



E-ISSN: 2278-4136
P-ISSN: 2349-8234
JPP 2019; 8(2): 1333-1337
Received: 04-01-2019
Accepted: 06-02-2019

HT Valvi
Department of Plant Pathology,
College of Agriculture, Dr.
BSK KV, Dapoli, Maharashtra,
India

AD Saykar
Department of Plant Pathology,
College of Agriculture, Dr.
BSK KV, Dapoli, Maharashtra,
India

VR Bangar
Department of Plant Pathology,
College of Agriculture, Dr.
BSK KV, Dapoli, Maharashtra,
India

In vitro and *in vivo* field efficacy of different fungicides against *Alternaria brassicae* (Berk.) sacc. causing *Alternaria* leaf spot of cauliflower

HT Valvi, AD Saykar and VR Bangar

Abstract

The *Alternaria* leaf spot caused by *A. brassicae* (Berk.) Sacc is one of the destructive disease of Cauliflower in Konkan region of Maharashtra. In the present study, bio efficacy of seven fungicides tested under *in vitro* conditions against the pathogen. Among that, Mancozeb 75% WP (0.25 %) was completely inhibited the growth of the test fungus. It was followed by Propiconazole 25% EC (0.1%) which showed 96.29 per cent inhibition of the test fungus and was at par with Mancozeb. No sporulation was observed in Mancozeb 75% WP, Propiconazole 25% EC and Difenconazole 25% EC. Among the different fungicides tested under field conditions, all the fungicidal treatments were significantly superior over control after last spray. The treatment Mancozeb 75% WP was the most effective as it recorded minimum disease incidence (28 %). It was followed by Azoxystrobin 23% EC which recorded 29 % disease incidence.

Keywords: Cauliflower, *Alternaria brassicae* (Berk.) Sacc. Fungicides, Management, Sporulation and Mycelial growth

Introduction

Several factors are responsible for low production of cauliflower crop, among which diseases also play an important role. The important diseases of Cauliflower crop are leaf spot, Downy mildew, Damping off, Club root, Powdery mildew, White rust, Black rot, Bacterial soft rot and Cauliflower mosaic. Among these diseases *Alternaria* leaf spot is a serious disease of cauliflower. About 20 to 80 per cent loss in yield and 59 per cent loss in seed may occur due to this disease. The disease appears as minute specks on the leaves, which enlarge over a time and result in substantial lesions with concentric rings where spores are produced. The disease starts from lower leaves and slowly progresses towards the upper shoots, leaves, petioles, pods and heads. Defoliation of the outer leaves may occur on severely infected plants and extensive trimming may be required to remove infected leaves from the cabbage head at harvest. In susceptible varieties, apart from yield, significant reduction in quality may occur.

During recent years the fungal diseases were found to occur in severe proportions under favorable environmental conditions particularly during *Rabi* season. *Alternaria* leaf spot caused by *Alternaria brassicae* has been reported to inflict heavy yield losses (Pattanamahakul and Strange, 1999; Azevedo *et al.*, 2000; Peruch *et al.*, 2006; Chauhan *et al.*, 2009; Mishra *et al.*, 2009; Deep and Sharma, 2012 and Sharma *et al.*, 2013) [14, 1, 15, 2, 11, 4, 17].

Alternaria leaf spot is difficult to control because the fungus penetrates in infected seed, plant debris in soil and collateral hosts such as other cruciferous crops in the vicinity of the main crop or weeds. One of the most effective and old method for disease control is the use of fungicides. There are several fungicides which are being commercially available while several others are being evaluated in different laboratories. Even though cauliflower is not a regularly cultivated crop in Konkan region, some progressive vegetable growers are switching to this crop due to its high marketability. Considering importance of the crop and disease, present study on *Alternaria* leaf spot of cauliflower was planned and conducted to evolve *in vitro* and *in vivo* management strategy.

Materials and Methods

In vitro evaluation of fungicides

The laboratory experiments in the present study were conducted in the Department of Plant Pathology, Dr. DBSKKV, Dapoli during 2015-16. The principle involved in poisoned food technique is to poison the nutrient medium with a fungi-toxicant and then allowing the test fungus to grow on it and finally recording the extent of growth (Nene and Thapliyal, 1997) [13]. The details of treatments used are mentioned below.

Correspondence

VR Bangar
Department of Plant Pathology,
College of Agriculture, Dr.
BSK KV, Dapoli, Maharashtra,
India

Potato dextrose agar medium (PDA) was used as basal medium and dispensed in 100 ml aliquots in each 250 ml Erlenmeyer conical flasks, which were sterilized at 1.054 kg/cm² pressure for 20 minutes. The quantity of every fungicide for each concentration was calculated for 100 ml medium separately. The weighed quantity of each fungicide was added in lukewarm PDA at 40 ± 2°C, mixed thoroughly and poured into sterilized Petri plates and allowed to solidify. The mycelial discs of 5 mm diameter were cut from 7 day old culture of the test fungus with the help of a sterilized cork borer. A single disc was transferred aseptically to the centre of each plate already poured with poisoned medium. The plates with PDA without fungicide but inoculated with fungal culture, served as control.

List of fungicides tested under *in vitro* and *in vivo* against the pathogen.

T. No.	Name of the fungicide and formulation	Trade name	Conc. (%) tested
T ₁	Azoxystrobin 23% EC	Amistar	0.1
T ₂	Carbendazim 50% WP	Fungiguard	0.1
T ₃	Copper hydroxide 77% WP	Kocide 101	0.1
T ₄	Mancozeb 75% WP	Dithane M-45	0.25
T ₅	Difenconazole 25% EC	Score	0.1
T ₆	Thiophanate methyl 70% WP	Roko	0.1
T ₇	Propiconazole 25% EC	Tilt	0.1
T ₈	Control	-	-

The plates were incubated at room temperature (27 ± 2°C). Three replications of each treatment were maintained. The observation on colony diameter of the fungus were recorded when Petri plate in control treatment was fully covered with mycelial growth. Per cent inhibition of growth of the test pathogen was calculated.

Per cent inhibition of growth of the test fungus was calculated by following formula (Horsfall, 1956) [7].

$$X = \frac{Y - Z}{Y} \times 100$$

Where, X = Per cent inhibition, Y = Growth of fungus in control (mm), Z = Growth of fungus in treatment (mm)

Field evaluation of fungicides

The field experiment was conducted on variety "Super-fast crop" of cauliflower during *Rabi* 2015-16 at the Botany farm, College of Agriculture, Dapoli.

Raising of cauliflower seedlings

The seeds of cauliflower were sown on in portraits containing cocopeat as growing medium. The trays were staked one above the other and the uppermost tray was covered with a clean plastic sheet for 4 days. Seeds germinated within 4 days and then each tray was arranged separately on raised beds in shed net. The trays were watered regularly to maintain sufficient moisture. The Cauliflower seedlings were ready for transplanting in 25 days.

Preparation of experimental plot

The experimental plot was prepared with a tractor drawn mould board plough by two crisscross ploughings with tractor operated cultivator. Planking was done to level the field. The experiment was laid in randomized block design. Three replications were maintained per treatment.

Transplanting of seedlings

Initially light irrigation was given to the experimental plot in the morning so as to achieve optimum moisture in the soil to facilitate establishment of seedling during transplanting. On the same day evening, 25 days old healthy seedling were carefully uprooted and immediately transplanted in the field. Light irrigation was given to the plot immediately after transplanting for better crop stand.

Schedule of spraying of Fungicide:

The crop was observed carefully for initiation of the disease. First spray of each fungicide treatment was given immediately after disease incidence was noticed. Two more sprays of fungicides were given at an interval of 15 days.

Method of recording observations:

Five plants per treatment per replication were randomly selected for recording disease incidence of *Alternaria* leaf spot. Initial observations were recorded before first spray and final observations were recorded 15 days after the last spray on the basis of development of spots and lesions by considering the whole leaf area as hundred per cent.

Observations on disease incidence:

The disease intensity was recorded in 0-5 scale (Conn. *et al.* 1990) as described below.

Score/grade	Description
0	No disease
1	<5% leaf area affected
2	6-10% leaf area affected
3	11-25% leaf area affected
4	26-50% leaf area affected
5	>50% leaf area affected

Per cent disease intensity (PDI):

Per cent disease intensity was calculated by the following formula given by (McKinney, (1923)

$$PDI = \frac{\text{Sum of all numerical ratings}}{\text{Total number of leaves examined} \times \text{Maximum rating}} \times 100$$

Per cent disease control (PDC)

The per cent disease control was calculated by using the formula given below:

$$PDC = \frac{PDI \text{ in control} - PDI \text{ in treatment}}{PDI \text{ in control}} \times 100$$

Statistical analysis

The data obtained were statistically analysed by the methods suggested by Gomez and Gomez (1986) [6]. The standard error and critical difference were worked out and the results obtained were compared statistically.

Results and Discussion

In vitro evaluation of fungicides

Seven fungicides belonging to different groups were tested for their efficacy against *A. brassicae* by employing poisoned food technique. The data obtained on the effect of different fungicides on the vegetative growth and sporulations of *A. brassicae in vitro* are presented in Table 1, PLATE-I and depicted in Fig-1.

The data presented in Table 1 revealed that, among the different fungicides tested under *in vitro* Mancozeb 75% WP (0.25 %) completely inhibited the growth of the test fungus. It was followed by Propiconazole 25% EC (0.1%) which showed 96.29 per cent inhibition of the test fungus and was at par with Mancozeb. The next fungicide in order of merit was Difenconazole 25% EC (0.1 %) which recorded 87.77 per cent inhibition. Copper hydroxide 77% WP (0.1 %) and

Azoxystrobin 23% EC (0.1 %) showed 52.96 per cent and 49.25 per cent inhibition of *A. brassicae*, respectively over control and were at par with each other. In the treatment of Thiophanate methyl 70% WP (0.1%), 38.88 per cent inhibition of the fungus was achieved. Carbendazim 50% WP (0.1) was appeared as least effective fungicide which showed only 27.77 % inhibition of the test pathogen.

Table 1: *In vitro* efficacy of fungicides on growth and sporulation of *A. brassicae* (Berk.) Sacc.

T. No.	Name of the fungicide	Conc. (%)	Mean colony diameter (mm)	Per cent inhibition over control	Sporulation
T ₁	Copper hydroxide 77% WP	0.1	42.33	52.96	+
T ₂	Azoxystrobin 23% EC	0.1	45.66	49.25	+
T ₃	Difenconazole 25% EC	0.1	11.00	87.77	-
T ₄	Propiconazole 25% EC	0.1	3.33	96.29	-
T ₅	Mancozeb 75% WP	0.25	0.00	100	-
T ₆	Thiophanate methyl 70% WP	0.1	55	38.88	++
T ₇	Carbendazim 50% WP	0.1	86.66	3.70	+++
T ₈	Control		90.00	-	++++
S.Em.±			0.91		
C.D.at 1%			3.85		

Sporulation

- = No sporulation, +++ = Good,
 + = Poor, ++++ = Excellent.
 ++ = Fair,

No sporulation was observed in Mancozeb 75% WP (0.25%), Propiconazole 25% EC (0.1%) and Difenconazole 25% EC (0.1%). Poor sporulation was recorded in Copper hydroxide 77% WP (0.1%) and Azoxystrobin 23% EC (0.1%). Fair sporulation was observed in Thiophanate methyl 70% WP (0.1%). Good sporulation was observed in Carbendazim 50% WP (0.1%) as compared to control which showed excellent sporulation. The result findings are in agreement with those reported by Kumar *et al.*, (1999) ^[9] who reported that Carbendazim 0.05 and 0.1 per cent completely inhibited mycelial growth of *A. brassicae*. Beatrice *et al.*, (2004) showed that difenconazole was most effective against *A.*

brassicae and *A. brassicicola*. Hossain and Mian (2004) reported that Mancozeb, Carbendazim and Propiconazole inhibited the mycelial growth of the *A. brassicicola* infecting cabbage. Survilieue *et al.*, (2006) reported that Azoxystrobin showed significant inhibitory effect on mycelial growth of the *Alternaria* spp. Mishra *et al.*, (2009) ^[11] reported that Propiconazole was the most effective against *A. brassicae* which showed maximum per cent inhibition of mycelial growth and was followed by Mancozeb and Azoxystrobin. Gaikwad (2013) also reported that Propiconazole, Difenconazole and Mancozeb at different concentrations completely inhibited the mycelial growth of *A. brassicae*.



Plate 1: Efficacy of fungicides on growth and sporulation of *Alternaria brassicae*

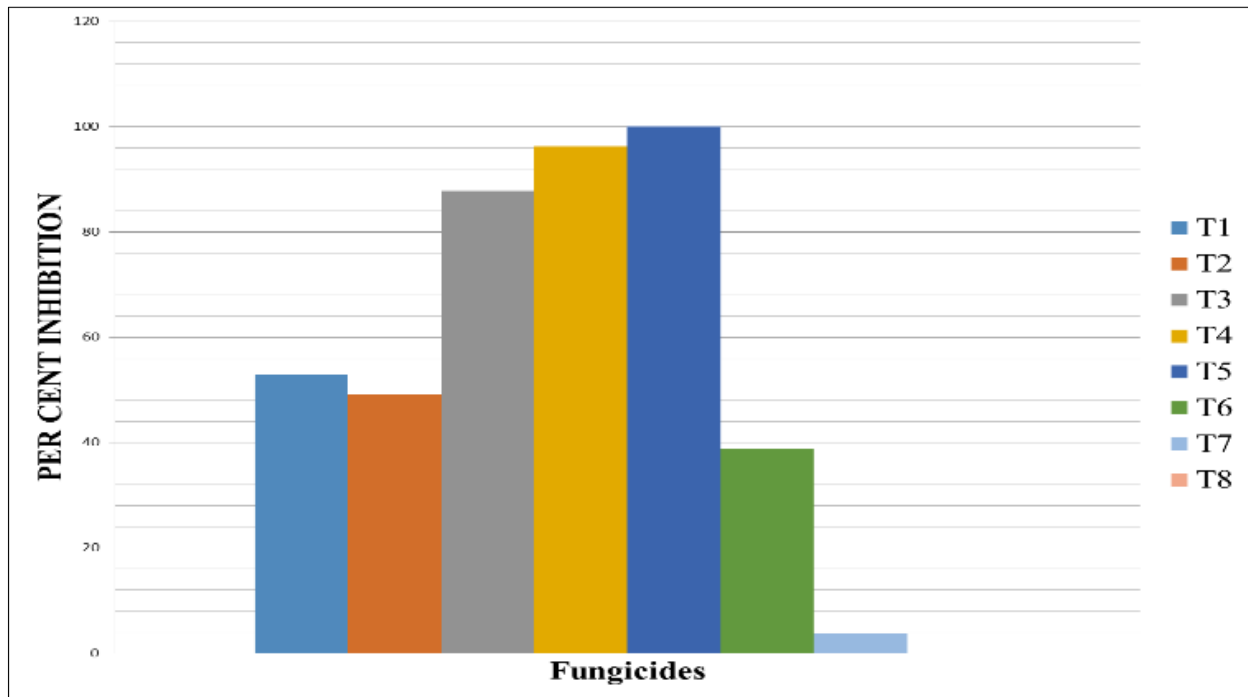


Fig 1: Efficacy of fungicides on growth and sporulation of *A. brassicae* (Berk.) Sacc.

Management of the disease in field conditions

The results of the experiment on evaluation of the fungicides

against the disease under field conditions are presented in Table 2 and depicted in Fig-2.

Table 2: Per cent disease incidence (PDI) and per cent disease control (PDC) against *A. brassicae* (Berk.) Sacc.

T. No.	Treatments	Conc. (%)	Mean PDI before first spray	Mean PDI after last spray	PDC
T ₁	Copper hydroxide 77% WP	0.1	19.60 (26.25)*	36.83 (37.36)	42.45
T ₂	Azoxystrobin 23% EC	0.1	19.67 (26.32)	29.00 (32.58)	54.69
T ₃	Difconazole 25% EC	0.1	20.17 (26.61)	33.50 (35.35)	47.66
T ₄	Propiconazole 25% EC	0.1	19.83 (26.41)	31.67 (34.24)	50.52
T ₅	Mancozeb 75% WP	0.25	19.75 (26.38)	28.00 (31.93)	56.25
T ₆	Thiophanate methyl 70% WP	0.1	19.83 (26.45)	36.00 (36.86)	43.75
T ₇	Carbendazim 50% WP	0.1	20.00 (26.56)	34.00 (35.67)	46.88
T ₈	Control		20.15 (26.67)	64.00 (53.13)	
S. Em.±			0.54	0.63	
C.D.at 5%			1.64	1.91	

(*Figures in parentheses indicate arc sin values)

It is apparent from the data presented in Table 2 that, among the different fungicides tested under field conditions all the fungicidal treatments were significantly superior over control after last spray. The treatment T₅ (Mancozeb 75% WP) was the most effective as it recorded minimum disease incidence (28 %). It was followed by T₂ (Azoxystrobin 23% EC) which recorded 29.00 per cent disease incidence. The treatment T₄ (Propiconazole 25% EC) was at par with T₃ (Difconazole 25% EC). Treatment T₇ (Carbendazim 50% WP) showed 34 % disease incidence and it was followed by T₆ (Thiophanate methyl 70% WP) and T₁ (Copper hydroxide 77% WP) with 36 and 36.63 per cent disease incidence respectively over control (64 %). Maximum per cent disease control was achieved in treatment T₅ (Mancozeb 75% WP) and T₂ (Azoxystrobin 23% EC) which showed 56.25 per cent and 54.69 per cent disease control over control, respectively. Both the treatments were found statistically at par with each other. The next fungicides in merit of order were treatment T₄ (Propiconazole 25% EC) & T₃ (Difconazole 25% EC)

which recorded 50.52 per cent and 47.66 per cent disease control, respectively.

These were followed by treatment T₇ (Carbendazim 50% WP) with 46.88 per cent disease control and was at par with T₄. Treatment T₆ (Thiophanate methyl 70% WP) and T₁ (Copper hydroxide 77% WP) showed 43.75 and 42.45 per cent disease control over control treatment, respectively and were statistically at par with each other. Sreedhar *et al.*, (2003) [18], showed that Azoxystrobin was the most effective fungicides as it reduced disease incidence to the tune of 88 to 93 per cent. Narain *et al.*, (2006) [12] reported that Mancozeb was the most effective fungicide against *A. brassicae*. Prasad and Lallu (2006) reported that first spray of Carbendazim (0.1%) + Mancozeb (0.2%) followed by two sprays of Mancozeb (0.2%) at early date of sowing (October) was the best combination in reducing the disease severity on leaves. Gaikwad (2013) showed that Mancozeb showed the least diseases incidence (15.30%) followed by Propiconazole (18.36%) and Copper oxychloride (20.65%).

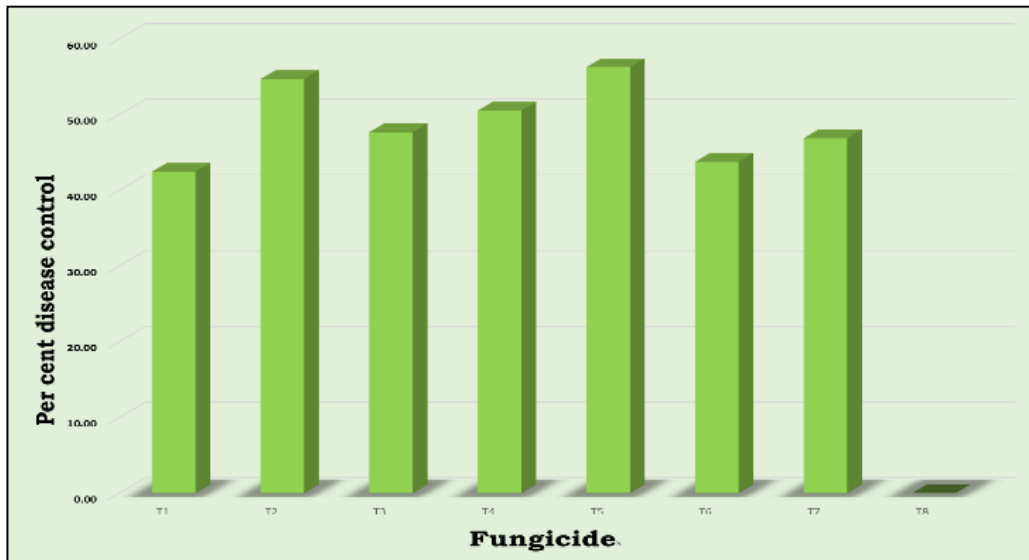


Fig 2: Management of the disease in the field by using different fungicides.

Conclusion

On the basis of the results of present study it can be concluded that *Alternaria* leaf spot of cauliflower caused by *Alternaria brassicae* (Berk.) Sacc. is an important disease of cauliflower in Konkan region. The fungicides viz., Mancozeb (0.25%) and Azoxystrobin (0.1%) are very effective against the pathogen under field conditions. Three sprays of Mancozeb (0.25%) or Azoxystrobin (0.1%) at an interval of 15 days will be the best strategy for management of *Alternaria* leaf spot of cauliflower. However, these findings need to be confirmed at different locations.

References

- Azevedo SS, Mariano R, De LR, Michereff SI. Survey of intensity of black rot and *Alternaria* leaf spot cabbage in Agreste in Pernambuco and determination of sample size for disease quantification. *Summa phytopathologica*. 2000; 26(3):299-306.
- Chauhan JS, Anoop B, Indrakumar N, Singh Ali Seema. Effect of *Alternaria* on some members of family Brassicaceae of Garhwal Himalaya. *New York S. J.* 2009; 2(6):80-85
- Conn KL, Tewari JP, Awasthi RP, A disease assessment key for *Alternaria* black spot in rapeseed and mustard. *Canadian Pl. Dis. Serv.* 1990; 70:19-22.
- Deep Swati, Sharma Pratibha. Host age as predisposing factor for incidence of black leaf spot of cauliflower caused by *Alternaria brassicae* and *Alternaria brassicicola*. *Indian phytopath.* 2012; 65(1):71-75.
- Gaikwad PA. Studies on Leaf Spot of Cabbage Caused by *Alternaria brassicae* (Berk.) Sacc. M.Sc. (Ag) Thesis, submitted to V.N.M.K.V., Parbhani, Maharashtra, 2013.
- Gomez KA, Gomez AA. *Statistical Procedures for Agricultural Research*. 2nd ed. John Wiley and Sons, London, 680, 1986.
- Horsfall JG Principles of fungicidal action. *Cronical boronica* (U.S.A.), 1956, 207.
- Hossain MS, Mian IH. Effect of foliar fungicides on the control of *Alternaria* blight of Cabbage seed crop. *Bangladesh J Pl. Pathol.* 2005; 20:43-48.
- Kumar A, Jha AK, Sinha SK, Ojha KL. Evaluation of different fungicides for the control of *Alternaria* leaf blight of Cauliflower. *J App. Biol.* 1999; 9(1):77-80.
- Mackinney. A new system of grading plant disease. *J Agric. Res.* 1923; 26:195-218.
- Mishra PK, Rai RK, Saha S, Pandey KK, Singh RP, Rai AB. Effect of fungicides, botanicals and bioagent leaf spot of cauliflower caused by *Alternaria brassicae* (Berk.) Sacc. *Vegetable Science.* 2009; 36(3):55-59.
- Narain U, Chand G, Pandey R. Efficacy of fungicides against *Alternaria* leaf spot of broccoli. *Annals of Plant Protection Sciences.* 2006; 14(2):487-488.
- Nene YL, Thapliyal PN. *Fungicides in plant disease control*. Oxford and IBH Publishing Co., New Delhi, 1997, 531-532.
- Pattanamahakul P, Strange RN. Identification and toxicity of *Alternaria brassicicola*, the causal agent of dark leaf spot disease of *Brassica* species grown in Thailand. *Plant Pathology.* 1999; 48(6):749-755.
- Peruch LAM, Michereff S, Araujo IB. Survey of intensity of *Alternaria* black spot and black rot on brassica species under organic farming systems in Pernambuco and anta catarina states, Brazil. *Horti. Bras.* 2006, 24(4).
- Prasad, R. and Lallu. 2006. Management of *Alternaria* blight of Mustard with Combination of Chemicals and Botanicals. *Annals of Plant Protection Sciences.*, 14(1)400-403.
- Sharma P, Deep S, Sharma M, Bhati DS. Genetic variation of *Alternaria brassicae* (Berk.) Sacc. causing dark leaf spot of Cauliflower and Mustard in India. *J Gen. Plant Pathology.* 2013, 79:41-45.
- Sreedhar KN, Ashokkumar CT, Padamabha K, Sudhir Kumar AS. Compatibility of insecticides and fungicides mixtures against cabbage leaf spot, *Alternaria brassicae* (Sacc.) Berk. *The Asian Journal of Horticulture*, 2003; 8:659-666.
- Surviliene, E. and Dambrauskiene, E. Effect of Different active Ingredients of Fungicides on *Alternaria spp.* growth *in vitro*. *Agronomy Research*, 2006, 403-406.
- Thaware DS, Fugro PA, Jadhav YT, Magar SV, Karande RA, *In vitro* evaluation of different fungicides, plant extracts and bio-agents against *Alternaria alternata* (Fr.) Keissler causing leaf blight of cowpea. *International Journal of Plant.* 2010; 3(2):356-360.