

# Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2018; 7(5): 2517-2521 Received: 07-07-2018 Accepted: 08-08-2018

## Sukhpreet Kaur Sidhu

Department of Botany, Punjab Agricultural University, Ludhiana, Punjab, India

#### Jagmeet Kaur

Department of Plant Breeding and Genetics, Punjab Agricultural University, Ludhiana, Punjab, India Cluster, correlation and path coefficient analysis of phosphorus use efficiency related traits in pigeonpea

# Sukhpreet Kaur Sidhu and Jagmeet Kaur

#### Abstract

The present investigation was undertaken to study the genetic diversity, correlation and path coefficient analysis for 16 metric traits in 43 pigeonpea genotypes at Department of Plant Breeding and Genetics in the Punjab Agricultural University, Ludhiana during *Kharif* 2013. Pigeonpea genotypes were grown in pots and two phosphorus doses were supplied to soil in the pots; one was recommended dose of phosphorus supplied [Single Super Phosphate@ 250 kg/ha<sup>-1</sup> nutrient (+P)] and in second, P fertilizer was not added in soil (-P). Within clusters sum of squares were maximum in cluster I. This cluster showed highest average values of root area and perimeter was highest of this cluster than other four clusters under recommended dose of P. Root perimeter was highly positively correlated with root area and a negative correlation was found among leaf P and root length, root P and root length, root P and root dry weight, stem P and root area, leaf P and root area. Path analysis of leaf P indicated that root dry weight had maximum significant positive direct effect followed by root volume, leaf area on leaf P. Genetic diversity, correlation coefficient and path analysis approaches may be used in selecting traits which contribute for phosphorus use efficiency from large pool of germplasm for breeding purpose.

**Keywords:** Cluster analysis, pearson correlation coefficient, path analysis, pigeonpea genotypes

## Introduction

Pigeon pea (Cajanus cajan (L.) Millsp.) is the second most important pulse crop and it has advantages over other legumes, such as tolerant toward drought condition because of its long deep rooting system (Cook et al., 2005)<sup>[3]</sup>. Pigeon pea seed contains 57 – 59% of carbohydrate, 14 – 30% of protein, 1 -9% of fat, 100g of mature raw pigeon peas also provide 76% for pregnant women of the daily requirement of folate and high vitamin A, B complex, and C (Mahalaya et al., 2015)<sup>[8]</sup>. In India, pigeonpea is cultivated in 3.9 million hectare with production of 3.2 million tonnes (INDIASTAT, 2015)<sup>[5]</sup>. Efforts to enhance the phosphorus (P) efficiency of cropping systems have typically focused on optimizing P-fertilizer application by regulating the rate of fertilizer needed to achieve highest production. Moreover, small amount of P fertilizer confines yields, whereas too large amount is an unnecessary expenditure and can cause damage to environment through leaching (Shapley et al., 1994)<sup>[11]</sup>. There are a number of possible adaptive mechanisms that the P-efficient plants can utilize for superior root growth on low P soils, including modification in root morphology, root symbiosis, improvement of internal phosphatase activity, secretion of organic acids into the rhizosphere, activation of high-affinity phosphate transporters (Wang et al., 2010)<sup>[14]</sup>. Both internal phosphorus use efficiency (PUE) and external PUE are independent processes, some approaches of improvement in equalizing P uptake be supposed to employed (Veneklaas et al., 2012)<sup>[12]</sup>. In soil supplied with fertilizers, P availability higher in top layers of soil. Plants with better root traits (more root surface area, hairs, branching and volume) are more capable to acquire P from top soil (Manschadi et al., 2013)<sup>[9]</sup>. Root hair density and increase in lateral branching are the most useful root traits for phosphorus use efficiency (Clemens et al., 2016) <sup>[2]</sup>. Plants can also turn on a set of adaptive responses to boost P uptake and P recycling by reprogramming of physiology processes and alter root structure (Jain et al., 2007) <sup>[6]</sup> to maintain their development rate as feasible (Gutschick and Kay 1995)<sup>[4]</sup>. Root characters *i.e* root length, fineness, surface area, and root hair density affect the behavior of plant in P deficient soils (Rao et al., 1996)<sup>[10]</sup>. Keeping in view the investigation of forty three pigeonpea genotypes were undertaken to estimate the correlation and path coefficient for PUE among sixteen characters associated to root and shoot.

Correspondence Sukhpreet Kaur Sidhu Department of Botany, Punjab Agricultural University, Ludhiana, Punjab, India

## Materials and Methods

Forty three genotypes (AL1584, AL1778, AL1812, AL1816, AL1817, AL1836, AL1838, AL1839, AL1842, AL1843, AL1847, AL1849, AL1853, AL1873, AL1881, AL1931, AL1932, AL1933, AL1941, AL15, AL1593, AL1747, AL1756, AL1758, AL201, IC245504, IC245506, IC245507, ICPL 20329, ICPL 20330, ICPL 20340, ICPL 88039, Manak, MN1, MN5, P2001, P2002, P992, Pant A 402, Paras, PAU881, UPAS120 and VLA11) were assessed in pots with eight replications in factorial complete randomized design. Seeds of genotypes were obtained from International Crops Research Institute for the Semi-Arid Tropics, Hyderabad, Chaudhary Charan Singh Haryana Agricultural University, Hisar, Punjab Agricultural University, Ludhiana. Two phosphorus doses were supplied to soil in the pots; one was recommended dose of phosphorus supplied [Single Super Phosphate@ 250 kg/ha-1 nutrient (+P)] and in second, P fertilizer was not added in soil (-P). The present investigation was carried out at the experimental area and laboratories of the Pulses section, Department of Plant Breeding and Genetics in the Punjab Agricultural University, Ludhiana during Kharif 2013. Observations were recorded for sixteen characters viz., root length, root volume (cm<sup>3</sup>), root dry weight (g), root area (mm<sup>2</sup>,) root perimeter (mm<sup>2</sup>), number of root tips, shoot length (cm), shoot dry weight (g), number of leaves, leaves dry weight (g), leaf area (cm<sup>2</sup>), photosynthesis rate, stem p, root p and leaf p at 60 days after sowing. Genotypes were classified on the basis of their root traits was done using the ward cluster method. Pigeonpea genotypes were divided into different clusters under both P treatments by using Manitab17 software.

# **Result and Discussion**

## Growth and root characteristics based clustering

Forty three pigeonpea genotypes based on root acid phosphatase activity, photosynthetic rate, root and shoot morphological traits were categorized into five groups using ward cluster squared Euclidean distance method.

# First cluster genotypes

Genotypes AL1584, AL1843, AL1873, AL1593, AL1758, IC245507, MN1, P992, Paras and UPAS120 were classified in the first cluster out of 43 genotypes (Table 1). The minimum inter-cluster distance (D = 2.334) was found between cluster I and II under no added P condition.

AL 1584, AL 1778, AL 1812, AL 1816, AL 1817. AL 1836, AL 1839, AL 1842, AL 1843, AL 1853, AL 1873, AL 1881, AL 1932, AL 1933, AL 1941, AL 1593, AL 1756, AL 1758, IC 245504, Pant A 402 and VLA11 (Table 2). Least intercluster distance (D = 4.202) between cluster I and II was observed (Table 3). Within clusters sum of squares were maximum in cluster I. This cluster showed highest average values of root area and perimeter was highest of this cluster than other four clusters under recommended dose of P.

# Second cluster genotypes

AL 1778, AL 1747, AL 1756, IC 245504, ICPL 20329, P 2001, P2002, Pant A 402 and VLA11 genotypes were categorized in the second cluster. Highest and two successively lower inter-cluster distance values were 5.584, 5.257 and 3.973 for cluster V and II, IV and II, III and II, respectively under no added P condition.

Genotypes AL1838, AL 1847, AL 1849, AL 1931, AL15, AL 201, ICPL 20329, Manak, MN1, P 2001, P992, Paras, PAU 881 and UPAS120 belonged to this cluster under recommended dose of P. Inter-cluster distance 7.565, 7.413 and 5.481 was found between cluster V and II, III and II, IV and II respectively.

**Table 1:** Distribution of pigeonpea genotypes to different clusters on the basis of ward cluster method (P not added)

Clusters	Number of genotypes	Genotypes									
Ι	10	AL1584, AL 1843, AL 1873, AL 1593, AL 1758, IC245507, MN1 P992, Paras, UPAS120									
II	9	AL 1778, AL 1747, AL 1756, IC 245504, ICPL 20329, P 2001, P2002, Pant A 402, VLA11									
III	10	AL 1816, AL 1839, AL 1842, AL 1881, AL 1932, AL 1933 AL 1941, AL 15, IC245506 , PAU 881									
IV	10	AL 1812, AL 1817, AL 1836, AL 1838, AL 1847, AL 1849, AL 1853, AL 1931, AL 201, Manak,									
V	4	ICPL 20330, ICPL 20340, ICPL 88039, MN5									

Cluste rs	Number of genotypes	Genotypes										
Ι	21	AL 1584, AL 1778, AL 1812, AL 1816, AL 1817. AL 1836, AL 1839 AL 1842, AL 1843, AL 1853, AL 1873, AL 1881, AL 1932, AL 1933 AL 1941, AL 1593, AL 1756, AL 1758, IC 245504, Pant A 402, VLA11										
II	14	AL 1838, AL 1847, AL 1849, AL 1931, AL 15, AL 201, ICPL 20329 Manak, MN1, P 2001, P992, Paras, PAU 881, UPAS120										
III	3	AL 1747, IC245506, IC245507										
IV	4	ICPL 20330, ICPL 20340, ICPL 88039, MN5										
V	1	P2002										

# Third cluster genotypes

Ten genotypes namely AL 1812, AL 1816, AL 1839, AL 1842, AL 1881, AL 1932, AL 1933, AL 1941, AL 15 and IC245506 were included in this cluster. Highest average mean value of stem P content was observed under no added P condition.

Genotypes AL 1747, IC245506 and IC245507 were classified in third cluster under recommended dose of P. Mean values of root length, root volume, root dry weight, number of root tips, shoot length, shoot dry weight, number of leaves, leaf dry weight, leaf area were more of this cluster.

#### Fourth cluster genotypes

Cluster IV was formed by AL 1817, AL 1836, AL 1838, AL 1847, AL 1849, AL 1853, AL 1931, AL 201, Manak and PAU 881. Inter-cluster distance values revealed that the maximum inter cluster distance (D= 10.298) was found between cluster V and cluster IV under no added P condition.

ICPL 20330, ICPL 20340, ICPL 88039, MN5 were belonged to this cluster under recommended dose of P. This cluster showed highest average values of photosynthetic rate, root acid phosphatase activity, stem P content, root P content and leaf P content. The maximum inter-distance (D=7.767) found between cluster V and IV (Table 3).

Table 3: Distance values between cluster centroids of pigeonpea genotypes under P not added and recommended P dose conditions

Closeter			-P		+P							
Cluster	Ι	II	III	IV	V	Ι	II	III	IV	V		
Ι	0					0						
II	2.334	0				4.202	0					
III	3.047	3.973	0			4.593	7.413	0				
IV	3.534	5.257	2.414	0		4.489	5.481	5.223	0			
V	7.218	5.584	8.930	10.298	0	6.846	7.565	7.655	7.767	0		

#### Fifth cluster genotypes

Genotypes ICPL 20330, ICPL 20340, ICPL 88039 and MN5 were classified in this cluster. Mean values of root length, root volume, root dry weight, root area, root perimeter, number of root tips, shoot length, shoot dry weight, number of leaves, leaf dry weight, leaf area, photosynthetic rate, root acid phosphatase activity root P content and leaf P content were more in cluster V under no added P condition. The P2002 was only one genotype belonged to this cluster under recommended dose of P condition.

## Pearson correlation coefficient and Path analysis

Correlation coefficient analysis analyze quantitative relationship among root-shoot traits and leaf P. Root perimeter was highly positively correlated with root area (r = 0.934) followed by leaf area and leaf dry weight (r = 0.872), shoot dry weight and shoot length (r = 0.733), root dry weight and root volume (r = 0.695), leaf dry weight and root dry weight (r = 0.684) while leaf P had no correlation with root volume, shoot dry weight, stem P, photosynthetic rate, number of root tips, acid phosphatase activity and root dry weight. A negative correlation was found among leaf P and root length (r = -0.091), root P and root length (r = -0.087), root P and root area, leaf P and root area. Number of root tips were negatively correlated with photosynthetic rate, acid phosphatase activity, stem P and root r puder +P condition

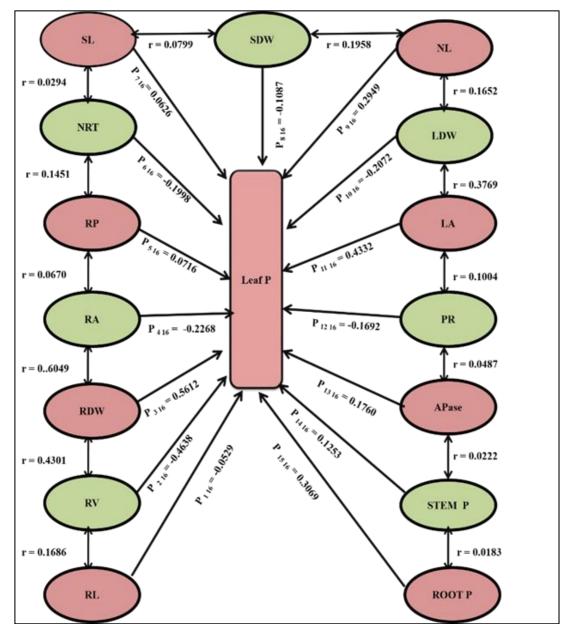
(Table 4). Root area of *Arabidopsis* increased with increase in fine root length in response to P deficient condition (Bates and Lynch, 2001)<sup>[1]</sup>.

Value of coefficient varied from -0.034 (number of root tips and stem P) to (root perimeter and root area) under - P condition (Table 4). Leaf P was positively correlated with leaf dry weight (r = 0.626) followed by root P (r = 0.622), photosynthetic rate (r = 0.618), root dry weight (r = 0.577). There was negative correlation found between stem P and root area (r = -0.038), root perimeter (r = -0.037), number of root tips (r = -0.034), leaf area (r = -0.097), acid phosphatase activity (r = -0.154). Root area with root perimeter (r = 0.980) followed by leaf dry weight and leaf area (r = 0.869), root volume and root dry weight (r = 0.859) were highly positively correlated. It was observed that stem P and root P, root length, root volume, root dry weight, shoot dry weight, number of leaves, root length and acid phosphatase activity had no correlation. A positive significant correlation was observed between total P uptake and root dry weight (Vesterager et al., 2006) <sup>[13]</sup>. Genotypes with maximum P acquisition efficiency represented more root area, volume, dry weight (Zhang et al., 2009) <sup>[15]</sup> and released organic acids to mobilize P under P deficiency (Wang et al., 2010). Shoot P content, root dry weight was positively correlated with total dry weight of pigeonpea genotypes in response to low P soil (Krishnappa et al., 2014)<sup>[7]</sup>.

 Table 4: Pearson correlation coefficient among different variables and leaf P at 60 DAS of pigeonpea under -P (above the diagonal) and +P (below the diagonal)

	RL	RV	RDW	RA	RP	NRT	SL	SDW	NL	LDW	LA	PR	APase	Stem P	Root P	Leaf P
RL	1	0.614**	0.522**	0.515**	0.438**	0.463**	0.585**	0.219**	0.487**	0.428**	0.361*	0.510**	0.284	0.103	0.537**	0.501**
RV	0.635**	1	0.859**	0.754**	0.736**	0.715**	0.621**	0.571**	0.799**	0.739**	0.662**	0.681**	0.563**	0.083	0.655**	0.526**
RDW	0.570**	0.695**	1	0.797**	0.803**	0.714**	0.564**	0.636**	0.697**	0.797**	0.715**	0.698**	0.605**	0.092	0.563**	0.577**
RA	0.609**	0.570**	0.596**	1	0.980**	0.751**	0.605**	0.540**	0.582**	0.723**	0.716**	0.647**	0.425**	-0.038	0.512**	0.408**
RP	0.667**	0.538**	0.547**	0.934**	1	0.751**	0.588**	0.571**	0.595**	0.703**	0.693**	0.630**	0.426**	-0.037	0.478**	0.364*
NRT	0.366*	0.351*	0.472**	0.434**	0.497**	1	0.745**	0.621**	0.765**	0.716**	0.665**	0.567**	0.559**	-0.034	0.485**	0.359*
SL	0.658**	0.537**	0.503**	0.644**	0.626**	0.358*	1	0.685**	0.743**	0.661**	0.576**	0.467**	0.421**	0.062	0.430**	0.412**
SDW	0.552**	0.614**	0.711**	0.632**	0.646**	0.543**	0.773**	1	0.743**	0.750**	$0.588^{**}$	0.477**	0.475**	0.085	0.360*	0.436**
NL	0.165	0.311*	0.027	0.058	0.050	0.282	0.073	0.193	1	0.811**	0.699**	$0.588^{**}$	0.489**	0.133	0.620**	0.557**
LDW	0.380*	0.646**	0.684**	0.289	0.317*	0.371*	0.346*	0.596**	0.246	1	0.869**	0.695**	0.613**	0.031	0.603**	0.626**
LA	0.247	0.581**	0.584**	0.212	0.247	0.361	0.170	0.487**	0.246	0.872**	1	0.652**	0.460**	-0.097	0.532**	0.416**
PR	0.196	0.022	0.034	0.117	0.077	-0.037	0.053	0.012	0.004	0.037	0.207	1	0.374*	0.112	0.648**	0.618**
APase	0.057	0.418**	0.248	0.108	0.055	-0.083	-0.125	0.053	0.048	0.400**	0.403**	0.213	1	-0.154	0.433**	0.397**
Stem P	-0.325	-0.258	-0.149	-0.292	-0.319	-0.315	-0.457**	-0.360	-0.130	-0.141	-0.146	-0.062	0.307	1	0.090	0.372*
ROOT P	-0.087	0.169	-0.102	0.028	-0.045	-0.109	-0.025	-0.158	-0.045	0.047	0.191	0.250	0.286	0.044	1	0.622**
Leaf P	-0.091	0.034	0.226	-0.171	-0.164	0.080	-0.079	0.042	-0.005	0.333*	0.431**	0.061	0.175	0.049	0.292	1

RL-Root length, RV- Root volume, RDW- Root dry weight, RA- Root area, RP-Root perimeter, NRT-Number of root tips, SL-Shoot length, SDW-Shoot dry weight, NL-Number of leaves, LDW- Leaf dry weight, LA- Leaf area, PR- Photosynthetic rate, APase- acid phosphatase, P-Phosphorus \* Correlation significant at 0.05 level \*\* Correlation significant at 0.01 level



**Fig 1:** Path coefficient showing direct effects of root length (RL), root volume (RV), root dry weight (RDW), root area (RA), root perimeter (RP), number of root tips (NRT), shoot length (SL), shoot dry weight (SDW), number of leaves (NL), leaf dry weight (LDW), leaf area (LA), photosynthetic rate (PR), acid phosphatase (APase) activity, stem and root phosphorus (P) on leaf P The single headed arrows indicate direct path correlation coefficient. (Data pooled for both P treatments)

Path analysis of leaf P showed that root dry weight had maximum significant positive direct effect (P<sub>3 16</sub> = 0.5612) followed by root volume (P<sub>2 16</sub> = 0.4638), leaf area (P<sub>11 16</sub> = 0.4332) on leaf P (Fig 1.). The direct effect of shoot length found to be minimum (P<sub>7 16</sub> = 0.0626). Root length (P<sub>1 16</sub> = -0.0529), root volume (P<sub>2 16</sub> = -0.4638), root area (P<sub>4 16</sub> = -0.2268), number of root tips (P<sub>6 16</sub> = -0.1998), shoot dry weight (P<sub>8 16</sub> = 0.5612), photosynthetic rate (P<sub>12 16</sub> = -0.1692) had negative effect on leaf P.

## Conclusion

Thus from the present study, it can be concluded that root perimeter was highly positively correlated with root area under +P condition. Stem P and root P, root length, root volume, root dry weight, shoot dry weight, number of leaves, root length and acid phosphatase activity had no correlation under –P condition. Path analysis of leaf P revealed that root dry weight had maximum significant positive direct effect followed by root volume, leaf area on phosphorus use efficiency.

# Acknowledgement

The first author would like to thank Department of Science and Technology, Ministry of Science and Technology, New Delhi, for the financial support to conduct this study through Women Scientist Scheme (A) programme.

## References

- 1. Bates TR, Lynch JP. Root hairs confer a competitive advantage under low P availability. Plant and Soil. 2001; 236:243-50.
- **2.** Clemens CM, Gerard C, Olga ES. Improving phosphorus use efficiency in agriculture: opportunities for breeding. Euphy. 2016; 207:1-22.
- 3. Cook BG, Pengelly BC, Brown SD, Donnelly JL, Eagles DA, Franco MA *et al* Tropical Forages: an interactive selection tool *Cajanus cajun* (Brisbane: CSIRO, DPI and F(Qld), CIAT, and ILRI, 2005.
- 4. Gutschick VP, Kay LE. Nutrient-limited growth rates: quantitative benefits of stress responses and some aspects of regulation. J Exp Bot.1995; 46:995-1009.

- 5. INDIASTAT 2015. http://www.indiastat.com
- Jain A, Poling MD, Karthikeyan AS, Blakeslee JJ, Peer WA, Titapiwatanakun B *et al* Differential effects of sucrose and auxin on localized phosphate deficiencyinduced modulation of different traits of root system architecture in *Arabidopsis*. Pl Physiol. 2007; 144:232-47.
- Krishnappa R, Hussain ISA. Phosphorus acquisition from deficient soil: involvement of organic acids and acid phosphatase in pigeonpea. Indian Journal Plant Physiology. 2014; 19(3):197-204. doi:10.1007/s40502-014-0101-z.
- 8. Mahalaya S, Aris TS, Alberth S, Luther K. dan Nakeus M. *Cultivation techniques and livestock production* (Canberra: International Potato Center (CIP) and Australian Centre for International Agricultural Research (ACIAR), 2015.
- Manschadi AM, Manske GGB, Vlek PLG. Root Architecture and Resource Acquisition – Wheat as a Model Plant. In: Eschel, A., Beeckman, T. (Eds.), Plant Roots: The Hidden Half., fourth ed. CRC Press, Taylor & Francis Group, USA. 2013; 22:1-22, 18.
- Rao IM, Kerridge PC, Macedo M. Nutritional requirements of Brachiaria and adaptation to acid soils. In Miles JW, Maass BL, DoValle CB (eds) Brachiaria: biology, agronomy and improvement. CIAT, Colombia, 1996, 53-71.
- 11. Shapley AN, Chapra SC, Wedepohl R, Sims JT, Daniel TC, Reddy KR. Managing agricultural phosphorus for protection of surface waters: issues and options. J Env Quality. 1994; 23:437-51.
- 12. Veneklaas EJ, Lambers H, Bragg J, Finnegan PM, Lovelock CE, Plaxton WC *et al* Opportunities for improving phosphorus-use efficiency in crop plants. New Phytol. 2012; 195:306-20.
- 13. Vesterager JM, Hogh-Jensen H, Nielsen NE. Variation in phosphorus uptake and use efficiencies between pigeonpea genotypes and cowpea. J Pl Nutri. 2006; 29(10):1869-88.
- 14. Wang X, Shen J, Liao H. Acquisition or utilization, which is more critical for enhancing phosphorus efficiency in modern crops? Pl Sci., 2010; 179:302-06.
- 15. Zhang D, Meng Q, Gai J, Yu D, Cui S, Cheng H *et al* Detection of quantitative trait loci for phosphorus deficiency tolerance at soybean seedling stage. Euphy. 2009; 167(3):313-22.