

E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2018; 7(6): 2116-2118 Received: 28-09-2018 Accepted: 30-10-2018

Ishrat Mohamad

Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Farooz A Bhat

Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

MH Balkhi

Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Tasaduq H Shah

Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Bilal A Bhat

Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Asifa Wali

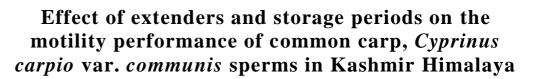
Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Correspondence

Ishrat Mohamad Faculty of Fisheries, Rangil, Ganderbal, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Jammu and Kashmir, India

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



Journal of Pharmacognosy and

Phytochemistry

Ishrat Mohamad, Farooz A Bhat, MH Balkhi, Tasaduq H Shah, Bilal A Bhat and Asifa Wali

Abstract

The effect of short term storage on the motility rate and motility duration was assessed at different storage periods. The sperms were diluted with3 different extenders viz. Extender I contained 300 mM glucose solution. Extender II contained 1% NaCl solution. Extender III contained 75 mM NaCl, 70 mM KCl, 2 mM CaCl₂, 1 mM MgSO4, 20 mM Tris (Modified ionic solution) at ratio of 1:3. Glucose containing extender showed the best results. The highest motility rate and duration was found with the glucose based extender at 0 hour storage period. Therefore it is found that glucose based extender is the best extenders used in the study and motility performance of sperm of *Cyprinus carpio* var. *communis* decreases with the storage time.

Keywords: Motility rate, extender, Cyprinus carpio var. communis, sperms

1. Introduction

Common carp is the most commercially important species. Major production of the species comes from the aquaculture operation. However, the quality of seed gets deteriorated in hatchery due to inbreeding depression, negative selection etc. So, the preservation of genetic diversity is a great challenge. Short term preservation of sperms can be considered as an important tool for reducing the genetic erosion that invariably results from inbreeding and that can allow the preservation of the various male donors from all the possible strains ^[1].

For the successful hatchery operations, short-term preservation of fish sperm is very important. The short-term storage of sperm at low temperature (4°C) is mostly used to obtain the good quality gamete of both male and female sex at the same time during artificial insemination, also in research programs for genetic studies. Maturation of gametes and the time of spawning between the sexes at different times has been documented in number of species. To overcome these problems, researchers have examined procedures such as cryopreservation, short-term, refrigerated storage of milt. Short-term, refrigerated storage of milt have been developed for several teleosts, such as walleye ^[2] Red drum Sciaenops ocellatus ^[3], Atlantic sturgeon Acipenser oxyrinchus^[4] Mozambique tilapia Tilapia mossambica^[5], and Paddlefish Polyodon spathula^[6]. The fish sperm could be preserved by storage in undiluted and diluted form. Sperm stored at low temperature in undiluted form has been reported to cause a reduction in fertilization capacity ^[7]. Undiluted storage provides less control compared to the diluted storage of sperm with extender ^[5]. The dilution of the sperm in a suitable medium helps in the preservation for short time periods, while transporting from the place of collection to the place of employment, as it helps to keep the physicochemical properties of semen stable ^[8,9]. These solutions, however, helps in keeping the sperms inactive, and thus prevent the exhausting energy reserves of the spermatozoa^[10].

2. Materials and methods

2.1 Milt collection

Mature male brooders were collected from the hatchery of Faculty of fisheries SKUAST K during the breeding season. Each male was stripped once only and the total amount of expressible milt was collected individually by gently pressing the abdomen. The semen was collected directly into clean 15 ml graduated centrifuge tubes. Care was taken to avoid the contamination of semen with water, urine, blood or faecal matter. The tubes were covered and immediately transported on ice (4°C) to the laboratory for analyses.

2.2 Estimation of Sperm motility

Motility was evaluated using a light microscope (Olympus CX31) at 40x magnification and was expressed as percentage of motile spermatozoa. An activating solution of 0.3% NaCl was used to estimate motility. For the evaluation of motility, about 10 μ l of semen was placed on a glass microscope slide and 100 μ l of activation solution was added ^[11].

2.3Short-term storage of sperm

Three extenders were selected for evaluating potential extender for fishes as shown in table 1. The semen was

diluted at ratio of 1:3 with one of three extenders. Extender I contained 300 mM glucose solution as described by Tekin *et al.* (2003). Extender II contained 1% NaCl solution. Extender III contained 75 mM NaCl, 70 mM KCl, 2 mM CaCl₂, 1 mM MgSO4, 20 mM Tris (Modified ionic solution) as described by Lahnsteiner *et al.* (1998). The diluted sperm was packaged in small laboratory tubes. Undiluted semen samples were used as control. Experimental and control samples were stored at 4°C for 4 hours in a refrigerator. During refrigerated preservation, motility (%), motility durations (s) of stored sperm were evaluated at 1 hour intervals.

Table-1: Composition of semen extenders

Extenders	Composition
Extender I	300 mM of glucose solution ^[12]
Extender II	1% NaCl solution
Extender III	75 mM NaCl, 70 mM KCl, 2 mM CaCl ₂ , 1 mM MgSO4, 20 mM Tris (Modified ionic solution. ^[13]

3. Results

3.1 Short term preservation of Common carp sperm 3.1.1 Post-activation motility rate

Data on the effect of extenders and storage periods on motility rate are given in Table 2. Spermatozoa motility percentage decreased significantly with increasing storage period in diluted and undiluted sperm during spawning season. When the effect of extenders were tested, the best results (74% motility) were achieved with extender I containing glucose solution at the e 0 hours storage periods. On the contrary, the lowest motility (4.66%) was obtained with control group at 4 hour storage periods. Table 2, shows that statistically there was a significant difference in motility rate in extenders, extender 1, extender 2, extender 3 and no extender at storage time 0, 1, 2, 3 and 4 hr. The maximum value of extender 1 was at 0 hr and minimum value was at 4 hr storage period indicating significant decrease in motility percentage from 0 hr storage to 4 hr storage. The decrease in motility (%) was also found in extender 2, extender 3 and no extender from 0 hr storage to 4 hr storage. At 0 hr storage the maximum value of motility rate (74%) was of extender 1 and minimum value (53.66%) was of extender 3. At 1 hr storage, the maximum value (56.33%) was of extender 1 and minimum value (42.66%) was of extender 2. The maximum value (40%) at 2 hr storage was of extender 1 and minimum value (31%) was that of control (undiluted milt). The maximum value (30%) at 3 hr storage was of extender 1 and minimum value (15%) was of extender control (undiluted milt). The maximum value (14.33%) at 4 hr was of extender 1 and minimum value (4.66%) was of control (undiluted milt). The Extender 1 showed better performance of motility rate at all storage periods with a mean value of 42.9% followed by Extender 2 with a mean value of 35.66%.

Storage period (hour)	Extender (%)			Control (coo)	Maan (aaa)
Storage period (hour)	1	2	3	Control (sec)	Mean (sec)
0	74.00	66.33	53.66	69.66	65.91
1	56.33	42.66	44.666	50.33	48.50
2	40.00	35.00	38.66	31.00	36.16
3	30.00	25.33	29.66	15.00	25.00
4	14.33	9.00	10.00	4.66	9.50
Mean	42.90	35.66	35.33	34.133	

Table 2:Effect of extenders and storage periods on motility rates of Scale carp sperm during spawning season

CD at 5% level of significance; Storage = 2.7435; Extender = 2.4538; Storage × Extender = 5.487

3.1.2 Post-activation motility durations

Data on the effect of extenders and storage periods on motility durations are given in Table 3. In case of testing extenders, the highest motility duration (54 sec) was achieved with extender 1 at 0 hours storage periods during spawning season. On the contrary, the lowest motility duration of 5 sec was observed with extender 3 at 4 hour storage period. Decrease in motility duration is significantly important with increasing storage period in diluted and undiluted sperm during spawning season, differences among storage hours and extenders were statistically significant during experiment. The maximum value (54 sec) of extender 1 was at 0 hr and minimum value at 4 hr storage (16.33 sec) indicated significant decrease in motility duration with storage. The decrease in motility duration was also observed in extender 2, extender 3 and control (undiluted milt) from 0 to 4 hr storage. The maximum value (56 sec) of motility duration at 0 hr was that of extender 2 and minimum value (42 s) was that of extender 1 (48 sec) and minimum value (33 sec) was that of extender 3. The maximum value (31 sec) at 2 hr was that of extender 1 and minimum value (24 sec) was that of extender 3. The maximum value (21 sec) at 3 hr was that of extender 1 and minimum value (11.66 sec) was that of control (undiluted milt). The maximum value (16.33 s) at 4 hr was that of extender 3. The extender 1 shows better performance of motility duration at all storage periods with a mean value of 34.067 sec of followed by extender 2 with a mean value 29.22 sec, control (undiluted milt) with a mean value of 26.133 sec and extender 3 with a mean value 24.33 sec.

Table 3: Effect of extenders and storage periods on motility duration of scale carp carp sperm during spawning season

Storage period	Ext	ender ((sec)	Control (ma)	Maan (aaa)		
(hour)	1	2	3	Control (sec)	Mean (sec)		
0	54.00	56.00	42.00	46.00	49.50		
1	48.00	42.00	33.00	39.00	40.50		
2	31.00	27.00	24.00	25.00	26.75		
3	21.00	13.00	19.66	11.66	16.33		
4	16.33	8.00	5.00	9.00	9.588		
Mean	34.067	29.22	24.733	26.133			
SD at 50/ land af air if ann an Stamper 2 2828; Entendan 2 0257							

CD at 5% level of significance; Storage = 3.3828; Extender = 3.0257; Storage X Extender = 6.766

4. Discussion

The most important indicator for the success of a preservation protocol is the post-activation motility. The present work reports that the spermatozoa motility get affected during preservation. Glucose containing extender was found with the best motility results. The motility percentage decreased faster with time in undiluted sperm samples than diluted ones. The motility of diluted sperm was always significantly greater than that of undiluted sperm, Similar results for the motility parameters of spermatozoa were reported by different worker ^[14,15, 16] also reported that no significant changes in the quality were found when the spermatozoa are kept in appropriate extender at low temperature (4°C), ^[5] also observed that diluted sperm with extender provides better control compared to undiluted storage. ^[17]Also reported that the best results in fertilization and motility are obtained from the sperm samples of rainbow trout diluted with artificial seminal plasma with the rate of 1:1 stored for 7 days. It is possible that by using suitable activating mediums, duration of motility will be increased that will enhance fertilizing capacity of the fish. Sperm dilution is a major factor in the induction of sperm motility [18, 19,20] reported that the duration of sperm motility

and intensity of spermatozoa in *A. baeri* increased when the dilution rate increased from 1:6 to 1:100. In the present study, the maximum duration of motility has been determined with extender I containing 300 mM glucose. Therefore the extender I provided longer duration of motility since glucose served as an energy resource for spermatozoa. Similar findings were reported by ^[21] Hatipoglu and Akçay (2010) who found that glucose based extender is a better preservative than Ringer solution for the short term preservation of Abant trout (*Salmo trutta abanticus*) semen.

5. Conclusion

It is concluded from the present study that glucose based extender is best at the dilution ratio of 1:3. But the development of suitable protocol for short term preservation of milt more research in future.

6. References

- 1. FAO. The state of food and agriculture Food & Agriculture Organization of the UN (FAO), 1989, 37.
- 2. Moore AA. Short-term storage and cryopreservation of walleye semen Progressive Fish-Culturist. 1987; 49:40-43.
- Wayman WR, Tiersch TR, Thomas RG. Refrigerated storage and cryopreservation of sperm of red drum, *Sciaenops ocellatus* L. Aquaculture Research 1998; 29:267-273.
- Dilauro MN, Krise WF, Hendrix MA, Baker SE. Shortterm cold storage of Atlantic sturgeon sperm. Progressive Fish-Culturist. 1994; 56:143-144
- 5. Harvey B, Kelley RN. Chilled storage of *Sarotherodon mossambicus* semen. Aquaculture. 1984; 36:85-95.

- Brown GG, Mims SD. Storage, transportation and fertility of undiluted and diluted paddlefish milt. Progressive Fish-Culturist. 1995; 57:64-69.
- 7. Lahnsteiner F, Berger R, Weismand T, Patzner R. Sperm motility and seminal fluid composition in the burbot, *Lota lota*. Journal of Applied Ichthyology. 1997; 13:113-119.
- 8. Gwo JC. Cryopreservation of yellowfin seabream (*Acanthopagrus latus*) spermatozoa (teleost, perciformes, sparides), Theriogenology. 1994; 41:989-1004.
- Ohta H, Ilzawa T. Diluent for cold storage of the Japanese eel (Anguilla japonica) spermatozoa. Aquaculture. 1996; 142:107-118.
- Gwo JC. Cryopreservation of black grouper (*Epinephelus malabaricus*) spermatozoa. Theriogenology. 1993; 39:1331-1342.
- 11. Mims SD. Evaluation of activator solutions, motility duration and short term storage of paddle fish spermatozoa. Journal of World Aquaculture Society. 1991; 22:224-229.
- 12. Tekin N, Secer S, Akcay E, Bozkurt Y, Kayam S. The effect of age on spermatological properties in rainbow trout (*Oncorhynchus mykiss* W. 1792). Turkish Journal of Veterinary and Animal Sciences. 2003; 27:37-44.
- 13. Lahnsteiner F, Berger B, Weismann T, Patzner RA. Determination of semen quality of the rainbow trout, *Oncorhynchus mykiss*, by sperm motility, seminal plasma parameters and spermatozoa metabolism. Aquaculture. 1998; 16:163-181.
- Stoss J, Holtz W. Cryopreservation of rainbow trout (*Salmo gairdneri*) sperm. III. Effect of proteins in the diluent, sperm from different males and interval between sperm collection and freezing. Aquaculture. 1983; 31:275-282
- 15. Bozkurt Y, Secer S. Effect of short-term preservation of mirror carp (*Cyprinus carpio*) semen on motility, fertilization and hatching rates. Israeli Journal of Aquaculture-Bamidgeh. 2005; 57(3):207-212.
- 16. Kime DE, Ebrahimi M, Nysten K, Roelants I, Rurangwa E, Moore HDM, Ollevier F. Use of computer assisted sperm analysis (CASA) for monitoring the effects of pollution on sperm quality of fish; application to effects of heavy metals. Aquatic Toxicology. 1996; 36:223-237.
- Canyurt MA, Akhan S, Takma C. A study on short-therm storage of rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792) milt. EU. Journal of Fisheries and Aquatic Sciences. 2003; 20(3-4):537-542.
- 18. Billard R, Cosson, MP. Some problems related to the assessment of sperm motility in freshwater fish. Journal of Experimental Zoology. 1992; 261:122-131.
- 19. Ginsburg AS. Fertilization in fishes and the problem of polyspermy. Izdatelnaya Nauka, Mosco, 1968.
- Gallis JL, Fedrigo E, Jatteau P, Bonpunt E, Billard R. Siberian sturgeon spermatozoa. Effects of dilution, pH, osmotic pressure, sodium and potassium ions on motility. In: Acipenser. Cemagref Publ., Antony, France (Ed. P. Williot), 1991, 143-151.
- Hatipoglu T, Akçay E. Fertilizing ability of short-term preserved spermatozoa Abant trout (*Salmo trutta abanticus* T, 1954). Ankara Univ Vet Fak Derg. 2010; 57:33-38.