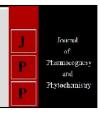


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A review on *Chaetomium globosum* is versatile weapons for various plant pathogens

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Abstract

Chaetomium globosum is a potential bio control agent against various seed and soil borne pathogens. Plant pathogens are the main threat for profitable agricultural productivity. Currently, Chemical fungicides are highly effective and convenient to use but they are a potential threat for the environment. Therefore the use of biocontrol agents for the management of plant pathogens is considered as a safer and sustainable strategy for safe and profitable agricultural productivity. Many experiments and studies revealed by various researchers *C. globosum* used as plant growth promoter and resulted into high yield of crops in field conditions. *C. globosum* produces pectinolytic enzymes polygalacturonate transeliminase (PGTE), pectin trans-eliminase (PTE), viz., polygalacturonase (PG), pectin methyl esterase (PME), protopectinase (PP), xylanase and cellulolytic (C and C) 1 xenzymes and various biologically active substances, such as chaetoglobosin A, Chaetomium B, C, D, Q, R, T, chaetomin, chaetocin, chaetochalasin A, chaetoviridins A and C. The present aim of this article we have discussed the various aspects of biocontrol potential of *Chaetomium globosum*.

Keywords: Biological control, Chaetomium globosum, plant pathogens

Introduction

Chaetomium globosum is so far commonest and most cosmopolitan fungi especially on plant remains, seeds, compost paper and other cellulosic substrates. (Domsch *et al.* 2007) ^[7]. It is a common colonizer of soil and cellulose producer with ability to degrade cellulosic and other organic materials, it is one of the commonest species growing saprophytically in the rhizosphere and phyllosphere. It has been reported to be a potential bio-control agent suppresses the growth of bacteria and fungi through competition, mycoparasitism, antibiosis, or their various combinations (Zhang & Yang 2007). There are many studies with promising results on using endophyte *Chaetomium* spp. as a biocontrol agent. Endophytes are capable to reduce in the host the effect of fungi diseases, through secondary metabolites production as alkaloids. (Poulina Moya *et al.* 2016) ^[14]. Antagonistic mechanisms of this fungus include competition for space and nutrients (Vannaci & Harman 1987) ^[23], mycoparasitism (Mandal *et al.*, 1999) and metabolite production (antibiosis) such as chaetomin, chaetoglobosin, cochliodinol, chaetosin and prenisatin (Brewer *et al.*, 1970; Brewer *et al.*, 1972; Brewer & Taylor, 1978.

Moreover, the hazardous use of heavy chemical fungicide caused environmental pollution and build up of chemical residues in the air, soil, water and agricultural products. Recently, the biological control of plant pathogens has been taking place and it has served as a new strategy for disease control (Soytong, 1995) [18]. This successful use of biological control measure particularly reduce the chemical usage and improved agro ecosystem for sustainable agriculture and has maintained ecological balance. It was found that using the specific strain of *C. globosum* Kunze could control many plant pathogens, for example *C. globosum* was shown to be antagonistic to *Fusarium* spp. and *Helminthosporium* spp. (Tveit and Moore, 1954) [19] and was antagonistic to *Alternaria brassicicola* (Vannacci and Harman, 1987) [23], and reduced the inoculum of *Botrytis cinerea* (Kohl *et al.*, 1995). In many years of research work strain of *C. globosum* have been screened and found to control other economically important plant pathogens like *Phytopthora palmivora*, *Phytopthora parasitica*, and *Colletotrichum gloeosporioides* (Soytong, 1991).

Morphology of Chaetomium globosum

Chaetomium is a genus of the class Pyrenomycetes (Ascomycotina), Order Sordariales and family Chaetomiaceae. It is a dematiaceous (dark-walled) mold normally found in soil, air, and plant debris. There are about 95 species in the widespread genus (Kirk *et al.* 2008) [11]. Members of this genus typically have superficial, ostiolar perithecia, covered in hairs.

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Horticulture Polytechnic College, Sri Konda Laxman Telangana State Horticultural University, Telangana, India Asci are often clavate and evan evanescent, bearing eight spores. *Chaetomium* colonies grow rapidly, they have a cottony appearance and are initially white in colour, the mature colonies becoming grey to olive, and later sometimes tan to red or brown to black.

Microscopic observation of *Chaetomium globosum* can produce an Acremonium-like state (imperfect stage) on culture media and it is characterized by superficial flask-shaped perithecia, which are surrounded by dark and stiff hair. Asci are often clavate and evanescent, bearing eight spores. Ascospores are usually lemon-shaped, commonly coloured olive-brown. Perithecia showed densely haired surfaces and the shape of perithecia varied from globose to subglobose. The ascospores released inside the perithecium were seen oozing out from ostiole. The results were much similar to those reported by Ahammed *et al.* (2005), Prokhorov *et al.* (2011), Maheswari *et al.* (2013).

Growth and sporulation of *Chaetomium globosum* in synthetic and non synthetic media

Chatterji (1966) studied that Chaetomium globosum, grown at 30 °C on media in which the source of nitrogen is ammonium tartrate, produces no mature perithecia, although perithecial initials appear freely. Maturation of initials occurs at 26 °C, or in the presence of certain other fungi. Organic phosphates, biotin and other substances also affect the development of perithecia. Domsch et al. (2007) [7] revealed that good sporulation of most Chaetomium globosum was obtained on substrates with a high C/N ratio. Therefore the microscopic characters were recorded mainly for colonies on cornmeal agar. In case of Neeta Sharma et al. (2011) revealed that growth and sporulation of Chaetomium globosum was excellent on Malt yeast extract agar medium(64.96mm) followed by Nutreint yeast agar medium (27.53mm) while poor growth was recorded Czapeks dox agar medium (11.56mm) and Richard medium (7.8mm) Kiran et al. (2015) studied the growth and sporulation of Chaetomium globosum was excellent on sabouraud medium(90 mm) because glucose or maltose and peptone were the most suitable carbon and nitrogen source for Chaetomium growth followed by PDA (63 mm), while minimum growth was observed on Czapeks dox agar medium. Sharma et al. (2010) who studied colony diameters of C. globosum over 4 weeks on different agar media viz., potato dextrose agar (PDA), oatmeal agar (OA), cornmeal agar (CMA) and malt extract agar (MEA) and found that oatmeal agar exhibited comparatively higher mycelial growth and sporulation. Poulina Moya et al. (2016) [14] In oatmeal agar the isolates showed olivaceous-yellow colonies according to the color of Isolate C5 showed globose or oval ostiolar ascomata with straight or undulate hairs unlike isolate 2, which showed irregular hairs with no undulations. Both isolates showed terminal hairs all unbranched and displayed one-celled, lemon-shaped ascospores, In PDA isolate C5 was slightly yellower than C2 and showed margins wavy or irregular, while C2 developed full edges. The growth on the several media did not evidence major differences. It was slightly higher on OA followed by MEA. At 7th day on the three media, C2 grew faster than C5. Andrez fojutowski et al. (2015) [2] Growth of Chaetomium globosum fungus in laboratory conditions on salt-agar medium quiet fast and growth rate of 5, 7, day is 29mm, 43mm respectively.

Effect of different temperature on the growth of Chaetomium globosum

Basu *et al.* (1948) reported that vegetative growth of *Chaetomium globosum* was more rapid at 30 °C than at 22 °C.

Incase of Matthew et al. (2008) [12] studied the growth and mycotoxin production by Chaetomium globosum which was favored in a neutral P^H. In this study, the influence of ambient pH on the growth of C. globosum was examined on an artificial medium. Whereas Asgari et al. (2011) [3] revealed that growth-temperature relationships of the *Chaetomium* sp. growing well at 20 °C-30 °C and maximum at 35 °C-40 °C. The results were conformity with the Neeta Sharma et al. (2011) who revealed that optimum temperature radial growth of C. globosum appeared in between 25 to 35 °C. Excellent growth on 30°C and good growth recorded on growth 25 °C, while almost nil growth on 15 °C and 10 °C. Similar results obtained by Kiran et al. (2015) in temperature study it was observed that Chaetomium can nicely grow between 20 to 35 ⁰C temperature range but it can flourish nicely at 35 ⁰C temperature while temperature below 15 °C and above 45 °C were almost nil growth of Chaetomium. Similar reports of Ahammed (2005) who studied optimum temperature for growth of C. globosum by growing the fungus in medium along with best nitrogen, carbon, vitamin, having optimum pH, incubated at different temperatures viz., 18 °C, 25 °C, 28 ⁰C, 35 ⁰C and 40 ⁰C, this investigations also showed that thermophilism of C. globosum, as it could grow best at temperature of 35 °C. Most of the species grow best between 25 to 35 °C and require a cellulose-rich medium for sporulation. Chaetomium globosum is a saprobic organism and their ability to suppress plant pathogens resulted to induced growth, and high yield of the plant (Sibounnayong et al. 2005).

Chaetomium globosum as a biocontrol agent

Miedtke et al. (1990) reported that Chaetomium globosum as antagonists of the perfect stage of the apple scab pathogen (Venturia inaequalis) under field condition. In autumn, Chaetomium globosum were applied to detached apple leaves naturally infected by V. inaequalis. Treated leaves were overwintered on the orchard floor in boxes covered with coarse plastic mesh. Guang et al. (1991) [8] tested the study of endophytic fungi Chaetomium ND35 antagonism to plant pathogens viz., Macrophoma kuwatsukai, Rhizoctonia solani and Sclerotium rolfsii (Corticium rolfsii) were evaluated in vitro and in vivo. In case of wheat Biswas et al. (2000) [5] studied antagonism of Chaetomium globosum to Drechslera sorokiniana, the spot blotch pathogen. Interaction between Chaetomium globosum and Drechslera sorokiniana produced inhibition zone, indicating biocontrol mechanism of C. globosum through antibiosis. (Soytong et al., 2001) [16] the biocontrol potential of Chaetomium spp. was reported against Fusarium, Helminthosporium, Pythium ultimum, Alternaria raphani, A. brassicicola and Phytophthora pathogens. Singh et al. (2002) studied the management of Pigeonpea wilt by Chaetomium globosum was used as bioagent against Fusarium udum in vitro. Aggarwal et al. (2004) [1] reported the role of antibiosis in the biological control of spot blotch (Cochliobolus sativus) of wheat by Chaetomium globosum. Production of antifungal compounds by Chaetomium globosum (Cg) and their role in suppression of spot blotch of wheat caused by this fungus under in vitro. Istifadah et al. (2006) revealed that isolates of endophytic Chaetomium spp. inhibit the fungal pathogen Pyrenophora tritici-repentis in vitro. Tomilova et al. (2006) [20] studied the effect of a preparation from Chaetomium fungi on the growth of phytopathogenic fungi. They studied the fungicidal activity of a biological preparation from the fungi of the genus Chaetomium against soil phytopathogenic fungi Rhizoctonia solani and Fusarium oxysporum. Emmanuel et al. (2013) evaluated the inhibitory activity of Chaetomium globosum extract against Philippine strain of Pyricularia oryzae. Pv. oryzae were isolated from blast infected leaves of rice in Pangasinan, Philippines. They were cultured and their identities were validated morphologically. Results implied the growth of *P. oryzae* could be inhibited with 500 to 1000 ppm of ethanol extract of *C. globosum in vitro*. Kumar *et al.* (2013) identified antifungal principle in the solvent extract of an endophytic fungus Chaetomium globosum from Withania somnifera. Extracts of Chaetomium globosum EF18, was found effective against *Sclerotinia sclerotiorum*. V. Shanthiyaa $et\ al.\ (2013)^{[22]}$ reported that among eight C. globosum isolate Cg-6 showed greater inhibition to mycelial growth of P. infestans in vitro. C. globosum Cg-6 was formulated as a liquid and applied as a tuber, soil and foliar treatment either individually or in combination against Phytophthora infection in potato plants. Among different treatments, combined application of C. globosum as a tuber treatment @ 1 ml/kg of tubers, as a soil application @ 1 ml/kg of Farm Yard Manure (FYM) and foliar spray @ 0.7% resulted in significantly less late blight infection (72%) compared to untreated control (100%) under field conditions. The application of C. globosum resulted in greater tuber yield by reducing late blight infection in two field trials when compared to untreated controls. The study clearly demonstrated the potential use of C. globosum as a biocontrol agent in the management of late blight disease in potato plants. Poulina Moya et al. (2016) [14] The results confirm the identity of the pathogens and the isolates of Chaetomium globosum species group. Inhibition of B. sorokiniana and D. teres by C2 and C5 accounted for 30% and 31.2%, and 40% and 36% respectively, compared with the control. The mechanisms of action against B. sorokiniana and D. teres were antibiosis and competition and mycoparasitism, respectively. Moreover, it was also reported that C. globosum could produce certain antibiotics and release these to suppress the damping-off of sugar-beet caused by Pythium ultimum (Di Pietro et al. 1991)

Conclusion

In recent years, biological control of soil-borne pathogens has received increasing attention as a promising supplement or alternative to chemical control. To improve efficacy of *Chaetomium globosum* bioagent mechanism of action, nutrition, and ecology of understanding is needed.

Several methods have been established to quantify fungi in soils, including counting the number of fungal spores physically or by plating on selective medium. These techniques have limited success, since they neither ensure adequate sensitivity and accuracy, nor differentiate the desired species isolate from the native organisms. Compared to the above methods, the use of molecular markers provides much promise for the rapid identification and quantification of specific bio control agents in soil and plant. The genome of a microorganism offers several possibilities for monitoring. Genetic interaction of Chaetomium globosum other (fungi and plants) an in depth understanding of mechanisms is lacking. The absence of high throughput studies in these organisms has been due to the lack of whole genome sequences. The genome of Chaetomium spp. has been extensively investigated has proven to contain many useful genes, along with the ability to produce a great variety of expression patterns, which allows these fungi to adapt to many different environments (soil, water, dead tissues, inside the plant, etc.). Finally, understanding the mechanism of interaction *Chaetomium* spp. between and the plant has provided for the first time the opportunity to genetically increase the ability of a Chaetomium spp. strain for the management plant diseases.

References

- 1. Agarwal R, Tiwari AK, Srivastava KD, Singh DV. Role of antibiosis in the biological control of spot blotch (*Cochliobolus sativus*) of wheat by *Chaetomium globosum. Mycopathologia.* 2004; 15:369-377.
- 2. Andrzej Fojutowski, Aleks Andra Kropaczi. Growth of *Chaetomium globosum* fungus in laboratory conditions on agar medium and Scots pine wood Forestry and Wood Technology no. 2015; 92:97-103.
- 3. Asgari B, Zare R. The genus *Chaetomium* in Iran, a phylogenetic study including six new species. Mycologia. 2011; 103(4):863-882.
- 4. Benny GL. A second species of *Chaetomidium* with a cephalothecoid peridium wall, Mycologia. 1980; 72:832-840.
- 5. Biswas SK, Srivastav KD, Aggarwal R, Dureja P, Singh DV. Antagonism of *Chaetomium globosum* to *Drechslera sorokiniana*, the spot blotch pathogen of wheat. Indian Phytopath. 2000; 53(4):436-440.
- 6. Chang I, Kommedahl T. Biological control of seedling of corn by coating kernels with antagonistic microorganisms. Phytopathology. 1968; 77:1470.
- 7. Domsch KH, Gams W, Anderson TH. Compendium of soil fungi. 2nd ed. Munchen: IHW Verlag, 2007, 672.
- 8. Guang LX. Testing on the antagonism of the dominant of Endophytic fungi from *Populas tomentosa: Chaetomium* ND 35 in the laboratory. J Scientia Silvae Sinicae. 1991; 35(5):57-61.
- 9. Harman GE, Eckenrode CJ, Webb DR. Alteration of spermosphere ecosystems affecting oviposition by the bean seed fly and attack by soil borne fungi on germinating seeds. Annals of Applied Biology. 1978; 90:1-6.
- 10. Khan AL, Shinwari ZN, Kim YH, Waqas M, Hamayun M, Kamran M *et al.* Role of endophyte *Chaetomium globosum* Lk4 in growth of *Capsicum annuum* by producion of gibberellins and indole acetic acid. Pak. J Bot. 2012; 44(5):1601-1607.
- 11. Kirk PM, Cannon PF, Minter DW, Stalpers JA. Dictionary of fungi (10th ed.), ISBN 978-085199-826-8. 2008, 131.
- 12. Matthew R, Fogle DR, Straus DC. Growth and mycotoxin production by *Chaetomium globosum* is favoured in a neutral ph. Int. J Mol. Sci. 2008; 9:2357-2365.
- 13. Marwah RG, Fatope MO, Deadman ML, Al-Maqbali YM, Husband J. Musanahol: a new aureonitol-related metabolite from a *Chaetomium* sp. Tetrahedron. 2007; 63:8174-8180.
- 14. Paulina Moya, Debroa Pedemonte, Susana Amengual, Mario Franco EE, Marina Sisterna N. Antagonism and modes of action of *Chaetomium globosum* species group, potential biocontrol agent of barley foliar diseases. Bol. Soc. Argent. Bot. 2016; 51(4):569-578.
- 15. Rajkumar E, Aggarwal R, Singh B. Fungal antagonists for the biological control of Ascochyta blight of chickpea. Acta Phytopathologica et Entomologica Hungarica. 2005; 40:35-42.
- 16. Soytong K, Kanokmedhakul S, Kukongviriyapa V, Isobe M. Application of *Chaetomium* species (Ketomium®) as

- a new broad spectrum biological fungicide for plant disease control: A review article. Fungal Diversity. 2001; 7·1-15
- 17. Tyler JA, Richard C, Richard H, Belanger R. Approaches to molecular characterization of fungal biocontrol agents: some case studies. Canadian Journal of Plant Pathology 2001; 23:8-12.
- 18. Soytong K. *Chaetomium* as a biocontrol agent against plant pathogens. (Abstr.). The XIII International Plant Protection Congress. The Hague. Netherlands, 1995.
- 19. Tveit M, Moore MB. Isolates of *Chaetomium* that protect oats from *Helminthosporium victoriae*. Phytopathology. 1954; 44:686-689.
- 20. Tomilova OG, Shternshis MV. The effect of a preparation from *Chaetomium* fungi on the growth of phytopathogenic fungi. Appl. Biochem. Microbiol. 2006; 42:76e80.
- 21. Raguchander T, Manikandan R, Arunkumar K, Senthil R. *Chaetomium globosum:* A Potential Biocontrol Agent for the Oomycetes Pathogens. J Mycol Plant Pathol, 2014, 44(4).
- 22. Shanthiyaa V, Saravanakumar D, Rajendran L, Karthikeyan G, Prabakar K, Raguchander T. Use of *Chaetomium globosum* for biocontrol of potato late blight disease Crop Protection. 2013; 52:33-38.
- 23. Vannacci G, Harman GE. Biocontrol of seed-borne, *Alternaria raphani* and *Abrassicicola*. Can. Journal of Microbiology. 1987; 33:850-856.
- 24. Von Arx JA, Guarro J, Figueras MJ. The Ascomycete Genus *Chaetomium*. Nova Hedwigia Beihefte, Lubrecht & Cramer Ltd., 1986.