

# Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2018; 7(4): 2828-2832 Received: 11-05-2018 Accepted: 15-06-2018

Monika Nasrin Munni Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Shakera Islam Keya Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Rafiqul Hasan Khan Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Nura Ahmed Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Mahbuba Mohosina Runa Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Amina Ferdous Chowdhury Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Kazi Nuruddin Al Masud Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Correspondence Kazi Nuruddin Al Masud Department of Pharmacy, BRAC University, Dhaka, Bangladesh

# In-vitro investigation of antioxidant activity and phytochemical screening of Baccaurea ramiflora

Monika Nasrin Munni, Shakera Islam Keya, Rafiqul Hasan Khan, Nura Ahmed, Mahbuba Mohosina Runa, Amina Ferdous Chowdhury and Kazi Nuruddin Al Masud

#### Abstract

This study was conducted to evaluate the antioxidant property and perform the phytochemical screening of *Baccaurea ramiflora*. The plant belongs to the Euphorbiaceae family and locally used analgesic agent. This study provides a scientific basis for the use of *Baccaurea ramiflora* in traditional medicine. The seed was extracted using Ethanol. The extract was subjected to antioxidant activity determination and phytochemical screening. The phytochemical screening of *Baccaurea ramiflora* seeds showed the presence of alkaloids, flavonoids, glycosides, phenol, phlobatannin, resins, sterol and tannins but showed absence of tannin, resin and quinone. The total phenolic content of the ethanolic extract of the seeds of *Baccaurea ramiflora* was found to be 108.877 (mg of GAE / gm of extractives). Ethanolic extract solution presented notable free radical scavenging activity with an IC<sub>50</sub> value of 4.524μg/ml which is comparable with the value of standard Ascorbic Acid, which provided an IC<sub>50</sub> value of 3.01μg/ml.

Keywords: antioxidant, total phenolic content, DPPH, Ethanol, Baccaurea ramiflora

#### Introduction

Medicinal plants are an essential part of traditional medicines. The traditional drug is the collection of the knowledge, practices, and skills which are actually constructed on the theories, experiences and beliefs of indigenous to various cultures, whether explainable or not, used to keep good health as well as in the diagnosis, prevention, improvement or treatment of mental and physical illness. Traditional medicines use parts like leafs or roots to treat different diseases. Nature is the basic source of 87% of drugs used to treat all type of human diseases. About 25% of recommended drugs made from the plant. In developing countries, around 80% people depend on traditional based medications for their wellbeing [1]. In this project, I have focused on traditional plants used in Bangladesh to discover some unknown pharmacological effect of those plants like antioxidant, antimicrobial and cytotoxicity to discover a new source of drugs to treat diseases efficiently and cost effectively. Phytochemical investigations have been run on 15% medicinal plants [2]. Baccaurea ramiflora is one of the traditional plants that is used as an analgesic agent in Sylhet and anti-cancer agent and the juice of this herbs is used as painkiller and leaf paste is used as an anti-cancer drug. The other species of Baccaurea genus possesses several activities. Seeds of Baccaurea ramiflora (L). Are to possess antiemetic, purgative, stimulant properties and can be used to relieve colic pain [3]. Infusion of Baccaurea ramiflora (L.) is a good tonic and can be used in ulcerative colitis and remedy of Fever [4]. Baccaurea ramiflora contains flavonoids and chalcones which show powerful antioxidant effects and can be used to reverse oxidative stress-causing pathologic conditions, for example arteriosclerosis and cancer [5]. Based on the several activities provided by the other species of Baccaurea genus, Baccaurea ramiflora seeds has been chosen to perform the phytochemical screening and analysis of antioxidant property.

#### **Materials and Methods**

### Plant collection and identification

The seeds of *Baccaurea ramiflora* was collected in the month of October 2017 from Kumilla, Bangladesh. After that, its verification (Verification code number: 46964) was done by the National Herbarium of Bangladesh (NHB), Mirpur, Dhaka by submitting plant sample.

#### Chemicals

1, 1-diphenyl-2-picrylhydrazyl (DPPH), Potassium mercuric iodide, Iodine in potassium iodide, Dragendorff's reagent, Saturated picric acid, Tannic acid, Neutral ferric chloride, Sodium nitroprusside, Glacial acetic acid, Concentrated H<sub>2</sub>SO<sub>4</sub> Molish's reagents, CuSO<sub>4</sub>,

Alcoholic KOH, Basic lead acetate, Acetic anhydride, NaOH, Chloroform, NaNO<sub>2</sub>, Methanol, Folin-Ciocalteu reagent, Na<sub>2</sub>CO<sub>3</sub> were used.

# **Preparation of plant extract**

After the washing the leaves with clean water the leaves were shade dried for several days and were grounded finely as a granular particle with a high power grinding machine. About 400gm of grounded leave powder of *Baccaurea ramiflora* which was drenched in 2L of ethanol for 14 days period in a room temperature (22-25°C) with random agitation. After 14 days of soaking, the substances of the bottle were emptied out first to filter them by using Whatman filter paper (pore size 100nm). The filtrate was concentrated by using rotary evaporator (Heidolph) at 30°C temperature with a rotation speed of 100rpm up to form the concentrated ethanolic extract.

#### Phytochemical screening

The crude extract of *Baccaurea ramiflora* was used for phytochemical screening to identify its chemical compound present in its leaves.

# Procedure of extract preparation for screening

2-3 grams of dried ethanol extract was mixed with 50ml ethanol in a 100ml of conical flask. After that, that flask was labeled properly closing with cotton plugs and kept still for 1 to 2 hours. Later, the mixture was filtered through Whatman filter paper. Collected filtrates were used for phytochemical screening by following the standard process <sup>[6, 7]</sup>. The following qualitative tests were performed sequentially:

#### **Tests for Alkaloids**

**Mayer's test:** A few drops of Mayer's reagent (Potassium mercuric iodide solution) was added in 1ml of seed extract. If cream color precipitation form then it will contain the presence of alkaloids.

**Wagner's test:** In 1ml of seed extract was added with the same amount of Wagner's reagent (Iodine in potassium iodide). If reddish brown color precipitation form then it will point to the presence of alkaloids.

**Dragendorff's reagent test:** 2ml of Dragendorff's reagent was added in 1ml of seed extract and later dilute HCL of 2ml was added in that solution. If orange color precipitation forms then it will confirm the presence of alkaloids.

**Hager's test:** A few drops of Hager's reagent (Saturated picric acid solution) was added in 2ml of seed extract. If bright yellow color precipitation form then it will point to the presence of alkaloids.

**Tannic acid test:** A few drops of tannic acids was added in 1ml of seed extract. If yellow-brown colored precipitation form then it will point to the presence of alkaloids.

**FeCl<sub>3</sub> test:** About 1-2 ml extract was mixed with a little amount of neutral ferric chloride solution in dropwise. If cream yellow precipitation forms then it will point to the presence of alkaloids.

#### Tests for glycosides

**Legal's Test:** Addition of alkaline sodium nitroprusside and pyridine in extract solution results in the formation of cherry red color then it will confirm to the presence of glycosides.

**Keller Killiani test:** At first 1ml of glacial acetic acid was mixed-up with 1 ml of extract and cooled. After that 2-3 drops of ferric chloride was mixed and 2ml of concentrated H<sub>2</sub>SO<sub>4</sub> was added carefully in sideways of test tube walls. If reddish brown colored ring at the junction of two layers form then it will point to the presence of glycosides.

Concentrated H<sub>2</sub>SO<sub>4</sub> test: 1ml of Concentrated H<sub>2</sub>SO<sub>4</sub> was added in 1ml of seed extract and kept still for 2 minutes. If reddish color precipitate form then it will point to the presence of glycosides.

Molish's test: In seed extract around 2-3 drops of Molish's reagents was added. Later, a few drops of concentrated H<sub>2</sub>SO<sub>4</sub> was added properly. If reddish purple colored ring at the junction of two layers form then it will point to the presence of glycosides.

**Test for phlobatannins:** At first 2-3ml of 10% HCl was added in 10ml of seed extract in a boiling test tube which was boiled for 5-6 minutes. If red color precipitate occurs then it will point to the presence of phlobatannins.

**Test for resins:** 3-4ml of the CuSO<sub>4</sub> solution was mixed-up with seed extract which was shaken vigorously for 1-2 minutes and allowed to discrete. If green color precipitate occurs then it will point to the presence of resins.

**Test for quinones:** Alcoholic KOH solution was added in seed extract. If color ranging from red to blue occur then it will point to the presence of quinones.

**Test for Saponins:** In the test tube 5ml of the extract was taken and shaked vigorously to get a stable froth. 5-6 drops of olive oil were added into frothing solution. If the emulsion is formed then it will point to the presence of saponins.

# **Tests for phenols**

**Ellagic acid test:** A few drops of 5% (w/v) glacial acetic acid was added in seed extract. After that 5% (w/v) NaNO<sub>2</sub> solution was added. If muddy brown color form then it will point to the presence of phenols.

**Phenol test:** 1ml of the FeCl<sub>3</sub> solution was added in 2ml of seed extract. If the development of intense color form then it will point to the presence of phenols.

# **Tests for Tannins**

**Ferric chloride test:** A few drops of FeCl<sub>3</sub> was added in seed extract. If blackish color precipitate form then it will point to the presence of tannins.

**Lead acetate test:** A few drops of basic lead acetate was added in 1-2ml of seed extract. If bulky red color precipitate form then it will point to the presence of tannins.

**Alkaline reagent test:** A few drops of sodium hydroxide solution was added in seed extract. If red color form then it will point to the presence of tannins.

#### **Tests for flavonoids**

**Lead-acetate test:** A few drops of basic lead acetate solution was added in 1-2ml of seed extract. If reddish brown color precipitate form then it will point to the presence of flavonoids.

**FeCl<sub>3</sub> test:** A few drops of neutral ferric chloride solution was added in 1-2ml of seed extract. If the deposition of blackish red color precipitate form then it will point to the presence of flavonoids.

**Alkaline reagent test:** A few drops of sodium hydroxide was added in 1-2ml of seed extract. If yellowish red color occurs then it will point to the presence of flavonoids.

# Test for sterols

**Libermann-Burchard test:** A few drops of acetic anhydride solution was mixed with 1-2ml of seed extract. After that, a few drops of concentrated H<sub>2</sub>SO<sub>4</sub> was given beside the test tube walls in the mixture. If reddish brown color ring at the junction of two layers occur then it will point to the presence of sterols.

**Salkowski test:** 5ml of chloroform was added in 1-2ml of seed extract. After that, 1ml of concentrated H<sub>2</sub>SO<sub>4</sub> was put beside the test tube walls. If the reddish color in the lower layer occurs then it will point to the presence of sterols.

# Evaluation of antioxidant activity Determination of total phenolic content

Generally, the antioxidative action is shown by phenolics, phenolic acid, phenolic diterpenes, and flavonoids. Chemical properties of the phenolic compounds show that they exert their antioxidative properties by redox reaction [8]. Researches show that various amount of the phytochemicals retain antioxidant capacities which might be related to lower mortality rate and lower incidence human cancer [9]. Phenols get ionized in an alkaline condition which is why the Folin-Ciocalteu reagent is used which readily gets ionized in phenolic solution. Oxidized reagent turns blue from vellow. Color change intensity is measured as absorbance at 760 nm by UV spectrophotometer. Absorbance indicates the TPC (Total Phenolic Content) of particular test compound [10]. Total phenolic content of leaves of the Total phenolic content of leaves of Baccaurea ramiflora seed extract was measured by using the method which was designed [11] involving Folin-Ciocalteu reagent as an oxidizing agent and gallic acid as standard [12].

# **Determination of DPPH radical scavenging activity**

DPPH assay is simple and fast procedure to evaluate antioxidant activity of extract sample where its stability in the radical form is good <sup>[13]</sup>. Basic law of this assay is color change of DPPH solution from purple to yellow as the radical is quenched by antioxidant <sup>[14]</sup>. Based on the method described by Brand-Williams <sup>[15]</sup> the free radical scavenging activity or antioxidant property of plant extracts was

measured using DPPH (1, 1-diphenyl-2-picrylhydrazyl) reagent. Above mentioned procedure follows the addition of extract's ethanol solution (2 ml) with DPPH methanol solution (3 ml, conc.  $20\mu g/ml$ ). Decoloration of purple colored DPPH methanol solution by the test plant extract is compared with a standard ascorbic acid and BHT. DPPH was used to evaluate the free radical scavenging activity (antioxidant potential) of various compounds and medicinal plants [16, 17].

3ml of ethanolic solution of DPPH was mixed with 2ml of ethanolic sample (extract or control) solution at a various concentration which was ranging from 500 to 0.977µg/ml. The prepared solution was kept in dark for 30 minutes at room temperature for reaction. After that, prepared solution absorbance was measured against methanol as blank by UV spectrometer at 517nm.

Percent of inhibition of free radical DPPH (I%) was calculated by given equation:

$$(1\%) = (1 - A_{sample}/B_{blank}) \times 100$$

In the given equation,  $B_{lank}$  is denoted as absorbance of the control reaction (contain all chemical reagents except the test material).

The concentration of extract provided 50% inhibition ( $IC_{50}$ ) which was calculated by using the graph of inhibition percentage vs. extract concentration. In this graph we will use logartmic trendline to get an equation by which we will determine the IC<sub>50</sub> value for our extract sample.

General equation for calculating IC<sub>50</sub> is given below –

$$y = mln(x) + c$$

# Result and Discussion Phytochemical screening

Table 1: Phytochemical screening of Baccaurea ramiflora seed

Serial Number	Class of compound	Result
1	Alkaloid	+++++
2	Glycoside	+
3	Phlobatannin	+
4	Resin	-
5	Quinone	-
6	Phenol	++
7	Tannnin	-
8	Flavonoids	+
9	Sterol	++

**Note:** (+) = presence in a single method test, (++) = presence experimented in two methods, (+++) = presence experimented in three methods, (++++) = presence experimented in four methods, (+++++) = presence experimented in five methods and (-) = absence.

#### **Determination of total phenolic content**

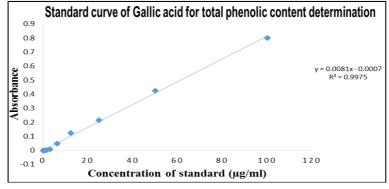


Fig 1: Standard curve of Gallic acid for total phenolic content determination

**Table 2:** Test samples for total phenolic content determination

Plant part	Sample code	Test Sample	Absorbance (Y)	Total phenolic content (mg of GAE / gm of extractives) (X)
Seeds of Baccaurea ramiflora	EE	Ethanolic extract	0.865	108.877

# DPPH free radical scavenging assay

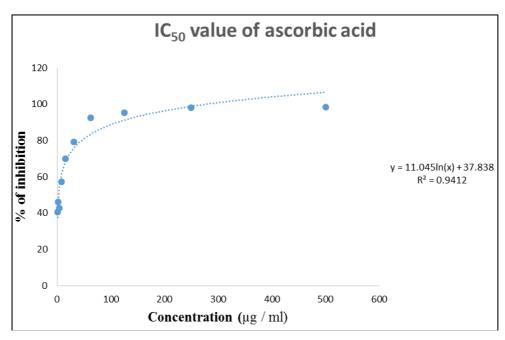


Fig 2: IC<sub>50</sub> value of ascorbic acid

Table 3: IC<sub>50</sub> value of Ethanolic extracts

Absorbance of the blank	Concentration (µg/ml) (X)	Absorbance of the extract	% Inhibition (Y)	IC <sub>50</sub>
0.325	500	0.021	93.54	
	250	0.028	91.38	
	125	0.042	87.08	4.524
	62.5	0.066	79.69	
	31.25	0.083	74.46	
	15.625	0.116	64.31	
	7.813	0.144	55.69	
	3.906	0.193	40.62	
	1.953	0.209	35.69	
	0.977	0.224	31.08	

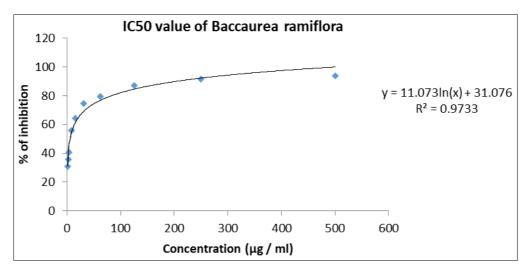


Fig 3: IC50 value of EE of seeds of Baccaurea ramiflora

# Discussion

The phytochemical screening of *Baccaurea ramiflora* showed the presence of alkaloids, flavonoids, glycosides, phenol,

phlobatannin, resins, sterol and tannins whereas showing the absence of tannin, resin and quinone.

The total phenolic content of the ethanolic extract of the seeds of *Baccaurea ramiflora* was found to be 108.877 (mg of GAE

/ gm of extractives). The Ethanolic extract of the seeds of *Baccaurea ramiflora* was tested to free radical scavenging activity by using the method suggested by Brand-Williams <sup>[15]</sup>. Reference standard was Ascorbic acid (ASA). Ethanolic extract solution presented the notable free radical scavenging activity with an  $IC_{50}$  value of  $4.524\mu g/ml$  which is comparable with the value of standard Ascorbic Acid, which provided an  $IC_{50}$  value of  $3.01\mu g/ml$ .

#### Conclusion

The phytochemical analysis showed the presence of several phytochemicals which can be further isolated using compound isolation. The antioxidant analysis showed *Baccaurea ramiflora* has higher level of antioxidant property and can be used as an antioxidative agent.

#### References

- Khatun A, Rahman M, Haque T, Rahman Md M, Akter M, Akter S *et al.* Cytotoxicity Potentials of Eleven Bangladeshi Medicinal Plants. The Scientific World Journal, 2014, 1-7. doi:http://dx.doi.org/10.1155/2014/913127.
- 2. Verpoorte R. Pharmacognosy in the new millennium: lead finding and biotechnology. J Pharm Pharmacol. 2000; 52:253-262.
- 3. Ahmed F, Rahman S, Ahmed N, Hossain M, Biswas A, Sarkar S *et al.* Evaluation of neolamarckia cadamba (roxb.) Bosser leaf extract on glucose tolerance in glucose-induced hyperglycemic mice. Afr J Tradit Complement Altern Med. 2011; 8(1):79-81. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/22238487
- 4. Keya MA, Rahman AHMM. Angiosperm Diversity at the Village Sabgram of Bogra, Bangladesh with Emphasis on Medicinal Plants. American Journal of Plant Biology, 2017; 2(1):25-34.
- Huq AKMM, Jamal JA, Stanslas J. Ethnobotanical, Phytochemical, Pharmacological, and Toxicological Aspects of *Persicaria hydropiper* (L.) Delarbre. PMC. US National Library of Medicine National Institutes of Health, 2014.
- Kokate CK. Practical Pharmacognosy. (4th Ed). New Delhi, India: Vallabh Prakashan Publication, 1999.
- Evans WC. Trease and Evans Pharmacognosy. (14th Ed.). Singapore: Harcourt Brace and company, Asia Pvt. Ltd 1997
- 8. Pietta A, Sionetti P, Mauri P. Antioxidant activity of selected medicinal plants. Agric Food Chem. 1998; 46:4487-4490.
- Velioglu YS, Mazza G, Gao YL, Oomah BD. Antioxidant activity and totalphenolics in selected fruits, vegetables and grain products. J Agric Food Chem. 1998; 46:4113-4117.
- 10. Harbertson JF, Spayd S. Measuring Phenolics in the Winery. American Journal of Enology and Viticulture. 2006; 57(3).
- 11. Skerget M, Kotnik P, Hadolin M, Hras A, Simonic M, Knez Z. Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food chemistry. 2005; 89:191-198.
- 12. Majhenik *et al.* Antioxidant and antimicrobial activity of guarana seed extracts, Food chemistry. 2007; 10:1016.
- 13. Bozin B, Mimica-Dukic N, Samojlik I, Goran A, Igic R. Phenolics as antioxidants in garlic (Allium sativum L., Alliaceae). Food Chem. 2008; 111:925-929.

- Karagözler AA, Erdag B, Emek YC, Uygum DA. Antioxidant activity and proline content of leaf extracts from Dorystoechas hastata. Food Chem. 2008; 111:400-407
- 15. Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. LWT Food Science and Technology. 1995; 28(1):25-30. https://doi.org/10.1016/S0023-6438(95)80008-5
- Choi SJ, Kim JK, Kim HK, Harris K, Kim C.-J, Park GG et al. 2,4-Di-tert-butylphenol from sweet potato protects against oxidative stress in PC12 cells and in mice. Journal of Medicinal Food. 2013; 16(11):977-83. https://doi.org/10.1089/jmf.2012.2739
- 17. Desmarchelier C, Mongelli E, Coussio J, Ciccia G. Evaluation of the *in vitro* antioxidant activity in extracts of *Uncaria tomentosa* (Willd.) DC. Phytotherapy Research. 1997; 11(3):254-256. https://doi.org/10.1002/(SICI)1099-1573(199705)11:3<254::AID-PTR76>3.0.CO;2-5
- Bondet V, Brand-Williams W, Berset C. Kinetics and Mechanisms of Antioxidant Activity using the DPPH. Free Radical Method. LWT - Food Science and Technology. 1997; 30(6):609-615. https://doi.org/10.1006/FSTL.1997.0240
- 19. Pandey G. Antioxidant and Antibacterial Activities of Leaf Extract of *Achyranthes aspera* Linn. (Prickly Chaff Flower). European Journal of Medicinal Plants. 2014; 4:695-708. https://doi.org/10.9734/EJMP/2014/8786.
- 20. Beaulah AG, Sadiq MA, Santhi JR. Antioxidant and Antibacterial activity of Achyranthes Aspera: An *in vitro* study. Der Pharma Chemica, 2011; 3(5):255-262. Retrieved from http://derpharmachemica.com/archive.html
- 21. Rishikesh. Phytochemical and Pharmacological Investigation of *Achyranthes Aspera* Linn. Scholars Academic Journal of Pharmacy. 2013; 2:74-80.
- 22. Priya CL, Kumar G, Loganathan K, Rao B. Antioxidant Activity of *Achyranthes Aspera* linn stem Extracts, Pharmacologyonline, 2010; 2:228-237.
- 23. Benariba N, Djaziri R, Bellakhdar W, Belkacem N, Kadiata M, Malaisse WJ *et al.* Phytochemical screening and free radical scavenging activity of Citrullus colocynthis seeds extracts. Asian Pacific Journal of Tropical Biomedicine. 2013; 3(1):35-40. http://doi.org/10.1016/S2221-1691(13)60020-9