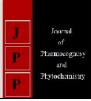


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# Management of stem rot of groundnut (Sclerotium rolfsii Sacc.) by plant oils

# DP Awasthi, NK Mishra and B Dasgupta

#### Abstract

Four types of plant oil viz., Palmarosa (*Cymbopogon martinii*), Karanja (*Pongamia pinnata*), Citronella (*Citronella* spp.) and Neem (*Azadirachta indica*) were tested against groundnut stem rot ((*Sclerotium rolfsii* Sacc) at 0.1 and 0.5 per cent concentration with twenty five different treatments (T) such as seed, soil and seed followed by soil treatment under *in situ* conditions excluding control, at BCKV, Nadia, West Bengal during two consecutive years i.e., 2003-04 and 2004-05. The observation was recorded for three parameters disease incidence, severity and yield. Among the treatments (T<sub>21</sub>) and (T<sub>22</sub>) (Seed + soil treatment with Citronella oil and Palma rosa oil @ 0.5%) showed economically significant positive response in all respective parameters compare to other however, the lowest variation was recorded in T<sub>25</sub> (control).

Keywords: groundnut, stem rot, management, plant oil

## Introduction

Groundnut (*Arachis hypogaea* L.) is one of the important economic oilseed crops of the world. It belongs to the family *leguminoseae*. Due to year round cultivation, groundnut crop is subjected to various diseases caused by fungi, bacteria, virus, phytoplasma, phytonematodes etc. Out of various diseases of groundnut, stem rot caused by *Sclerotium rolfsii* Sacc is a very important disease causing losses in different parts of India and throughout the world. In the first half of 20<sup>th</sup> Century, peanut production sustained losses of 10 to 20 million dollars annually due to this disease. Losses of 25 to 50% were recorded in 1938-1947. In India, this pathogen causes appreciable losses in chilli, bean, beet, carrot, cucurbit, potato, spinach betelvine, peanut etc. (Aycock, 1966)<sup>[1]</sup>

*Sclerotium rolfsii* Sacc. Teleomorph: *Athelia rolfsii* (Curzi) Tu and Kimbrough is a destructive fungal plant pathogen causing diseases in many monocotyledon and dicotyledon plants encompassing more than 500 host species (Punja, 1984)<sup>[8]</sup>. Despite continuous research over the past century since its first report on management of the pathogen has remained a challenge. Control efforts have often met with limited success, partially due to the extensive host range, prolific growth rate and ability of the pathogen to produce large numbers of sclerotia that may remain viable in soil for several years. Furthermore, control measures effective for a particular crop in an area may not be adaptable elsewhere due to regulatory or economic constraints. Spray of chemical fungicides possesses serious threat to beneficial organisms of eco-system. Since long, researchers are using extracts of botanicals to control plant disease. Plant oils are eco-friendly and less toxic to environment, thus use of plant oils for management of fungal and viral disease is becoming more and more popular and has became a very interesting area of research. In view of this, the present study was undertaken with the objective to manage collar rot of groundnut by use of different plant oils or botanicals.

## **Materials and Methods**

The experimental was carried out in two consecutive years i.e. 2003-04 and 2004-05 in Randomized Block Design having 25 treatments including a control and 3 replications at Jaguli Instructional Farm of BCKV, Nadia, West Bengal, India (Table 1). Four types of plant oils viz., Karanja (*Pongamia pinnata*), Citronella (*Citronella* spp.), Neem (*Azadirachta indica*) and Palmarosa (*Cymbopogon martinii*) were tested in two different concentrations of 0.1% and 0.5%. Three different types of method of application namely seed treatment, soil treatment and combination of seed treatment followed by soil treatment were used in the study. Before sowing, the seeds were soaked in respective oils in two different desired concentrations of 0.1% and 0.5% respectively. Control was maintained by soaking the seeds in sterile water. Percent disease incidence and percent disease index (PDI) was recorded at 90 DAS, respectively. Assessment of disease severity was done on 0-4 scale as given below:

Table 1: Efficacy of Plant oil used as seed and soil treatment against stem rot of groundnut during 2003-2005 (Pooled data of two years)

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Treatment	Disease Incidence (%)	% Disease Incidence over control	Disease Index (%)	% Disease Index over control	Yield (Kg/ha)
T <sub>1</sub> -Seed treatment with citronella oil @ 0.1%	39.0 (38.6)	16.4	24.5 (29.6)	47.1	383.2 (958)
T <sub>2</sub> -Seed treatment with palma rosa oil @ 0.1%	40.7 (39.6)	12.8	22.2 (28.1)	52.1	377.9 (944)
T <sub>3</sub> -Seed treatment with karanja oil @ 0.1%	42.8 (40.8)	8.3	35.0 (36.2)	24.5	333.0 (832)
T <sub>4</sub> -Seed treatment with neem oil @ 0.1%	37.9 (37.9)	18.8	43.3 (41.1)	6.6	402.1 (1005)
T <sub>5</sub> -Seed treatment with citronella oil @0.5%	32.6 (34.8)	30.1	32.6 (34.8)	29.7	419.6 (1049)
T <sub>6</sub> -Seed treatment with palma rosa oil @ 0.5%	26.7 (31.1)	42.8	41.0 (39.8)	11.6	441.2 (1103)
T <sub>7</sub> -Seed treatment with karanja oil @ 0.5%	38.7 (38.4)	17.1	23.0 (28.6)	50.43	390.6 (976)
T <sub>8</sub> -Seed treatment with neem oil @ 0.5%	35.7 (36.6)	23.5	26.2 (30.7)	43.5	410.8 (1027)
T <sub>9</sub> -Soil treatment with citronella oil @ 0.1%	28.3 (32.1)	39.4	34.4 (35.9)	25.8	429.5 (1073)
T <sub>10</sub> -Soil treatment with palma rosa oil @ 0.1%	27.2 (31.4)	41.7	33.4 (35.3)	28	438.1 (1095)
T <sub>11</sub> -Soil treatment with karanja oil @ 0.1%	29.9 (33.1)	26.3	22.9 (28.5)	50.6	412.4 (1031)
T <sub>12</sub> -Soil treatment with neem oil @ 0.1%	40.8 (39.6)	11.9	22.8 (28.5)	50.8	339.5 (848)
T <sub>13</sub> -Soil treatment with citronella oil @ 0.5%	26.1 (30.7)	44.1	20.2 (26.7)	56.4	443.3 (1108)
T <sub>14</sub> -Soil treatment with palma rosa oil @ 0.5%	25.7 (30.4)	44.9	17.7 (24.8)	61.8	449.2 (1123)
T <sub>15</sub> -Soil treatment with karanja oil @ 0.5%	27.8 (31.8)	39.1	23.6 (29.0)	49.1	428.2 (1070)
T <sub>16</sub> -Soil treatment with neem oil @ 0.5%	32.7 (34.8)	27.7	21.5 (27.6)	53.6	418.3 (1045)
$T_{17}$ -Seed + soil treatment with citronella oil @ 0.1%	29.9 (33.1)	35.8	31.5 (34.1)	32.1	420.6 (1051)
$T_{18}$ -Seed + soil treatment with palmarosa oil @ 0.1%	28.9 (32.5)	37.2	36.4 (37.1)	21.55	427.5 (1068)
$T_{19}$ -Seed + soil treatment with karanja oil @ 0.1%	25.0 (30.0)	44.7	26.0 (30.6)	43.9	444.5 (1111)
$T_{20}$ -Seed + soil treatment with neem oil @ 0.1%	30.5 (33.5)	34.6	16.9 (24.2)	63.5	420.2 (1050)
$T_{21}$ -Seed + soil treatment with citronella oil @ 0.5%	19.6 (26.2)	59.5	18.9 (25.7)	59.2	469.9 (1174)
T <sub>22</sub> -Seed + soil treatment with palmarosa oil @ 0.5%	19.4 (26.2)	58.8	23.0 (28.6)	50.4	464.6 (1161)
T <sub>23</sub> -Seed + soil treatment with karanja oil @ 0.5%	29.1 (32.6)	46.3	19.7 (26.3)	57.5	453.6 (1134)
$T_{24}$ -Seed + soil treatment with neem oil @ 0.5%	25.9 (30.5)	44.5	30.6 (33.5)	34	443.4 (1108)
T <sub>25</sub> - Control	46.7 (43.1)	0	46.4 (42.9)	0	235.9 (589)
CD at 0.05%	3.26		1.82		40.59
*Figures in the parenthesis are angular transformed					

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Scale	Disease severity		
0	Healthy plants		
1	Yellowing		
2	Browning and white mycelial growth near the basal part of stem		
3	Wilting		
4	Complete collapse of the plant		

Percent Disease Index (PDI) was calculated using the following formulae:

Sum of all numerical ratings

PDI =

Number of plants observed x maximum rating used

# **Results and Discussion**

Results of the experiment revealed that 90 days after sowing among the treatments  $T_{21}$  and  $T_{22}$  (Seed + soil treatment with Citronella oil and Palma rosa oil @ 0.5%) showed economically significant positive response with lowest variation in disease incidence (19.6 & 19.4%), severity (18.9 & 23%) and highest yield (469.9 & 464.6 kg/ha) respectively in all parameters. An another treatment  $T_{20}$  (Seed + soil treatment with neem oil @ 0.1%) is found to be one of the best treatment compare to other and lowest variation recorded in T<sub>25</sub> (control). Similar results were also finding out by Singh et al. (1989) <sup>[10]</sup>, Banerjee et al. (1989) <sup>[2]</sup>, Madhavi et al. (2011)<sup>[7]</sup> and Guerra et al. (2015)<sup>[3]</sup> against S. rolfsii by same plant oil in both in vitro and in vivo conditions. Although many findings of such type of research work references are available in respect to control of S. rolfsii or other plant pathogens by use of essential oils and plant extract Thakur et al. (1989) <sup>[12]</sup>, Handique and Singh (1990) <sup>[4]</sup>, Khanna and Johari (1991)<sup>[5]</sup>, Kole et al. (1993)<sup>[6]</sup> but not much work is known in regards of their effect as seed soaking prior to sowing. Thus it can be concluded that plant oils derived from Cymbopogon martinii, Citronella spp., Pongamia pinnata and Azadirachta indica as seed and soil treatment can prevent stem rot of groundnut caused by S. rolfsii.

# References

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- 1. Aycock R. Stem rot and diseases caused by *Sclerotium rolfsii*. NC Agric. Exp. Stn. Tech. Bull. 1966; 174:202
- 2. Banerjee S, Bhattacharya I, Mukherjee N. Sensitivity of three sclerotial rice pathogens to plant oils. International Rice Research Newsletter. 1989; 14:6-23.
- 3. Guerra YL, Oliveira TAS, Laranjeira D, Lima LM, Filo PAM, Santos RC. Control of *Sclerotium rolfsii* in peanut by using *Cymbopogon martini* essential oil. African Journal of Microbiology Research. 2015; 9(27):1684-1691.
- 4. Handique AK, Singh HB. Antifungal action of lemongrass oil on some soil borne plant pathogens. Indian Perfumer. 1990 34(3):232-234.
- 5. Khanna RK, Johri JK. Screening of alternative biocides amongst plant based essential oils. National Academy Science Letters. 1991; 14(1):3-6.
- 6. Kole C, Pattnaik S, Subramanyam VR, Narain A. Antifungal efficacy of oils and its genetic variability in citronella. Crop Research Hisar. 1993; 6(3):509-512.
- Madhavi GB, Bh attiprolu SL, Reddy VB. J Hortl. Sci. 2011; 6(2):156-158.
- 8. Punja ZK, Jenkins SF. Light and scanning electron microscopic observation of calcium oxalate crystals produced during growth of *Sclerotium rolfsii* in culture and in infected tissue. Can. J Bot. 1984; 62:2028-32.
- 9. Sesha Kiran K, Lingaraju S, Adiver SS. Effect of Plant extract on *S. rolfsii* the incitant of stem rot of groundnut. J Mycol. Pl. Pathol. 2006; 36(1):77-79.
- Singh RK, Shukla RP, Dwivedi RS. Studies of fungitoxicity of oils against *Sclerotium rolfsii* Sacc. And soil myco flora. National Academy Science Letters. 1989; 12:6:183-85.

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- 11. Srinivas T, Manjulatha M, Venkateswarlu D. Biointensive management of collar rot, *Aspergillus niger* and Stem rot, *Sclerotium rolfsii* Sacc in groundnut. Journal of Oilseed Research. 2005; 22(1):103-104.
- 12. Thakur RN, Singh P, Khosla MK. *In- vitro* studies on antifungal activities of some aromatic oils. Indian Perfumer. 1989; 33(4):257-260.