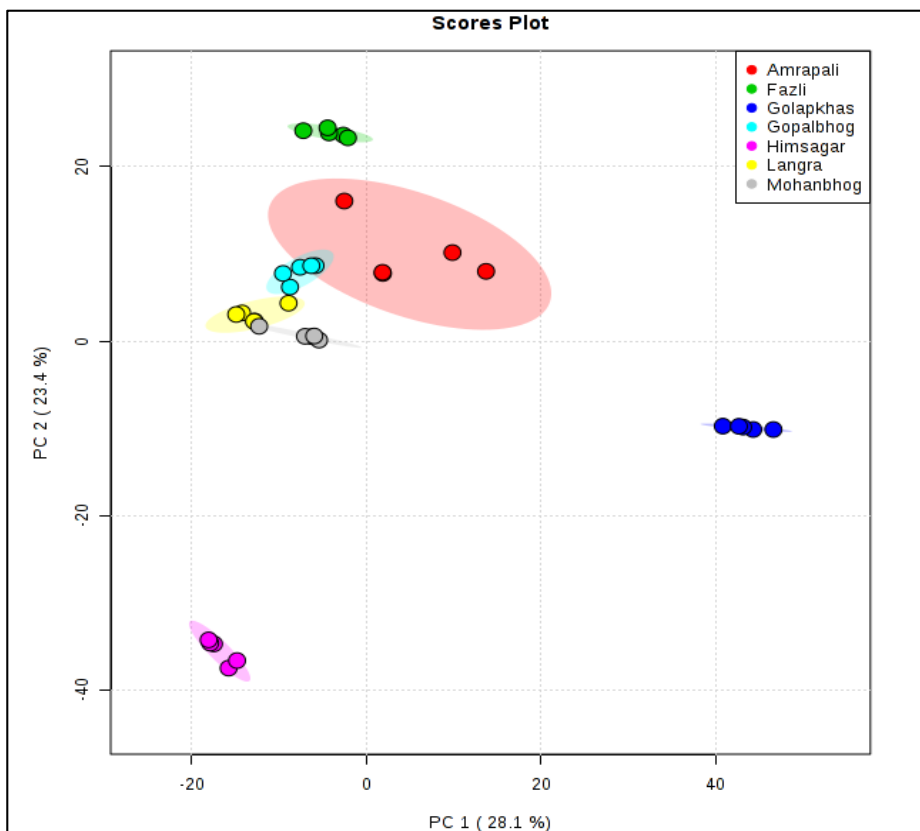
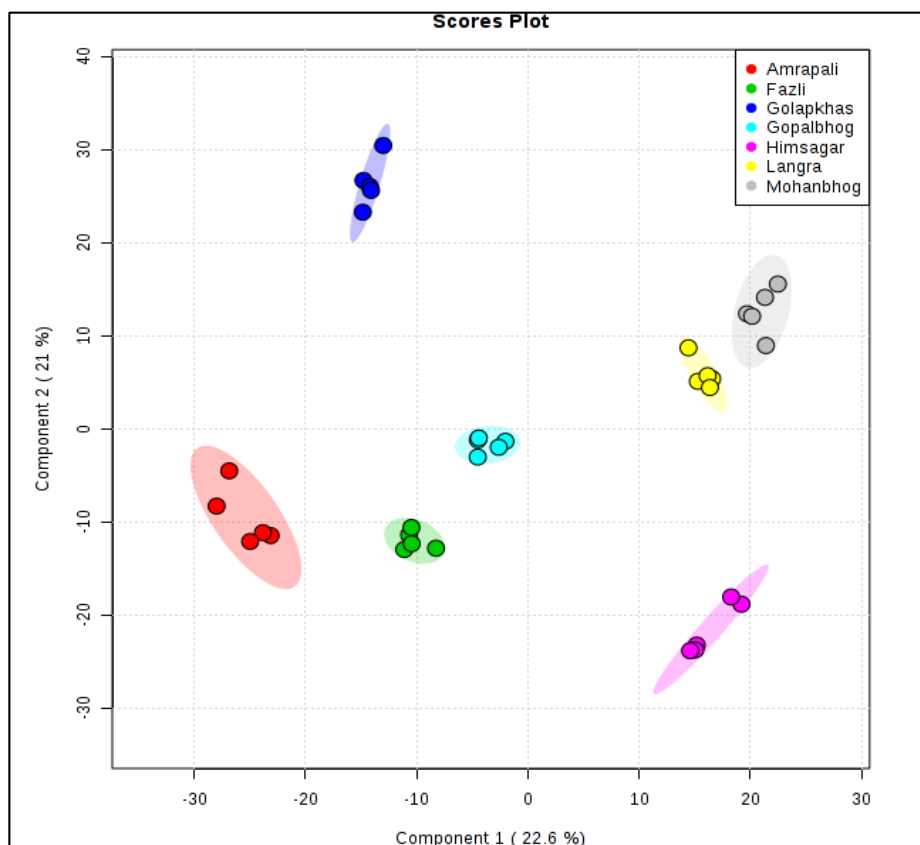


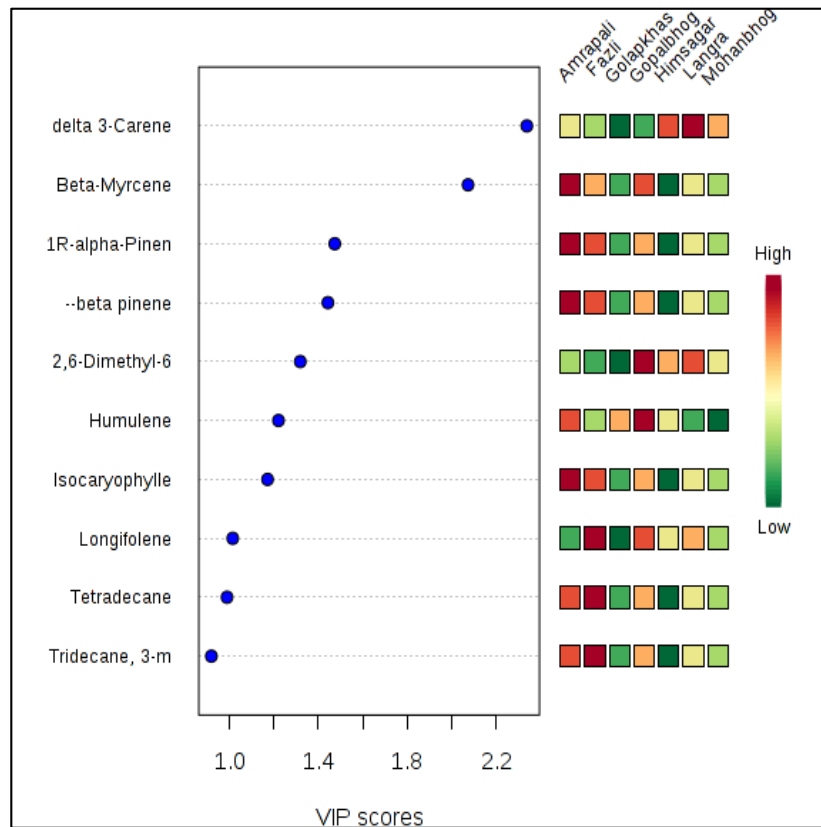
Fig 2: Multivariate analysis of identified metabolites in PCA score plot



PLS-DA cross validation details

| Measure  | 1 comps | 2 comps | 3 comps | 4 comps | 5 comps |
|----------|---------|---------|---------|---------|---------|
| Accuracy | 0.0     | 0.14286 | 0.48571 | 0.42857 | 0.94286 |
| R2       | 0.90773 | 0.97246 | 0.98945 | 0.99115 | 0.99208 |
| Q2       | 0.87679 | 0.95636 | 0.98247 | 0.98488 | 0.98527 |

Fig 3: Multivariate analysis of identified metabolites in PLS-DA score plot



**Fig 4:** 10 most influential metabolites for varietal separation represented in VIP plot

### Conclusion

Hexane soluble non-polar metabolites of seven varieties of Indian mango pulp were analyzed by GC-MS. On the basis of multivariate statistical analysis of the identified metabolites, the varieties could be distinctly differentiated from each other on the basis of four most important aromatic components such as delta-3-carene, beta-myrcene, 1R-alpha-pinene, beta-pinene.

### Acknowledgement

The authors acknowledge financial support from University Grants Commission (CAS Programme) and Department of Science and Technology, Government of India (FIST Programme).

### References

- Pino AJ, Queris O. Analysis of volatile compounds of mango wine. *Food Chem.* 2011; 125:1141-1146.
- Hassan FA, Al-Sheraji SH, Ismail A. Dried mangoes: antioxidant properties, and health benefits. In *Dried Fruits: Phytochemicals and health effects*. Alasalvar, C., Shahidi, F. (ed.) Chichester: John Wiley and Sons Inc., 2013, 457-470.
- Macleod AJ, Troconis NG. Volatile flavor components of mango fruits. *Phytochemistry.* 1982; 21(10):2523-2526.
- Etievant PX. Volatile compounds in food and beverages. In Maarse, H. (Ed.). New York: Marcel Dekker, 1991.
- Zhang Z, Zeng D, Li G. The study of aroma characteristics of Chinese mango cultivars by GC-MS with solid phase microextraction. *Journal of Plant Science.* 2006; 1:98-105.
- Helena E, Andrade A, Maia JGS, Gracas M, Zoghbi B. Aroma volatile composition and analysis. *J Food Compos. Anal.* 2000; 13:27-33.
- Lalel HJD, Singh Z, Tan SC. Distribution of aroma volatile compounds in different parts of mango fruit. *The*

*Journal of Horticultural Science and Biotechnology.* 2003; 78:131-138.

- Lalel HJD, Singh Z, Tan SC. Maturity stages at harvest affects fruit ripening, quality and biosynthesis of aroma volatile compounds in 'Kensington Pride' mango. *J Hort Sci Biotechnol.* 2003; 78:225-233.
- Singh Z, Lalel HJD, Nasir S. A review of mango fruit aroma volatile compounds: state of the art research. *Acta horticulturae.* 2004; 645:519-527.
- Laohaprasit N, Ambadipudi DS, Srzednicki G. Optimization of extraction conditions of volatile compounds in 'Nam Dok Mai' mangoes. *Int Food Res J.* 2011; 18:1043-1049.
- El Hadi MAM, Zhang FJ, Wu FF, Zhou CH, Tao J. Review advances in fruit aroma volatile research. *Molecules.* 2013; 18:8200-8229.
- Vahedi H, Lari J, Halimi M, Nasrabadi M, Vahedi A. Chemical composition of the n-hexane extract of *Verbascum speciosum* growing wild in Iran. *J Essential oil Bearing Plants.* 2012; 15(6):895-899.
- Nyegue M, Zollo PA, Bessiere JM, Rapior S. Volatile components of fresh *Pleurotus ostreatus* and *Termitomyces shimperi* from Cameroon. *J Essential oil Bearing Plants.* 2003; 6(3):153-160.
- Karak S, Bhattacharya P, Nandy A, Saha A, De B. Metabolite profiling and chemometric study for varietal variance in *Piper betle* L. leaf. *Current Metabolomics.* 2016; 4:129-140.
- Adams P. Identification of essential oil components by gas-chromatography/mass spectrometry. 4th ed, Allured Publishing: Carol Stream, IL, 2009.
- Baranauskiene R, Venskutonis RP and Demyttenaere JCR. Sensory and instrumental evaluation of catnip (*Nepeta cataria* L.) aroma. *J Agric. Food Chem.* 2003; 51(13):3840-3848.

17. Skaltsa HD, demetzos C, Lazari D, Sokovic M. Essential oil analysis and antimicrobial activity of eight *Stachys* species from Greece. *Phytochemistry*. 2003; 64(3):743-752.
18. Regnier FE, Nieh M, Holldobler B. The volatile dufour's gland components of the harvester ant *Pogonomyrmex rugosus* and *P. barbatus*. *J Insect Physiol*. 1973; 19(5):981-992.
19. White IR, Blake RS, Taylor AJ, Monks PS. Metabolite profiling of the ripening of mangoes *Mangifera indica* L. cv. 'Tomy Atkins' by real-time measurement of volatile organic compounds. *Metabolomics*. 2016; 12:57.
20. Oliveira GB, Costa HB, Ventura JA, Kondratyuk TP, Barroso MES, Correia RM *et al.* Chemical profile of mango (*Mangifera indica* L.) using electrospray ionization mass spectrometry (ESI-MS). *Food Chem*. 2016; 204:37-45.
21. De Pradhan I, Dutta M, Choudhury K, De B. Metabolic diversity and *in vitro* pancreatic lipase inhibition activity of some varieties of *Mangifera indica* L. fruits. *International Journal of Food Properties*. 2017; 20:S3212-323.