

E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2018; 7(3): 2371-2375 Received: 18-03-2018 Accepted: 24-04-2018

Auta Richard

Department of Biochemistry, Faculty of Science, Kaduna State University, Nigeria

Waziri M Peter

Department of Biochemistry, Faculty of Science, Kaduna State University, Nigeria

Efe M Omwirhiren

Chemistry Department, Federal College of Education, Zaria, Nigeria

Mercy Richard

Department of Environmental Management, Kaduna State University, Nigeria

Charles Luke Kumai

Department of Biochemistry, Faculty of Science, Kaduna State University, Nigeria

Correspondence Auta Richard Department of Biochemistry, Faculty of Science, Kaduna State University, Nigeria

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



Journal of Pharmacognosy and

Phytochemistry

Auta Richard, Waziri M Peter, Efe M Omwirhiren, Mercy Richard and Charles Luke Kumai

Abstract

African trypanosomiasis is a major disease of economic and public health importance caused by Trypanosoma brucei brucei that affects humans and livestock. The use of plant extract is fast becoming the choice method for the treatment of the disease. As a result, the current study investigated the *in vitro* and in vivo activity of methanolic seed extract of Garcinia kola on Trypanosoma brucei brucei. The in vitro assay was carried out by treating the parasites with 10, 5, 2.5, 1.3, 0.6 and 0.3 mg/ml of the extract. For the in vivo studies, the methanolic extract was administered orally at a dose of 200, 400 and 600 mg/kg body weight of rats 2 days post infection for 4 consecutive days. Parasitaemia and mean survival time were used as indices for monitoring the efficacy of the extracts. The phytochemical screening revealed the presence of alkaloids, flavonoids, terpene, quinines, saponins and tannins, with steroid having the highest concentration (31.13mg/100g) and quinone with the lowest concentration (0.08mg/100g). The in vitro screening showed antitrypanosomal activity at higher concentrations of the extract (5 and 10mg/ml) after the 60 minutes incubation period. Althoughin the in vivo study, the extract did not significantly decrease (p>0.05) parasitaemia of the infected rats, we presume this may have occurred due to inefficient absorption of the extract after oral administration. However, this study reveals that the extract has potential anti-trypanosomal activity in vitro and can be used to design drugs to reduce the global scourge of trypanosomiasis.

Keywords: antitrypanosomal, Garcinia kola, Trypanosoma brucei brucei, parasitemia.

Introduction

Trypanosomiasis, also known as sleeping sickness is an infectious disease of African origin that affects humans and animals. It is caused by the parasites *Trypanosoma species* and transmitted by the bite of infected tsetse flies (Abenga, 2014)^[1]. *Trypanosoma congolense, vivax* and *brucei brucei* are the responsible for the disease in animals while the subspecies of *Trypanosoma brucei, Trypanosoma brucei gambiense* and *brucei rhodiense* are responsible for the disease in humans (Ogbole *et al.*, 2016)^[30]. Trypanosomes kill more than 3 million cattle annually and those animals that survive display low productivity due to the wasting effects of the disease (Hurse, 2001)^[16]. In addition, the devastating effects of the disease affects the lives and livelihood of many rural farmers of sub-Saharan Africa that cannot afford good and quality health care services (Johnson and Omoniwa, 2014)^[20].

Clinically, the disease is treated with melarsoprol, suramin, pentadimine and efflornithine that are very expensive and fast becoming impotent because of the emergence of resistant strains of the Trypanosomes. Current study is focused on the use of natural products from plants that could help to overcome the challenges associated with the conventional drugs used for the treatment of trypanosomiasis.

Many natural products of plant origin have been reported to have activities against different species of protozoan parasites including Plasmodium, Trypanosoma, leishmania and Entamoeba (Hoet *et al.*, 2004) ^[15]. Garcinia kola (G. kola) a species of flowering plant of the Clusiaceae or Guttiferae family found in the tropical rain forest region of West Africa is a plant known for its medicinal properties. The presence of bioactive compounds with high therapeutic properties in the seeds of the plant has been reported (Tona *et al.*, 1999; Okunji *et al.*, 2000; Farombi *et al.*, 2002; Farombi, 2003; Omwirhiren *et al.*, 2017) ^[41, 12, 14, 32]. Phytochemical compounds that have been isolated from G. kola include oleoresin (Onayade *et al.*, 1998) ^[33], tannins, saponins, alkaloids, cardiac glycosides (Ebana *et al.*, 1991) ^[7]. Two new

chromanols, garcioic acid and garcinal, together with σ tocotrienol were also reported to be isolated from G. kola (Terashima *et al.*, 2002)^[40]. Other phytochemical compounds so far isolated from G. kola seeds are biflavonoids such as kolaflavone and 2hydroxybi-flavonols (Okunji et al., 2000; Terashima et al., 2002) ^[40]. Studies have shown that kolaviron, a natural biflavonoid from G. kola seeds possess the ability to protect against oxidation of lipoprotein in rats (Farombi and Nweokeafor, 2005)^[13]. This activity which was demonstrated to be presumably by Fe2+ chelation and antioxidant activity might be of immense benefit in the management of African trypanosomiasis. It has been suggested that removal of excess iron through chelation could possibly prevent iron mediated injury to cells thereby reducing the pathology of anaemia and tissue damage associated with African trypanosomiasis (Ekanem et al., 2009; Johnson and Omoniwa, 2014)^[23, 20]. Several medicinal plants are currently being investigated for their anti-trypanosomal properties in a bid to identify active components that can be used to combat the scourge of trypanosomiasis. Garcinia kola (Heckel) is one of the medicinal plants that is purported to have anti-trypanosomal effect. The plant is also reported to be useful for the treatment of diabetes, cough, laryngitis, infectious diseases, erectile problems, liver and lipid disorders among others (Iwu et al., 1990; Adegboye et al., 2008; Johnson et al., 2011)^[18, 2, 19]. However, the active components of the plant and their mechanism of action is still unknown. Therefore, the present study seeks to identify the phytochemicals of G. kola seed and also investigate their in vitro and in vivo anti-trypanosomal activity.

Materials and Method

Collection of plant material and parasites: *Garcinia kola* seeds, purchased at Kaduna Central Market, Kaduna State, Nigeria, was authenticated at the Department of Biological science, Kaduna State University, Nigeria, by Mr. U. S. Gallah and assigned the Voucher No, 9501.

The parasite, *Trypanosoma brucei*, (strain *Trypanosoma brucei* brucei) was obtained from the Nigerian Institute for Trypanosomiasis Research Kaduna, Nigeria and was maintained via routine inoculation of normal albino rats.

Preparation of methanolic extract of *G. kola* seed: The seeds were peeled off and cut into smaller pieces, shade-dried and then ground into fine powder using mortar and pestle. About 80 g of the pulverized sample was soaked in 800 mL of methanol (analytical grade) for two (2) days at 25°C. The extract was filtered with a clean muslin cloth and subsequently evaporated using a water bath at 40°C. The jelly-like concentrates obtained was weighed and placed in sterilized sample bottle for storage in a refrigerator at 4°C.

Phytochemical screening: The phytochemicals screening was carried out according the following methods: The total flavonoids content was estimated using the procedure described by Zhishen *et al (1999)*. Tannins content of the sample was estimated by the method of Siddhuraj and Manian (2007). Estimation of total saponins content was determined by the method described by Makkar *et al.*(2003). Steroids and terpenes were estimated according to the method described by Ejikeme *et al.* (2010).

Experimental animals: Eight (8) weeks old albino rats with average weight of 98.54 g obtained from the Animal house of Nigerian Institute for Trypanosomiasis Research (NITR),

Kaduna were used for the study. They were housed in plastic cages with wood shavings as beddings and maintained on a commercial poultry feed (Fitzer, Nigeria), with access to clean water. They were acclimatized for two (2) weeks before commencement of the experiment.

Experimental design: Fifteen (15) albino rats were weighed and grouped randomly into five experimental groups that contained 3 rats each, for the *in vivo* study. The groups are as follows: Group 1 – Normalrats treated with 2% DMSO (normal control); Group 2– Trypanosome-infected rats treated with 2% DMSO (negative control); Group 3– Trypanosome-infected rats treated with the extract at a dose of 200 mg/kg body weight of rat; Group 4– Trypanosome-infected rats treated with the extract at a dose of 400 mg/kg body weight of rat; Group 5– Trypanosome-infected rats treated with the extract at a dose of 600 mg/kg body weight of rat. The treatment was done for 4 days consecutively.

Infection of animals: Blood from a highly parasitized mouse was obtained by cardiac puncture using a syringe and needle and transferred into an EDTA- coated sample bottle. This was diluted appropriately with normal saline to serve as inoculum. Healthy rats were infected intraperitoneally with 0.3 mL each of the inoculum that contains $about10^6$ trypanosomes/mL (Ene *et al.*, 2009)^[9].

Determination of parasitaemia and antitrypanosomal activity of extract: Blood was collected from the tail of rats before and after treatment with the extract of G. kola and the level of parasitaemia was monitored in vitro and in vivo as described by Ene et al. (2014) [10]. The level of parasitaemia was expressed as log of the absolute number of parasites per mL of blood. Briefly, the *in vitro* anti-trypanosomal assay was performed in a 96 well plate seeded with the parasites. The parasites were treated with the extract and standard drug, veridium (positive control) and incubated at 37 °C for 60 min. The negative control rats were treated with PBS-G. The motility of parasites was used as a basis for assessing the antitrypanosomal activity of the treatments. For the in vivo study, parasitaemia was monitored every day until the last mortality was observed. The negative control group was treated with 2%DMSO.

Statistical analysis: The data obtained from the study were expressed as mean \pm standard deviation of mean (SD). Data analysis was performed using Statistical Package for Social Science (SPSS), version 20.0 (Inc. Chicago IL, USA). One way ANOVA was performed to determine statistical significance difference at 95% confidence interval.

Result and Discussion

 Table 1: Quantitative Photochemical Components of seed extract of Garcinia kola

Phytochemicals	Concentration (mg/100g)
Alkaloid	2.30 ± 0.05
Flavonoid	2.05 ± 0.03
Terpenes	3.05±0.03
Tannines	0.35 ± 0.03
Saponins	2.47±0.04
Quinones	0.08 ± 0.00
Anthraquinones	1.12 ± 0.02
Steroids	31.13±1.00

Data in duplicate: mean \pm S.D

Table 2: In vitro anti-trypanosomal activity of metholic extract of	G
kola against trypanosome brucei brucei	

Concentration	Trypanosome count	%
(mg/mL)	$(Mean \pm SEM)$	mortality
10	0.00 ± 0.00^{a}	100
5	0.00 ± 0.00^{a}	100
2.5	6.00 ± 1.15^{b}	74.3
1.25	5.33±0.33 ^b	71.2
0.625	8.33±0.67°	64.3
0.3125	8.67±0.66°	62.8
Positive control	0.00 ± 0.00^{a}	100
Negative control	23.3±1.45 ^d	0.00

Superscript: a, b, c and d indicates statistical differences

 Table 3: Effect of methanol seed extract of Garcinia kola on parasitaemia of Trypanosoma brucei brucei infected rats.

Crowns	Days post inoculation				
Groups	2	3	4	5	6
1	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
2	0.00 ± 0.00	2.67±1.15	38.7±24.1	98.0±42.4	-
3	0.13±0.23	4.00 ± 2.00	78.0 ± 55.7	171.3±73.3	-
4	0.67±1.15	4.22 ± 5.05	102.0±133	96.0±45.3	-
5	0.00 ± 0.00	2.72 ± 2.21	25.3±11.5	104.0±34.9	-

Values are given as mean \pm standard deviation (mean \pm SD)

 Table 4: Percentage (%) mortality of rats infected with

 Trypanosoma brucei brucei treated with various doses of methanolic

 seed extract of Garcinia kola

Cround	% mortality				MST (down)		
Groups	Day 2	Day 3	Day 4	Day 5	Day 6	$\mathbf{W}_{1,5,1}$ (days)	
1	0	0	0	0	0	>14 ^a	
2	0	0	0	33.3	100	5.7±0.6 ^b	
3	0	0	0	0	100	6.0±0.0 ^b	
4	0	0	0	33.3	100	5.7±0.6 ^b	
5	0	0	0	0	100	6.0 ± 0.0^{b}	

M.S.T= mean survival time, subscript: a and b indicates statistical differences. In each column, mean values with the same superscripts have no statistical significant difference (p > 0.05).

Despite the enormity of the health and economic implication of African trypanosomiasis, current chemotherapeutic options are very limited and very expensive (Legros et al., 2002). Therefore, there is a need for more potent and cheaper alternatives that could be used to manage the debilitating disease. Some plants have been reported to be very potent in the treatment of trypanosomiasis (Asuzu and Chineme, 1990; Nok, 2002; Mergia et al., 2015; Nwodo et al. 2015) [5, 27, 25, 28]. The present study investigated in vitro and in vivo antitrypanosomal activity of the methanolic extract of the seeds of Garcinia kola. The phytochemical screening of the methanolic seed extract of G. kola reveals the presence of alkaloids, saponins, flavonoid, terpene, tannine, steroids. anthraquinone and quinone (Table 1). From the steroid screened, phytochemicals had the highest concentration (31.13mg/100g) while quinone has the lowest concentration (0.08mg/100g). This finding is in agreement with previous study by Adesuyi et al. (2012) [3] that reported that G. kola seed contained 0.342, 2.471, 0.645 and 2.041% of tannins, saponins, alkaloids and flavonoids respectively. Findings from present study indicate the the presence of these phytochemicals may have contributed to the trypanocidal activity observed in the current study.

Numerous *in vitro* and *in vivo* studies conducted on the antitrypanosomal activities of the class of compounds listed

above reported the potential of each class of compounds in killing or inhibiting the growth of wide ranges of trypanosomes (Yabu et al., 2013; Johnson and Omoniwa, 2014)^[45, 20]. It has been well established that flavonoids and flavonoid-derived plant natural products are effective antioxidant (Nwodo et al., 2015)^[28]. Antioxidants have been shown to protect cells against the damaging effects of reactive oxygen species, such as singlet oxygen, superoxide, peroxyl radicals, hydroxyl radicals and peroxynitrite (Pieta, 2000). An imbalance between antioxidants and reactive oxygen species results in oxidative stress, leading to cellular damage. Cellular damage and consequently anaemia which is characterized by a significant reduction in PCV is one of the major pathologic effects of trypanosome infection (Ekanem et al, 2009; Johnson et al., 2011; Johnson and Omoniwa, 2014)^[8, 23, 19, 20]. The antioxidant property of flavonoids in G. kola promises to be of great benefit in the fight against this debilitating disease. Furthermore, the trypanocidal activity of a number of plants is attributed to the highly aromatic flavonoids they contain. Earlier, we observed the presence of flavonoid in the methanolic extract of G. kola which we presume that it contributes to the bioactivity of the extract (Johnson et al.,2011; Ogbadoyi et al.,2011) ^[19, 29]. Although in vitro activity of crude extract may need further verification, it remains one of the best tools widely used in bioassay guided identification of active components in plants (Wurochekke et al., 2014)^[43].

In the *in vitro* assay, the trypanocidal activity of the extract and standard drug (veridium) increased significantly (p < 0.05)as the concentration of extract was increased (Table 2). Similarly, the mortality increased as the concentration of the extract increased. The extract effectively inhibited the motility of Trypanosoma brucei brucei and at a concentration of 5 and 10 mg/mL of the extract, a 100% mortality was observed in vitro (Table 2). Therefore, the observed in vitro antitrypanosomal activity of Garcinia kola might be attributed to either the individual class of compounds present in the extract, or to the synergistic effect that each class of compounds exert to give the observed biological activity (Mergia et al., 2014)^[24]. Natural products can generate free radicals that causes peroxidative and DNA damage (Atawodi et al., 2003) ^[6]. The extract of G. kola may have exerted its trypanocidal activity through peroxidative and DNA damage and this corroborates previous studies (Ogbadoyi et al., 2011; Johnson and Omoniwa, 2014)^[29, 20]. Earlier reports has shown that The antitrypanosomal property of alkaloids has been suggested to be due to DNA intercalation in combination with protein biosynthesis inhibition while the trypanocidal activity exhibited by the saponin fraction may be as a result of cytotoxicity. Saponins are natural glycosides which possess a wide range of pharmacological properties including cytotoxic activity. Their cytotoxic effects have been suggested to be due to either apoptosis inducement or non-apoptotic cell death stimulation. Cell death may be as a result of mechanisms like - stimulation of autophagic cell death, decrease in NO production in cells, or cytoskeleton integrity disassembly (Merschjohann et al., 2001; Podolak et al., 2010; Mann et *al.*,)^[26, 37]. The doses administered to rats did not significantly decrease (p>0.05) the level parasitemia (Tables 3 and 4). Even though the in vitro screening shows the extract has trypanocidal activity, the in vivo studies does not confirm this fact. This we believe may be due to insufficient absorption of the extract after administration. Impaired absorption of drugs is known to affect the efficacy and therapeutic effect of drugs (Mann et al., 2009)^[23]. Therefore, it is very imperative to

Journal of Pharmacognosy and Phytochemistry

improve absorption of the extract and also understand the factors that affect the potency of the extract. It is also possible that the active component may need some form of activation as observed in many clinically used drugs. This study may actually lead to an increased potency of the seed extract of *Garcinia kola*.

However, our study has shown that the extract has potential anti-trypanosomal activity and this finding corroborates previous reports and justifies the traditional use of *Garcinia kola* in the treatment of trypanosomiasis.

Conclusion

Trypanosomiasis is a disease of humans and livestock that affects the livelihood of sub-Saharan Africans in rural settlements (Fairlamb, 1982)^[11]. The search for a potent drug that could help reduce the scourge of African trypanosomiasis remains elusive. Natural products are fast becoming the choice method for the treatment of the disease because they are safer and cheaper. The current study reveals that the extract of G. kola seed contains components that can effectively destroy the parasite and reduce mortalities associated with African trypanosomiasis. This findings no doubt is indeed an encouragement for the development of present and future African chemotherapy that promises succor to a region that has suffered from the debilitating effects of trypanosomiasis and consequently improve the quality of life. Further collaborative study in this area intend to focus on the isolation and spectroscopic characterization of the bioactive ingredients in G. kola which may serve as novel compounds in the quest for the development of new, affordable and more effective antitryponocidal therapies.

Acknowledgment

We are thankful to Mr. Emmanuel Enoh for assisting with the statistical analysis of the data.

References

- 1. Abenga JN. A comparative pathology of *Trypanosoma* brucei infections. Global Advanced Research Journal of Medicine and Medical Science (GARJMMS). 2014; 3(12):390-399. Available online http://garj.org/garjmms/index.htm
- Adegboye MF, Akinpelu DA, Okoh AI. The bioactive and phytochemical properties of *Garcinia kola* (Heckel) seed extract on some pathogens. *African Journal of Biotechnology*. 2008; 7(21):3934-3938.
- 3. Adesuyi AO, Elumm IK, Adaramola FB, Nwokocha AGM. Nutritional and Phytochemical Screening of *Garcinia kola. Advance Journal of Food Science and Technology*. 2012; 4(1):9-14.
- 4. Anosa VO. Haematological and Biochemical changes in human and animal trypanosomiasis. Parts I and II. *Rev. Elev.Med. Vet. Pays Trop.* 14 65-78, 151-164 Vet. Parasitol. 1988; 115(2):125-145
- Asuzu IU, Chineme CN. Effects of Morinda lucida leaf extracts on Trypanosoma brucei brucei infection. J Ethnoparmacol. 1990; 30:307-313.
- 6. Atawodi SE, Bulus T, Ibrahim S. *In vitro* trypanocidal effect of Methanol extract of some Nigerian Savannah Plants. Afri J. Biotechnol. 2003; 2(9):317-321.
- Ebana RU, Madunagu BE, Ekpe ED, Otung IN. Microbiological exploitation of cardiac glycosides and alkaloids from Garcinia kola, Borreria ocymoides, Kola nitida and Citrus auratifolia. J. Appl. Bacteriol. 1991; 71(5):398-401.

- Ekanem JT, Johnson TO, Balogun EA. Serum Iron and Nitric Oxide Production in Trypanosoma brucei Infected Rats Treated with Tetracycline. BIOKEMISTRI. 2009; 21(1):41-51.
- 9. Ene AC, Atawodi SE, Ameh DA, Nnamani CN, Apeh YEO. Antitrypanosomal Effects of Petroleum Ether, Chloroform and Methanol Extracts of *Artemisia maciverae* Linn. Indian Journal of Experimental Biology. 2009; 47:981-986.
- Ene AC, Edeh NG, Bonny-Okoli C, Ojiako OA, Ujowundu CO. Igwe CU..*In vitro* and *in vivo* Antitrypanosomal Effects of Methanol and Aqueous Extracts of *Picrali manitida*. British Journal of Pharmaceutical Research. 2014; 4(5):644-653. SCIENCEDOMAIN international www.sciencedomain. org
- Fairlamb A. Biochemistry of trypanosomiasis and rational approaches to chemotherapy. Trends Biochem. Sci. 1982; 23-26.
- Farombi EO, Akanni OO, Emerole GO. Antioxidant and scavenging activities of flavonoid extract (kolaviron) of Garcinia kola seeds *in vitro*. Pharm. Biol. 2002; 40(2):107-116.
- Farombi EO, Nwaokeafor IA. Antioxidant Mechanisms of Kolaviron: Studies on Serum Lipoprotein oxidation, Metal Chelation and Oxidative Membrane Damage. Clin. Exp. Pharmacol. Physiol. 2005; 33(8):667-674.
- 14. Farombi EO. African indigenous plants with chemotherapeutic potentials and biotechnological approach to the production of bioactive prophylactic agents. African J Biotech. 2003; 2:662-671.
- Hoet S, Opperdoes FR, Brun R, Quetin-Leclercq J. Natural products active against African trypanosomes: a step towards new drugs. Nat Prod Rep. 2004; 21(3):353-364.
- 16. Hursey BS. The programme against African trypanosomiasis-aims, objectives and achivements. Trends Parasitol. 2001; 17:2-3.
- Igweh AC, Onabanjo AO. Chemotherapautic effects of Annona senegalensis in Trypanosoma brucei brucei. Ann. Trop. Med. Parasitol. 1989; 83:527-534
- Iwu MM, Igboko OA, Okunji CO, Tempesta MS. Antidiabetic and aldose reductase activities of flavones of Garginia Kola. J pharm. Pharmacol. 1990; 42:290-292
- 19. Johnson TO, Ijeoma KO, Ekanem EE, Nelson E, Mohammed B. *In vitro* Studies on the Trypanocidal Activities of various Phytochemical fractions obtained from Garcinia.kola seed. Journal of Medicine in the Tropics. 2011; 13(2):124-128.
- 20. Johnson TO, Omoniwa BP. *In vivo* Trypanocidal Activity of Ethanolic Crude Extract and Phytochemical Fractions of Garcinia kola Seeds Annual Research & Review in Biology. 2014; 4(1):212-222
- 21. Legros D, Ollivier G, Gastellu-Etchegorry M, Paquet C, Burri C. Treatment of human African trypanosomiasis-present situation and needs for research and development. Lancet Infect Dis. 2002; 2:437-440.
- 22. Mann A, Ogbadoyi EO. Evaluation of Medicinal Plants from Nupeland for Their *in vivo* Antitrypanosomal Activity. Am. J Biochem. 2012; 2(1):1-6
- 23. Mann A, Egwim EC, Banji B, Abdulkadir N, Gbate M, Ekanem JT. Efficacy of Dissotisrotundiolia on Trypanosomabrucei brucei infectious rats. *African Journal of Biochemistry Research*. 2009; 3:5-8

- 24. Mergia E, Shibeshi W, Terefe G, Teklehaymanot T. Evaluation of *In vivo* Antitrypanosomal Activity of Aqueous and Methanol Leaf Extracts of *Clutia abyssinica* (Euphorbiaceae) against *Trypanosoma congolense* Field Isolate. Journal of Natural Product Chemistry and Research. 2014; 2:4.
- 25. Mergia E, Terefe G, Teklehaymanot T, Shibeshi W. Phytochemical screening and *in vitro* antitrypanosomal of aqueous and methanol leave extract of *Clutia abyssinica* (Euphorbiceae) against T. congolense. Pharmacologia, 2015; 6(3):79-87.
- Merschjohann K, Sporer F, Steverding D, Wink M. In vitro Effect of Alkaloids on Bloodstream forms of Trypanosoma brucei and T. congolense. Planta Med. 2001; 67:623-627.
- 27. Nok AJ. Azaanthraqinone inhibits respiration and in-vitro growth of long slender blood stream forms of *T. congolense*. Cell Biochem. Funct. 2002; 20:205-212.
- Nwodo NJ, Ibezim A, Ntie-Kang F, Adikwu MU, Mbah C. Anti-Trypanosomal Activity of Nigerian Plants and Their Constituents Molecules. 2015; 20:7750-7771; doi:10.3390
- 29. Ogbadoyi EO, Kabiru AY, Omotosho RF. Preliminary Studies of the Antitrypanosomal Activity of Garcinia kola nut Extract in Mice Infected with Trypanosoma brucei brucei, Journal of Medicine and Medical Sciences. 2011; 2(1):628-631.
- Ogbole E, Dashak DA, Nvau JB, Daben MR, Abongaby G, Obaloto OB et al. Phytochemical screening and *in vitro* evaluation of the antitrypanosomal action of the methanolic leaf extract of Corymbiatorelliana. International Journal of Ethnomedicine and Pharmacognosis. 2016; 3(1):20-29.
- Okunji CO, Ware TA, Hicks RP, Iwu MM, Skanchy DJ. Capillary electrophoresis determination of biflavanones from Garcinia kola in three traditional African medicinal formulations. Planta Med. 2002; 68:440-444.
- 32. Omwirhiren EM, Asefon OA, James SA. The phytochemical constituents and relative antimicrobial activities against clinical pathogens of different seed extracts of Cola nitida (Vent.), Cola acuminata (Beauvoir) and Garcinia kola (Heckel) grown in South West, Nigeria. Journal of Pharmacognosy and Phytochemistry. 2017; 6(1):493-501.
- Onayade OA, Looman AMG, Scheffer JJC, Gbile ZO. Lavender lactone and other volatile constituents of the oleoresin from seeds of Garcinia kola Hechel. Flavour Frangrance J. 1998; 13(6):409-412.
- Onyeyili RA, Egwu GO. Chemotherapy of Africa trypanomiasis: A historical review. Protozool Abstr. 1995; 5:229-243.
- 35. Pepin J, Meda HA. The epidemiology and control of human African trypanosomiasis Advances in Parasitol. 2001; 49:71-132.
- Pietta PG. Flavonoids as antioxidants. J Nat. Prod. 2000; 63:1035-1042.
- Podolak I, Galanty A, Sobolewska D. Saponins as cytotoxic agents: a review. Phytochem Rev. 2010; 9(3):425-474.
- 38. Solomon CI, Arukwe UI, Onuoha I. Preliminary phytochemical screening of different solvent extracts of stem bark and roots of *Dennetiatripetela* G. Baker. *Asian* Journal of Plant Science and Research. 2013; 3(3):10-13. Available online at: www.pelagiaresearchlibrary.com.

- 39. Terashima K, Kondo Y, Aqil M, Waziri M. A study of biflavanones from the stem of Garcinia kola. Heterocycles. 1990; 50:238-290.
- 40. Terashima K, Takaya Y, Niwa M. Powerful antioxidative agents based on garcinoic acid from Garcinia kola. Bioorgan. Med. Chem. 2002; 10(5):1619-1625.
- 41. Tona L, Ngimbi NP, Tsakala M, Mesia K, Cimanga K, Apers S. Antimalarial activity of 20 crude extracts from nine African medicinal plants used in Kinshasa, Congo. J. Ethnopharmacol. 1999; 68:193-203.
- 42. Welburn SC, Coleman PG, Fevre E, Mandling I. Sleeping Sickness, a tale of two Diseases. Trends in parasitology. 2001; 17:19-24.
- 43. Wurochekke AU, Nuhu N, Anyanwu GO. Trypanocidal potential of Carrisaedulis in male wistar rats infected with *T. congolense*. American Journal of Research Communication. 2014; 2(1)
- 44. Wurochekke AU, Chechet G, Nok AJ. In vitro and in vivo antitrypanosomal activity of the leaf of Lawsonia inermis against Trypanosoma bruceibruce infection in mice. J Medical. Sci. 2004; 4:236-239.
- 45. Yabu Y, Yoshida A, Suzuki T, Niher C, Kawai K. The efficacy of ascofuranone in a consecutive treatment on *Trypanosoma brucei brucei* in mice. Parasitol Int. 2003; 52:155-164.