

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2019; 8(2): 854-856 Received: 19-01-2019 Accepted: 21-02-2019

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Isolation and identification of local antagonistic *Trichoderma harzianum* from rhizosphere of Garlic, redgram and potato

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Abstract

A total of 18 rhizosphere soil samples were collected from healthy plants present in garlic, Red gram and potato fields, Piprauli block of Gorakhpur district. Five *Trichoderma* spp. were obtained from soil samples on the basis of morphological identification such as shape of phialides, conidia and growth characters. Further on the basis of farmer's response two isolates were highly effective as compare to others *viz*; TCIPMCG-1 and TCIPMCP-2 and were identified as *Trichoderma harzianum*, morphologically and molecularly from ITCC, New Delhi.

Keywords: Rhizosphere, Trichoderma harzianum, antagonistic, native strain

Introduction

The species of Trichoderma are well known for their biocontrol activity against many plant pathogens that cause major problems in the current agricultural scenario. *Trichoderma* spp. usually available in soil and root ecosystems has gained huge significance since last few decades due to its biological control ability against soil born plant pathogens. Trichoderma in managing soil borne plant pathogens are reported by different workers such as, eco friendly (Gaur et al., 2005) ^[5], effective in managing diseases caused by soil borne plant pathogens which cannot be easily controlled by chemicals (Manoranjitham et al., 2001)^[7], ease and cost effective mass culturing of antagonists (Das et al., 2006)^[3], growth promoting effect (Pan and Bhagat, 2007)^[9] and long lasting effective disease management (Sarojini *et al.*, 2007)^[10]. The potential of Trichoderma spp. as biocontrol agents of plant diseases was first recognised in early 1930's. Trichoderma is characterized as rapidly growing colonies bearing tufted or postulate, repeatedly branched conidiophores with lageniform phialides and hyaline or green conidia born in slimy heads (Shalini and Kotasthene, 2007)^[11]. Several strains of T. viride and Trichoderma harzianum had a significant reducing effect on plant diseases caused by pathogens such as Rhizoctonia solani, Sclerotium rolfsii, Pythium aphanidermatum, Fusarium oxysporum, and other soil borne pathogens under green house and field conditions (Bassim et al., 1999). Soil borne pathogens causing diseases like; collar rot, stem rot, root rot, dampingoff and wilt in crops like brinjal, Papaya, Redgram and Tomato. Root colonization ability of Trichoderma spp. is another important property which make them strong competitive colonizer in the rhizosphere of plant and protect the root infection by pathogens. Thus in present study we concentrated on isolation, identification of *Trichoderma* isolates existing in rhizosphere of Garlic, Redgram and Potato fields in Piprauli block of Gorakhpur district.

Materials and methods

Isolation and Identification of Trichoderma species

Soil samples were collected from rhizosphere of 18 different agricultural fields such as garlic, redgram and potato fields in Gorakhpur district, Uttar Pradesh. Among 18 soil samples, 6 samples from Garlic, 6 samples from Redgram and 6 samples from potato rhizospheric region. These soil samples were collected in clean poly bags and brought to laboratory and kept at 4°C until used. Dilution plate method was used for the isolation of antagonists from soil, for that serial dilutions of each soil samples was done with sterilized distilled water up to 10^{-6} and 0.1 ml diluted sample was poured on the surface of Trichoderma Specific Medium (TSM) (Elad and Chet, 1983). These plates were incubated at $25\pm2^{\circ}$ C and after 4 to 6 days of incubation, the fungal growths on petri plates were screened for *Trichoderma* fungus. Selected fungal isolates were purified through subcultures from single spores. The cultures were identified using the available literature Kiffer and Morelet (2000) ^[6] and monographic contribution provided by Bissett (1991 a, b) ^[1, 2].

Isolates were confirmed by morphological characters on the basis of colony colour and growth; size and shape of conidiophore, phialides and conidia and at species level by ITCC, Division of Plant Pathology IARI, New Delhi. *Trichoderma* spp. was maintained on PDA slants at 4°C for further studies.

Results and Discussion

Among 18 soil samples (6 samples from garlic, 6 samples from redgram and 6 samples from potato rhizosphere region) 5 *Trichoderma* spp. were isolated. From these isolates, 2 were from garlic, 2 from redgram and 1 from potato rhizosphere soil (Table 1 and 2). They were designated as TCIPMCG-1, TCIPMCP-2, TRGRH, TPRB and TRGRJ. In similar study several scientists reported the isolation of *Trichoderma* spp. from rhizosphere soils of different crops. Gajera *et al.* (2011) ^[4] isolated several *Trichoderma* from groundnut crop.

Sundaramoorthy and Balabaskar (2013)^[13] isolated fungal native isolates of Trichoderma from rhizosphere of Tomato. According to growth pattern 5 Trichoderma spp. were selected for identification. Out of them two species, TCIPMCG-1 and TCIPMCP-2 were very fast growing. According to farmer's response on effectiveness of these two species were identified as Trichoderma harzianum on the basis morphological and of molecular characterization from ITCC (Indian Type of Culture Collection), New Delhi. Similarly, Singh., (2006)^[12] obtained twenty seven isolates of T. harzianum from soil samples collected randomly from fallow agricultural fields throughout the Punjab and studied their growth rate on PDA medium and were categorised as medium, fast and very fast growing and Muthukumar and Pratibha Sharma (2011)^[8] used morphological description for characterization and grouping of Trichoderma isolates.

 Table 1: Soil Samples collected from rhizosphere region of garlic, redgram and potato fields from different villages of Piprauli block in Gorakhpur district

S. No	Name of the District	Name of the Block	Name of the village	Crops	No of soil samples
1.	Gorakhpur	Piprauli		Garlic (Farm sample)	2
2.	-do-	-do-	Chhapia	Red gram	1
3.	do	do		Potato	1
4.	do	do		Garlic	1
5.	do	do	Jaitpur	Red gram	2
6.	do	do	_	Potato	1
7.	do	do		Garlic	1
8.	do	do	Bargahan	Red gram	1
9.	do	do		Potato	1
10.	do	do		Garlic	1
11.	do	do	Hardia	Red gram	1
12.	do	do		Potato	2
13.	do	do		Garlic	1
14.	do	do	Haraya	Red gram	1
15	do	do		Potato	1
Total					18

Table 2: List of Trichoderma spp. isolated from soil samples collected from different villages in Gorakhpur Dist.

S. No.	Isolate No	Name of village
1.	TCIPMCG-1	Chhapia (Farm sample)
2.	TCIPMCP-2	Hardia
3.	TRGRH	Haraya
4.	TPRB	Bargahan
5.	TRGRJ	Jaitpur

TCIPMCG = *Trichoderma* Central IPM Centre Garlic, TCIPMCP = *Trichoderma* Central IPM Centre Potato TRGRH = *Trichoderma* Red gram Rhizosphere Haraya, TPRB= *Trichoderma* Potato Rhizosphere Bargahan, TRGRJ = *Trichoderma* Red gram Rhizosphere Jaitpur.

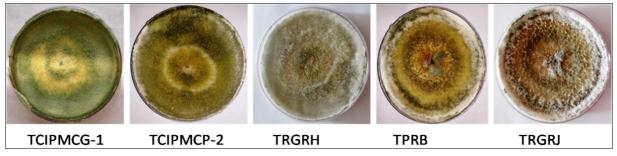


Fig 1: Pure cultures of Trichoderma isolates from rhizosphere region of Garlic, Potato and redgram.

Conclusion

Result of above finding's reveals that the TCIPMCG -1 and TCIPMCP-2 isolated from the rhizospheric soil of healthy garlic and potato plants, have strong biocontrol activity

against several plant diseases in field conditions. Therefore, the application of indigenous *Trichoderma harzianum* in the field tended to reduce the soil and seed borne diseases of several crops.

Acknowledgement

The authors express their gratitude to the Plant Protection Adviser and Joint Director (IPM), Directorate of Plant Protection Quarantine & Storage, N.H.IV, Faridabad-121001 (Haryana) for their encouragement and facilities provided during the work.

References

- 1. Bisset J. A revision of the genus *Trichoderma* II. Infragenric classification Canadian Journal of Botany. 1991a; 69:2373-2417.
- Bisset J. A revision of the genus *Trichoderma* III. Sect. *Pachybasium*. Canadian Journal of Botany. 1991b; 69:2373-2417.
- 3. Das BC, Das BK, Pranab D, Sarmah DK. Bioformulation of *Trichoderma harzianum* Rifai for management of soyabean stem rot caused by *Rhizoctonia solani* Khunn. Journal of Biological Control. 2006; 20(1):57-64.
- 4. Gajera H, Kalu R. Dinesh V. Bioefficacy of *Trichoderma* isolates against *Aspergillus niger* Van Tieghem inciting collar rot in groundnut (*Arachis hypogaea* L.). Journal of Plant Protection Research. 2011; 51:240-247.
- 5. Gaur RB, Sharma RN, Sharma RR, Singh VG. Efficacy of *Trichoderma* for *Rhizoctonia* root rot control in chickpea. Journal of Mycology and Plant Pathology. 2005; 35(1):144-150.
- 6. Kiffer E, Morelet E. The Deuteromycetes Mitosporic Fungi Classification and Generic Keys. Science Publications. Inc. USA, 2000, 1152.
- 7. Manoranjitham SK, Prakasam V, Rajappan K. Biocontrol of damping-off of tomato caused by *Pythium aphanidermatum*. Indian Phytopathology. 2001; 54:59-61.
- Muthukumar A, Sharma P. Molecular and morphological characters: An appurtenance for antagonism in *Trichoderma* spp. African Journal of Biotechnology. 2011; 10(22):4532-4543.
- 9. Pan S, Bhagat S. Antagonistic potential of *Trichoderma* and *Gliocladium* spp. from West Bengal.Journal of Mycology and Plant Pathology. 2007; 37(2):235-239.
- Sarojini K, Chakravarthy, Nagamani A. Efficacy of nonvolatile and volatile compounds of *Trichoderma* species on *Rhizoctonia solani*. Journal of Mycology and Plant Pathology. 2007; 37(1):82-86.
- 11. Shalini S, Kotasthane AS. Parasitism of *Rhizoctonia solani* by strains of *Trichoderma* spp. Electronic Journal of Environmental, Agriculture and Food Chemistry. 2007; 6(11):1220-1225.
- 12. Singh HB. Management of plant pathogens with microorganisms, Journal of Mycology and Plant Pathology. 2006; 36:373-384.
- 13. Sundaramoorthy S, Balabaskar P. Biocontrol efficacy of *Trichoderma* spp. against wilt of tomato caused by *Fusarium oxysporum* f. sp. *Lycopersici*. Journal of Applied Biology and Biotechnology. 2013; 1(3):36-40.